Reversible retinal vessel closure from VEGF-induced leukocyte plugging

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Clinical trials in patients with macular edema due to diabetic retinopathy or retinal vein occlusion (RVO) have shown that suppression of VEGF not only improves macular edema, but also reopens closed retinal vessels, prevents progression of vessel closure, and improves retinopathy. In this study, we show the molecular basis for those clinical observations. Increased retinal levels of VEGF in mice cause plugging of retinal vessels with leukocytes, vessel closure, and hypoxia. Suppression of VEGF reduces leukocyte plugging, causing reperfusion of closed vessels. Activation of VEGFR1 contributes to leukocyte recruitment, because it is significantly reduced by an anti-VEGFR1-neutralizing antibody. High VEGF increases transcriptional activity of NF-κB and expression of NF-κB target genes, particularly Vcam1. Injection of an anti-VCAM-1-neutralizing antibody reduces VEGF-induced leukocyte plugging. These data explain the broad range of benefits obtained by VEGF suppression in patients with ischemic retinopathies, provide an important insight into the pathogenesis of RVO and diabetic retinopathy, and suggest that sustained suppression of VEGF early in the course of these diseases may prevent vessel closure, worsening ischemia, and disease progression. This study also identifies VEGFR1 and VCAM-1 as molecular targets whose suppression could supplement VEGF neutralization for treatment of RVO and diabetic retinopathy.

Introduction

Diabetic retinopathy is a major complication of diabetes that is a highly prevalent cause of moderate and severe vision loss (1). The Diabetes Control and Complication Trial and the United Kingdom Prospective Diabetes Study demonstrated that hyperglycemia is a key driver of diabetic retinopathy because tight control of blood glucose provides substantial benefit (2, 3). Glucose enters the retina primarily through facilitative transport, and therefore, high blood glucose leads to high glucose in the retina, which, after many years, leads to the classic signs of diabetic retinopathy: retinal hemorrhages, microaneurysms, and cotton wool spots (microinfarcts). Patients are usually asymptomatic when these first signs of diabetic retinopathy appear, and it may be several years before decreased vision occurs from progression to more advanced stages of diabetic retinopathy, diabetic macular edema (DME), or proliferative diabetic retinopathy (PDR) (4). There are many hypotheses with supporting data as to how high retinal glucose leads to vascular damage and ultimately to DME and/or PDR, including production of advanced glycation end products (5), oxidative stress (6–8), increased cytokine expression and inflammation (9), or neuronal damage and neurovascular crosstalk (10), but conclusive proof for any of the competing hypotheses has been elusive. The long latent period between the onset of hyperglycemia and clinical signs of diabetic retinopathy and the variable and often long period between the onset of signs and the development of DME or PDR have prevented development of an animal model that closely mimics diabetic retinopathy (11); this has made it difficult to determine whether the hyperglycemia-induced abnormalities listed above contribute to the development of late vascular changes seen in human diabetic retinas.

The most consequential vascular change in diabetic retinopathy is retinal vessel closure resulting in areas of retinal nonperfusion and ischemia, because nonperfusion correlates with diabetic retinopathy severity and the development of PDR (12, 13). Retinal nonperfusion also occurs in other retinal diseases, and, along with diabetic retinopathy, they are classified as ischemic retinopathies. The second most preva-
Intraocular injection of VEGF causes transient leukostasis. Twenty-four hours after intravitreous injection of 1 μg VEGF165 (VEGF) or PBS in one eye, adult C57BL/6 mice were perfused through the left ventricle with rhodamine-conjugated concanavalin A (Con A). Compared with PBS-injected eyes in which fluorescence microscopy of retinal flat mounts showed narrow vessels that rarely contained leukocytes (Figure 1A), VEGF-injected eyes showed dilated retinal vessels, a well-establish effect of VEGF due to stimulation of eNOS (37–39), containing clumps of leukocytes adherent to the endothelium (Figure 1B). There was a marked decrease in leukocytes within retinal vessels at 48 hours after VEGF injection, but, 72 hours after injection, many leukocytes were again seen within large retinal vessels (Figure 1, C and D). At 96 hours after VEGF injection, there were few leukocytes in retinal vessels. Perfusion with fluorescein-labeled dextran 24 hours after injection showed the normal appearance of retinal vessels in PBS-injected eyes (Figure 1E), but eyes injected with VEGF had dilated vessels, with many leukocytes seen in negative relief (Figure 1F, arrowheads). Compared with fluorescein angiograms in PBS-injected eyes (Figure 1G), those done 24 hours after injection of 1 μg VEGF had dilated vessels, which combined with leakage and staining of vessel walls caused small vessels to stand out (Figure 1H). No perfusion defects were seen. Quantification of the mean

Results

Intraocular injection of VEGF causes transient leukostasis. Twenty-four hours after intravitreous injection of 1 μg VEGF165 (VEGF) or PBS in one eye, adult C57BL/6 mice were perfused through the left ventricle with rhodamine-conjugated concanavalin A (Con A). Compared with PBS-injected eyes in which fluorescence microscopy of retinal flat mounts showed narrow vessels that rarely contained leukocytes (Figure 1A), VEGF-injected eyes showed dilated retinal vessels, a well-establish effect of VEGF due to stimulation of eNOS (37–39), containing clumps of leukocytes adherent to the endothelium (Figure 1B). There was a marked decrease in leukocytes within retinal vessels at 48 hours after VEGF injection, but, 72 hours after injection, many leukocytes were again seen within large retinal vessels (Figure 1, C and D). At 96 hours after VEGF injection, there were few leukocytes in retinal vessels. Perfusion with fluorescein-labeled dextran 24 hours after injection showed the normal appearance of retinal vessels in PBS-injected eyes (Figure 1E), but eyes injected with VEGF had dilated vessels, with many leukocytes seen in negative relief (Figure 1F, arrowheads). Compared with fluorescein angiograms in PBS-injected eyes (Figure 1G), those done 24 hours after injection of 1 μg VEGF had dilated vessels, which combined with leakage and staining of vessel walls caused small vessels to stand out (Figure 1H). No perfusion defects were seen. Quantification of the mean
Figure 1. Intravitreous injection of VEGF causes transient severe leukostasis in retinal vessels. Retinal vessels in mice perfused with rhodamine-labeled concanavalin A (con A) were narrow and most did not contain any leukocytes 24 hours after intravitreous injection of PBS (A) but were dilated and packed with leukocytes 24 hours (B) or 72 hours (C) after intravitreous injection of 1 μg VEGF. The mean (± SEM) number of intravascular leukocytes per retina (PBS-treated mice, n = 5; VEGF-treated mice, n = 6 at each time point) was significantly greater in VEGF-injected eyes compared with PBS-injected eyes at 24 and 72 hours after injection (*P < 0.001 by unpaired t tests) but not at 48 or 96 hours (D, P = 0.6179 [48 hours], P = 0.4722 [96 hours] by unpaired t tests). Retinal vessels in a region around the optic nerve (ON) of mice perfused with fluorescein-labeled dextran showed normal retinal vasculature 24 hours after intravitreous injection of PBS (E), whereas vessels were dilated and packed with leukocytes seen in negative relief 24 hours after injection of 1 μg VEGF (F, arrowheads). Fluorescein angiography 24 hours after injection of PBS (G) or 1 μg VEGF (H) showed no identifiable nonperfusion. (I) The mean (± SEM) number of intravascular leukocytes per retina (NI group n = 6, mice treated with 200 ng, 500 ng, 1,000 ng VEGF n = 6; mice treated with 0 ng, 50 ng, 100 ng VEGF n = 5) was determined for several doses of VEGF and was significantly greater than PBS control for doses ≥100 ng (*P ≤ 0.002, †P < 0.001; P = 0.002 [100 ng, 24 hours], P = 0.001 [200 ng, 24 hours], P < 0.001 [500 ng, 24 hours], P < 0.001 [1,000 ng, 24 hours]).
number of intravascular leukocytes per retina after injection of various doses of VEGF showed a significant increase compared with PBS-injected eyes at 24 hours after injection of doses ≥100 ng (Figure 1I). There were also significantly more leukocytes in retinal vessels 72 hours after injection of 1 μg VEGF but not any of the lower doses. Compared with eyes injected with 1 μg VEGF, those injected with lower doses appeared to have fewer leukocytes in large vessels and more leukocytes adherent in smaller vessels (Figure 1J).

**Sustained increased expression of VEGF causes sustained leukostasis, reduced perfusion, and retinal hypoxia.** Intravitreal injection of VEGF results in a sudden increase and then a fairly rapid decrease in retinal VEGF, which differs from the situation in eyes with ischemic retinopathy, in which there is sustained expression of VEGF in ischemic retina. Mice with doxycycline-inducible expression of VEGF in photoreceptors (Tet/opsin/VEGF mice) have sustained expression of VEGF during treatment with doxycycline (40). One (Figure 2A) and two days (Figure 2B) after initiation of 2 mg/ml doxycycline in drinking water, many more leukocytes were adherent to the walls of small retinal vessels compared with those in PBS-injected eyes (Figure 1A) or uninjected eyes. Three days after the onset of doxycycline treatment, leukocytes were seen in vessels of all sizes, with large aggregates in large vessels (Figure 2, C and D). The mean number of intravascular leukocytes per retina was significantly greater 1, 2, and 3 days after starting doxycycline, compared with retinas of control mice not treated with doxycycline, and was significantly greater on day 3 compared with days 1 and 2 (Figure 2E). Fluorescein angiography 3 days after starting doxycycline showed dilated large retinal vessels radiating from the optic nerve. Between the large vessels, the network of small vessels was somewhat blurred by extravascular fluorescein leakage, punctuated by regions of hypofluorescence (Figure 2F, box). Magnification of the boxed area in Figure 2F shows patches of hypofluorescence, with sharp borders appearing cut out of the diffuse continuous fluorescence caused by retinal capillaries (Figure 2G, asterisks). This is very similar in appearance to sharply cut out black areas on human fluorescein angiograms recognized as capillary closure (32). Retinas from Con A–perfused doxycycline-treated Tet/opsin/VEGF mice that were also stained with pimonidazole, a stain for hypoxic tissue, showed regions of retinal hypoxia adjacent to vessels containing leukocytes (Figure 2H). Retinas from Tet/opsin/VEGF mice that were not treated with doxycycline showed no pimonidazole staining (Supplemental Figure 1, A, C, and E; supplemental material available online with this article; https://doi.org/10.1172/jci.insight.95530DS1). A low-magnification image of a retina from a Con A–perfused doxycycline-treated Tet/opsin/VEGF mouse (Supplemental Figure 1, B, D, and F), in which leukocytes in retinal vessels are small but still discernible (Supplemental Figure 1B, box, and Supplemental Figure 1F, arrows), provides more perspective and shows that the hypoxia is not uniform throughout the retina, but rather occurs in patches.

**Turning off VEGF expression or injection of a VEGF-neutralizing protein reduces leukostasis and opens closed vessels.** On day 3 after intraperitoneal injections of doxycycline on days 0, 1, and 2, retinal flat mounts from rhodamine-labeled Con A–perfused Tet/opsin/VEGF mice showed leukocytes adherent to the walls of retinal vessels (Figure 3A) that appeared to plug the lumen of some vessels (Figure 3B). On day 17, 2 weeks after the third injection of doxycycline, few intravascular leukocytes were seen (Figure 3C). Quantification of the number of intravascular leukocytes showed significantly more on day 3 when VEGF expression was high, compared with day 17 when VEGF levels had been normal for at least a week, indicating that most of the leukocytes had cleared (Figure 3D). Fluorescein angiograms obtained on day 3 showed dilation of large retinal vessels and severe extravascular leakage, resulting in diffuse hyperfluorescence filling spaces between small vessels; despite the increased overall fluorescence as a result of extravascular leakage, dark areas of retinal nonperfusion were seen (Figure 3E, boxes). A repeat fluorescein angiogram in the same Tet/opsin/VEGF mouse on day 17 showed that the large retinal vessels were no longer dilated, leakage had resolved allowing better resolution of small vessels; the previously black areas were filled in with a continuous network of vessels, indicating reopening of previously closed vessels (Figure 3F). Magnified images of the large boxed areas in Figure 3, E and F, allow better comparison of retinal vessels exposed to high levels of VEGF (Figure 3G) and the same vessels after reduction in VEGF levels (Figure 3H). High VEGF levels resulted in dilated large vessels with black patches between them indicating regions of vessel closure (Figure 3G, asterisks), while after levels of VEGF returned to normal, the same region of retina showed well-perfused vessels in the area where there previously were no vessels, indicating reopening of previously closed vessels.
Figure 2. Sustained expression of VEGF in the retina causes leukostasis, retinal vessel closure, and retinal hypoxia. Tet/opsin/VEGF double-transgenic mice with doxycycline-inducible expression of VEGF in the retina were given 2 mg/ml doxycycline in drinking water and perfused with rhodamine-labeled Con A 1 day (A), 2 days (B), or 3 days (C and D) after initiating doxycycline. Leukocytes were present in small vessels 1 day (A) and 2 days (B) after onset of VEGF expression and were present in vessels of all sizes after 3 days (C and D). The mean (± SEM) number of intravascular leukocytes per retina was significantly greater at each of the 3 time points compared with control (E) (*P < 0.001 by 1-way ANOVA with Bonferroni corrections) and at day 3 was greater than other time points (†P < 0.01, 1-way ANOVA with Bonferroni corrections). (F) Fluorescein angiography of Tet/opsin/VEGF mice 3 days after starting doxycycline showed dilated large retinal vessels radiating from the optic nerve (ON), between which the networks of small vessels were slightly blurred by extravascular leakage. There were hypofluorescent areas with sharp borders that appeared cut out of the diffuse fluorescence (box), better seen in a magnified view of the boxed area (G, asterisks). The dark black areas indicate absence of capillaries due to nonperfusion. Staining of a retinal flat mount with hydroxyprobe 3 days after starting doxycycline showed hypoxic retina (red) adjacent to vessels containing adherent FITC–Con A–stained leukocytes (H, arrowheads). Scale bar: 50 μm.
in 8-month-old normal control mice (Figure 4H). Fluorescein angiography in 7-month-old and 12-month-old rho/VEGF mice showed several areas of retinal nonperfusion (Figure 4I and Figure 4J, respectively, asterisks). In order to assess the effect of aflibercept, 7-month-old rho/VEGF mice had baseline fluorescein angiography on day 0 and then had subcutaneous injections of 25 mg/kg aflibercept on days 0, 2, 4, and 6 and repeat fluorescein angiography on day 7. There were many areas of retinal nonperfusion on the baseline fluorescein angiogram (Figure 5A) that were improved after treatment with aflibercept (Figure 5B). Even at low magnification, this was seen for the regions in the large and small boxes in which black regions surround asterisks in Figure 5A; the black areas are filled in with green, indicating reopening of vessels. Magnified images of the large boxed areas in Figure 5, A and B, demonstrate black regions devoid of vessels (Figure 5C, asterisks) that subsequently have perfused vessels (Figure 5D) but also show black areas (Figure 5C, dots), which remain black, indicating little or no reperfusion in those areas.
Stimulation of VEGFR1 contributes to leukocyte recruitment. VEGFR1 has been demonstrated on monocytes, macrophages, and neutrophils and mediates VEGF-stimulated leukocyte migration (42, 43). To determine if VEGFR1 on leukocytes contributes to VEGF-induced retinal leukostasis, mice were given an intravenous injection of 100 μg rat anti-mouse VEGFR1 or rat IgG and an intracocular injection of 200 ng VEGF, and, after 24 hours, they were perfused with FITC-labeled Con A. Compared with control IgG-injected mice (Figure 6A), there were many more intravascular leukocytes and they often appeared to occlude the lumen of vessels exemplified in 3 separate 7-month-old mice by plugging at a vessel bifurcation (C), by a line of leukocytes (D), or packing of a substantial length of a vessel lumen with a large aggregate (E). Similarly, there were leukocytes plugging vessels particularly at branch points in the retinas of 12-month-old (F) and 15-month-old (G) rho/VEGF mice. The mean (± SEM) number of intravascular leukocytes per retina in both 20-day-old and 30-day-old rho/VEGF mice, but not 16-day-old mice, was significantly greater than that in N-5W control mice \( (*) P < 0.001 \) compared to 5 weeks by unpaired \( t \) tests) and mean intravascular leukocytes were significantly greater in 7-month-old, 12-month-old, and 15-month-old rho/VEGF mice compared with N-8M control mice \( (**) P < 0.001 \) by unpaired \( t \) tests). Retinal flat mounts from fluorescein-dextran-perfused 7-month-old (I) and 12-month-old (J) rho/VEGF mice show abruptly ending vessels and large areas of nonperfusion (asterisks). Scale bar: 50 μm (A–D and G); 100 μm (E and F).
VEGF stimulates expression of mediators of leukocyte adhesion. The transcription factor NF-κB increases expression of many proteins involved in leukocyte–endothelial cell adhesion. To determine if VEGF activates NF-κB in retinal vascular endothelial cells, primary human retinal endothelial cell cultures (HRECs) were incubated in 25 ng/ml VEGF for 0, 1, or 2 hours, and NF-κB transcriptional activity was measured. There was a large increase in NF-κB transcriptional activity in HRECs exposed to VEGF for 1 hour that was decreased but still significantly higher than that of controls at 2 hours (Figure 7A). Expression of the NF-κB target gene Vcam1 was dramatically increased in HRECs after exposure to VEGF for 4 hours (Figure 7B). The constitutive level of mRNA for ICAM-1 was quite high in HRECs, and exposure to VEGF caused a modest, statistically significant increase (Figure 7C). Exposure of HRECs to VEGF for 4 hours also caused a significant increase in mRNA for E selectin (Figure 7D) but not in mRNA for ICAM-2, P selectin, or vascular adhesion protein (VAP; Supplemental Figure 2, A–C). There was little immunostaining for VCAM-1 in HRECs that were not exposed to VEGF (Figure 7, E and G), but those exposed to VEGF for 4 hours showed strong staining for VCAM-1 (Figure 7, F and H).

We sought to determine if the effects of VEGF on cultured endothelial cells predicted effects on retinal endothelial cells in vivo. In wild-type C57BL/6 mice, there was a significant increase in mRNA for VCAM-1 in the retina 24 hours after intravitreous injection of 1 μg VEGF as well as increases in mRNA for integrins β1 and αv, which have been implicated in leukocyte adhesion to vascular endothelium, and an increase in mRNA for ICAM-2 48 hours after injection (Figure 8A); however, mRNA for ICAM-1 was not significantly increased between 12 and 48 hours after injection (Supplemental Figure 2D). Sustained expression of VEGF in doxycycline-treated Tet/opsin/VEGF mice resulted in significant increases in mRNA for VCAM-1, ICAM-2, integrin β1, and integrin αv, which were first seen 3 days after onset of VEGF expression (Figure 8B), rather than after 1–2 days, as occurred after intravitreous injection of a large bolus of VEGF (Figure 8A). There was no increase in mRNA for ICAM-1, E selectin, P selectin, or VAP in the retinas of doxycycline-treated Tet/opsin/VEGF mice (Supplemental Figure 2, E–H).

Since the most dramatic and consistent VEGF-induced changes occurred in the mRNA for VCAM-1 and we also demonstrated an increase in VCAM-1 protein in cultured HRECs treated with VEGF, we investigated the effect of VEGF on VCAM-1 protein in retinal vessels in vivo. Twenty-four hours after intravitreous injection of PBS, there was little VCAM-1 in retinal vessels detectable by immunostaining (Figure 8, C–E), but 24 hours after injection of 1 μg VEGF, there was strong staining for VCAM-1 in retinal vessels near the surface of the retina (Figure 8, F–H). Two approaches were used to assess the effect of VEGF on NF-κB transcriptional activity in vivo. First, a nonviral gene transfer approach that provides good transduction of retinal vascular endothelial cells (44) was used to transduce retinal cells with a NF-κB reporter construct. Compared with intravitreous injection of PBS, injection of VEGF in mice with retinal transduction of the reporter construct caused significantly greater NF-κB transcriptional activity, as measured by dual-luciferase reporter assay (Figure 8I).
Second, we investigated the effect of doxycycline-induced expression of VEGF in the retinas of Tet/opsin/VEGF mice on nuclear localization of NF-κB. Compared with nuclear fractions from retinal homogenates of untreated Tet/opsin/VEGF mice, those from mice treated with doxycycline for 1, 2, or 3 days showed higher levels of NF-κB but equivalent levels of histone 3 (Figure 8J).

To determine the effect of blocking VCAM-1 on VEGF-induced retinal leukostasis Tet/opsin/VEGF mice were given an intravenous injection of 100 µg rat anti-mouse VCAM-1 or rat IgG at the onset of doxycycline treatment. Three days after doxycycline treatment, many intravascular leukocytes in the retinal vessels of rat IgG-treated mice (Figure 8K) and significantly fewer in those of anti-VCAM-1–treated mice (Figure 8, L and M).

Discussion
The treatment of DME, macular edema due to RVO, and neovascular age-related macular degeneration (AMD) has been revolutionized by the development of specific antagonists of VEGF. In addition, there has been an unexpected bonus, because neutralization of VEGF in the eyes of patients with ischemic retinopathies has shown that increased expression of VEGF in the retina does more than stimulate NV and excessive vascular leakage leading to macular edema, it also causes retinal hemorrhages, microaneurysms, and closure of retinal vessels (45). In this study, we have identified the mechanism by which high levels of VEGF cause retinal vessel closure; VEGF recruits leukocytes into retinal vessels and increases their adhesion to retinal vascular endothelial cells, resulting in vessel plugging and reduced perfusion. Activation of VEGFR1 on leukocytes contributes to their recruitment, because recruitment is reduced by systemic administration of an anti-VEGFR1 antibody. High levels of VEGF increase the transcriptional activity of NF-κB, which is a stimulator of expression of VCAM-1, other adhesion molecules, and cytokines that may further increase leukocyte recruitment and leukostasis. Neutralization of VCAM-1 with an anti-VCAM-1 antibody reduces VEGF-induced leukostasis.

These data provide important insights into the pathogenesis of RVO. Thrombosis of the central retinal vein or one of its branches disrupts venous return from the entire retina for the former and 25%–50% of the retina for the latter. Perfusion is compromised in the affected retina, resulting in retinal ischemia and
increased expression of VEGF. In some patients, particularly those that are older, there is immediate closure of some retinal capillaries and/or arterioles, probably due to preexistent arterial disease. Regardless of the amount of retinal vessel closure that occurs immediately after thrombosis, it is frequently followed by additional closure of retinal vessels that can be reversed by VEGF suppression (32–34). In untreated eyes with CRVO, the worsening of retinal nonperfusion can be so profound that it leads to severe loss of vision due to widespread ischemic death of retinal cells, including those in the macula and/or neovascular glaucoma, a severe complication that often leads to loss of the eye. This severe worsening of retinal nonperfusion was previously attributed to conversion from a nonischemic CRVO to an ischemic CRVO due to worsening of the vein occlusion, sometimes described as conversion of a partial CRVO to a complete CRVO (46, 47). However, measurement of area of retinal nonperfusion at several time points in RVO patients participating in clinical trials showed that progression of nonperfusion was the rule rather than the exception and that it did not occur abruptly, but rather progressed continuously through at least 24 months, the last measurement time point (30, 31). In these and other clinical trials (32–34), it has been noted that subjects with substantial nonperfusion at baseline showed more rapid and often more severe worsening of nonperfusion than those with little or no nonperfusion at baseline. This suggests that all CRVOs are ischemic but to different degrees, resulting in different levels of VEGF expression, and it is eyes at the most severe end of the ischemia spectrum at baseline that tend to show the most rapid and extensive progression of retinal nonperfusion, which...
Figure 8. VEGF stimulates expression of VCAM-1 in the retina, and neutralization of VCAM-1 reduces VEGF-induced leukostasis. (A) C57BL/6 mice were given an intravitreous injection of PBS or 1 μg VEGF, and qRT-PCR was used to measure retinal mRNA levels for several adhesion molecules. Compared with PBS controls, there was significant elevation of the mean (± SEM) number of mRNA transcripts per 10^5 Cyclophilin A transcripts for VCAM-1, ICAM-2, integrin β1, and integrin α4 (n in parentheses along x axis, *P < 0.05 by unpaired t tests). (B) In Tet/opsin/VEGF mice, the mean (± SEM) number of mRNA transcripts per 10^5 Cyclophilin A transcripts for VCAM-1, ICAM-2, integrin β1, and integrin α4 first became significantly elevated after 3 days of doxycycline treatment compared with controls not treated with doxycycline (n = 5 *P < 0.05 by unpaired t tests). (C–H) One day after C57BL/6 mice were given an intravitreous injection of 1 μg VEGF in one eye and PBS in the fellow eye, ocular frozen sections immunostained for VCAM-1 (red) and Griffonia simplicifolia lectin (green) showed no detectable staining for VCAM-1 in PBS-injected eyes (C–E), but eyes that had been injected with VEGF showed several vessels that stained for VCAM-1 (F–H). (I) Fourteen days after subretinal injection of 0.5 μg NF-κB luciferase reporter vector and 0.25 ng Renilla luciferase plasmid in C57BL/6 mice, an intravitreous injection of 1 μg VEGF was performed. Five days later, the ratio of Firefly/Renilla luciferase was measured in retinal/choroidal homogenates by Dual-Luciferase Reporter Assay. Lines show that the mean relative luciferase activity (n = 6) was significantly greater in VEGF-injected eyes compared with PBS-injected eyes (*P = 0.0138 by unpaired t test). (J) Immunoblots of nuclear fractions from Tet/opsin/VEGF mice treated with doxycycline for 1 (D1), 2 (D2), or 3 days (D3) showed higher levels of NF-κB but equivalent histone 3 in nuclei from retinas with high levels of VEGF compared with those in untreated Tet/opsin/VEGF mice with low levels of VEGF (Ctrl). (K–M) Tet/opsin/VEGF mice (n = 10 for each group) were given an intravenous injection of 100 μg anti-mouse VCAM-1 antibody or rat IgG; after 3 days of doxycycline treatment, there were many adherent leukocytes in retinal vessels of IgG-injected mice (K) and significantly fewer in eyes of anti-VCAM-1–injected mice (L and M) (*P < 0.0013 by unpaired t test). Scale bar: 100 μm.
occurs steadily, rather than as a sudden conversion. Since suppression of VEGF stops progression of non-perfusion and even causes reperfusion of some closed vessels, a causal role of high levels of VEGF has been established (32). The current study demonstrates the mechanism; high levels of VEGF in the retina stimulate recruitment and adhesion of leukocytes, resulting in plugging of large and small retinal vessels, and turning off VEGF expression or intraocular injection of a VEGF-neutralizing protein reduces leukostasis and allows closed vessels to reopen.

In early background diabetic retinopathy, there is not the sudden onset of retinal ischemia and high levels of VEGF that is seen in RVO, but there is evidence of small increases in retinal VEGF in experimental models of diabetes (48). The situation in RVO is mimicked by injecting a large bolus of VEGF into the vitreous or by doxycycline-induced expression of high levels of VEGF in Tet/opsin mice. In these models, there is severe leukostasis, resulting in plugging of large as well as small vessels over a relatively short time frame, as occurs in RVO. The situation in diabetic retinopathy is mimicked by rho/VEGF mice, in which there is a relatively small increase in VEGF, mild leukostasis that gradually increases, and, after several months, closure of retinal vessels occurs. Slowly progressing closure of retinal vessels is the underlying cause of progression of diabetic retinopathy (49), leading to substantial areas of ischemic retina, which further increases expression of VEGF and accelerates vessel closure and ischemia, which eventually reaches levels similar to those seen in RVO. Thus, while the time course is different, in both RVO and diabetic retinopathy increased expression of VEGF plays a major role in the progression of nonperfusion (32, 35), contributing to a positive feedback loop in which VEGF-induced leukostasis causes progressive closure of retinal vessels, increasing VEGF production, which worsens leukostasis and nonperfusion. This suggests that suppression of VEGF is not only important for treatment of macular edema and retinal NV, but is also necessary to halt progression of retinal nonperfusion and worsening of disease. Thus, normalization of VEGF levels should be the goal throughout the entire course of diabetic retinopathy and RVO; to achieve this, sustained delivery of VEGF antagonists or ocular gene transfer to express a VEGF-neutralizing protein is needed.

In addition to demonstrating the need for sustained suppression of VEGF throughout all stages of RVO and diabetic retinopathy, our study has identified additional potential therapeutic targets, VEGFR1 and VCAM-1. VEGFR1 is present on monocytes, macrophages, and neutrophils and mediates their migration along VEGF gradients in vitro and their recruitment into ischemic tissue in vivo (42, 43, 50, 51). Our data indicate that VEGFR1 contributes to leukocyte recruitment into the retinal vasculature when retinal levels of VEGF are high. Since blockade of VEGFR1 reduces leukocyte recruitment in the presence of elevated levels of VEGF, it may provide added benefit to VEGF-neutralizing agents, particularly in patients in whom neutralization of VEGF is incomplete and/or short lived. These data suggest that sustained delivery of a kinase inhibitor that blocks both VEGFR1 and VEGFR2 may have advantages over sustained delivery of a VEGF-binding agent for the long-term treatment of ischemic retinopathies.

High levels of VEGF increased NF-κB transcriptional activity in retinal endothelial cells and in the retina in vivo and increased the product of the NF-κB–responsive gene Vcam1. Interestingly, expression of Vcam1 is increased to a greater extent than other NF-κB–responsive genes in ischemic retina and is further increased much more than other NF-κB–responsive genes by exposure of ischemia retina to oxidative stress; its neutralization prevents exacerbation of ischemia-induced NV due to oxidative stress (52). Since neutralization of VCAM-1 reduces VEGF-induced leukostasis, it could provide added benefit to VEGF suppression in patients with ischemic retinopathies.

Leukostasis has previously been suggested to play a role in the pathogenesis of diabetic retinopathy. Mild leukocyte adhesion due to increased expression of ICAM-1 was observed within 3 days of the onset of hyperglycemia in rats with streptozotocin-induced diabetes and was associated with vascular leakage and endothelial cell death (53, 54). It was attenuated by aspirin, a COX-2 inhibitor, or suppression of TNF-α (55), and it was also reduced in diabetic mice deficient in CD18 or ICAM-1 (56). In hyperglycemic mice, the receptor for AGEs was implicated in upregulation of ICAM-1 and early leukocyte adhesion (57). This mild leukostasis is very different from the severe leukostasis described in the current study, and it is uncertain whether changes occurring within days of the onset of hyperglycemia play a role in vascular changes occurring many years later, but it is possible that mild leukostasis from hyperglycemia-induced upregulation of ICAM-1 could gradually lead to retinal ischemia and upregulation of VEGF that starts the positive feedback loop and progressive worsening of retinal nonperfusion described herein.
The suggestion that sustained suppression of VEGF be considered for treatment of all stages of RVO and diabetic retinopathy may raise concerns with some clinicians and investigators because of the perception that prolonged suppression of VEGF could cause retinal damage. While macular atrophy has been observed in patients with neovascular AMD treated with injections of VEGF-neutralizing proteins, it also occurs as part of the natural history of AMD and has not been observed in patients with DME or RVO treated with prolonged suppression of VEGF. Conditional knockout of Vegf in RPE cells leads to loss of the choriocapillaris and retinal atrophy (58), but it is unlikely that complete elimination of intercellular VEGF signaling can be achieved without targeted gene disruption, and no retinal damage occurred in mice after transgenic expression of a potent VEGF-neutralizing protein for up to 7 months (59) or prolonged blockade of VEGFRs (60). If it were possible to cause retinal atrophy from suppression of VEGF, one would expect it to occur after subretinal injection of an AAV vector expressing a VEGF-binding protein, because RPE cells are strongly transduced by AAV vectors, but this was not observed (61).

In summary, this study demonstrates that high levels of VEGF in the retina recruit leukocytes into the retinal vasculature, at least in part, through stimulation of VEGFR1 and promote endothelial cell adhesion and vessel plugging through activation of NF-xB and increased expression of VCAM-1 as well as other adhesion molecules. This explains the clinical observation of reperfusion of previously nonperfused retinal vessels and reduced progression of nonperfusion in patients with ischemic retinopathies treated with intravitreal injections of a VEGF-neutralizing protein. Our study also indicates that, despite different insults responsible for onset of retinal ischemia in RVO, diabetic retinopathy, and other ischemic retinopathies, they evolve into very similar diseases in which VEGF is the driver of an accelerating spiral of disease progression.

**Methods**

**Mice.** Wild-type C57BL/6 mice, Tet/opsin/VEGF double-transgenic mice (40), and rho/VEGF-transgenic mice (41) were treated in accordance with the Association for Research in Vision and Ophthalmology Guidelines on the care and use of animals in research. Adult C57BL/6 mice were anesthetized and given an intravitreous injection of 50, 100, 200, 500, or 1,000 ng VEGF (R&D Systems) or PBS. Adult Tet/opsin/VEGF mice had expression of VEGF induced by 2 mg/ml doxycycline in drinking water or daily intraperitoneal injections of doxycycline (50 mg/kg on day 0, 25 mg/kg on day 1, and 25 mg/kg on day 2). In some experiments, Tet/opsin/VEGF mice were given an intravitreous injection of 40 μg aflibercept or PBS prior to doxycycline administration. Seven-month-old rho/VEGF mice were given subcutaneous injections of 25 mg/kg aflibercept or PBS every other day for a total of 4 injections.

**Measurement of retinal leukostasis.** Leukostasis was measured as previously described (62). Briefly, mice were anesthetized and chests were opened to expose the heart; the right atrium was cut for drainage, a 27-gauge cannula was inserted into left ventricle, and the vasculature was flushed with PBS for 3 minutes for removal all nonadherent cells. Mice were then perfused over a span of 2–3 minutes with rhodamine- or FITC-labeled Con A (20 μg/ml in PBS, 5 mg/kg; Vector Labs, ThermoFisher). The vasculature was flushed for 4 minutes with PBS to remove residual unbound Con A. In some experiments, the vasculature was also perfused with fluorescein-labeled dextran (Sigma-Aldrich). Eyes were removed and fixed in 2% PFA for 2 hours at room temperature, and retinas were dissected and flat mounted. Retinas were examined by fluorescence microscopy, and the number of intravascular leukocytes throughout the entire retina were counted by an investigator masked with respect to treatment group. To identify subtypes of adherent leukocytes, mice were perfused with FITC-labeled Con A and retinas were dissected, fixed, and permeabilized with 1% Triton X-100 (Sigma-Aldrich) in PBS for 24 hours. Nonspecific binding was blocked by 1% bovine serum albumin (Sigma-Aldrich) in 5% horse serum (Invitrogen). After 3 washes with PBS, retinas were incubated with rat anti-mouse F4/80 (1:200; BioLegend) overnight at 4°C. After washing, retinas were incubated in Alexa Fluor 594–conjugated goat anti-rat secondary antibody (1:500; Invitrogen) for 1 hour at room temperature. Retinas were washed, flat mounted, and examined by fluorescence microscopy.

**Assessment for retinal hypoxia.** The Hypoxyprobe-RedAPC kit (Hypoxyprobe) was used to assess for retinal hypoxia. It utilizes pimonidazole, a 2-nitroimidazole, which is activated in hypoxic tissue and irreversibly binds thiol groups, and an Alexa Fluor 594–conjugated anti-pimonidazole antibody. Tet/opsin/VEGF mice were treated with doxycycline for 3 days and then given an intravitreous injection of 0.5 μl of 500 μM pimonidazole. After 90 minutes, mice were perfused with FITC-labeled Con A as described above. Eyes were removed, fixed in 2% PFA for 2 hours, and stained with the anti-pimonidazole antibody. Retinas were flat mounted and examined by fluorescence microscopy.
Fluorescein angiography. Mice had pupils dilated and were given a 50 μl intraperitoneal injection of sodium fluorescein (AK-Fluor 10%, Akorn), and fundus photographs were taken (Micro III Retinal Imaging Microscope, Phoenix Research Laboratories Inc.). For assessment of vascular perfusion, photographs were taken in the early recirculation phase 1 minute after fluorescein injection (63).

Quantitative RT-PCR. Retinas were dissected, immersed in 300 μl Trizol, and roughly shaken for 2 minutes until completely solubilized. Total RNA was extracted with Direct-zol RNA Kits (Zymo Research) using centrifuge columns pretreated with DNAase to remove contaminating genomic DNA. The concentration of RNA suspended in RNAase-free water was measured with Nanodrops, and 1 μg was reverse transcribed with Superscript III (Invitrogen). Primers specific for mouse VCAM-1, ICAM-1, ICAM-2, integrin β1, integrin α4, VAP-1, E selectin, P selectin, and cyclophilin A (Supplemental Table 1) were added to cDNA and SYBR Green Supermix and run in an iCycler real-time PCR detection system (Bio-Rad). Standard curves were made for each molecule before quantification. Expression of each target gene was normalized to 10^6 copies of cyclophilin A.

Effect of VEGF on NF-κB transcriptional activity in vivo. Two approaches were used to assess the effect of VEGF on NF-κB transcriptional activity in vivo. First, a nonviral gene transfer approach that provides good transduction of retinal vascular endothelial cells (44) was used to express a NF-κB luciferase reporter vector (Assymatrix) in the retina. Briefly, mice were given a subretinal injection of 0.5 μg NF-κB luciferase reporter vector together with 0.25 ng Renilla luciferase plasmid (Promega) in 1 μl PBS. Fourteen days after subretinal injection, mice were given an intravitreal injection of 1 μg VEGF or PBS for control. After 5 days, mice were euthanized; the retina and choroid from each eye were homogenized in 200 μl CERI reagent, and after addition of 5.5 μl CERII reagent, samples were centrifuged at 16,000 g for 15 minutes. Pellets were resuspended in 50 μl NER reagent, and protein concentration was measured. Samples containing 15 μg protein were resolved by 12% Nu-polyacrylamide gel electrophoresis (Invitrogen), transferred to polyvinylidene difluoride membrane (Invitrogen), and blocked with 5% BSA or 5% milk block buffer. The nuclear protein histone 3 was used as a control for loading, and due to the size difference between NF-κB (65 kD) and histone 3 (17 kD), membranes were cut into a relatively high-molecular-weight portion that was incubated in rabbit anti-mouse NF-κB (1:1,000; Cell Signaling) in 5% BSA at 4°C overnight and a relatively low-molecular-weight portion that was incubated in rabbit anti-mouse histone 3 (1:2,000; Cell Signaling) in 5% BSA at 4°C overnight. After washing, membranes were incubated in goat anti-rabbit antibody (1:10,000 Invitrogen) and then SuperSignal West Dura Extended Duration Substrate (ThermoFisher). Blots were imaged and analyzed with a Bio-Rad imaging system.

Measurement of the effect of VEGF on NF-κB activation and target gene expression in cultured retinal endothelial cells. Primary HRECs (Cell Systems) and primary C57BL/6 mouse retinal microvascular endothelial cells (Cell Biologics) were cultured with basal medium (EBM) (Lonza) containing 10% FBS and Single-Quots Kit supplements and growth factors (Lonza). The effect of VEGF on NF-κB transcriptional activity was measured as previously described (52). Briefly, HRECs were starved overnight in Opti-MEM (Life Technologies) and incubated in 25 ng/ml human VEGF for 4 hours, harvested, and total RNA was prepared for qRT-PCR to measure the mRNA for ICAM-1, VCAM-1, ICAM-2, E selectin, P selectin, and VAP-1 as described above. HRECs were harvested and nuclear proteins were extracted with NE-PER Nuclear and Cytoplasmic Extraction Kit as above.

Immunofluorescent staining for VCAM-1. Mouse retinal microvascular endothelial cells were seeded on the chamber slide (Lab-Tek) and stimulated with 25 ng/ml VEGF for 4 hours after starvation overnight. Cells were fixed with 4% PFA, blocked with 5% horse serum, and incubated with (1:100) rat anti-mouse VCAM-1
(6C7.1) (64, 65) overnight at 4°C followed by incubation in goat anti-rat secondary antibody conjugated with Alexa Fluor 594 and Hoechst (Sigma-Aldrich). Cells were examined by fluorescence microscope.

Eyes were frozen in OCT and 10-μm frozen ocular sections were fixed in 4% PFA, blocked with 5% horse serum in 0.05% PBST for 1 hour, and incubated overnight in 6C7.1 rat anti-mouse VCAM-1 and FITC-labeled Grifonia simplicifolia isoelectric B4 (Vector Labs) that selectively stains vascular cells. Slides were rinsed with 0.05% PBST, blocked with 5% horse serum in 0.05% PBST for 25 minutes, and incubated in goat-anti-rat secondary antibody conjugated with Alexa Fluor 594 and DAPI. Slides were examined by fluorescence microscopy.

**Effect of neutralization of VEGFR1 or VCAM-1 on VEGF-induced leukostasis.** Adult wild-type C57BL/6 mice were given an intravenous injection of 100 μg anti-mouse VEGFR1 antibody (n = 10, MF-1, Eli Lilly) or rat IgG (n = 6, Sigma-Aldrich), and 24 hours later, 200 ng VEGF was injected into the vitreous cavity. The MF-1 anti-VEGFR1 clone has previously been demonstrated to bind VEGFR1 on leukocytes (66). After an additional 24 hours, mice were perfused with FITC–Con A, and the mean number of intravascular leukocytes per retina was measured as described above. Tet/opsin/VEGF mice were given an intravenous injection of 100 μg 6C7.1 rat anti-mouse VCAM-1 antibody (n = 10) or rat IgG (n = 10) and 2 mg/ml doxycycline in drinking water. On day 3 of doxycycline treatment, mice were perfused with FITC–Con A, and the mean number of intravascular leukocytes per retina was measured.

**Statistics.** All statistical analyses were performed using Stata software, version 14.2 (StataCorp LP). Two-tailed Student's t test was performed in experiments with a single experiment group compared with control group, while 1-way ANOVA was performed in comparisons among multiple experimental groups and control groups, followed by Bonferroni correction for multiple comparisons. A P value of less than 0.05 was considered as statistically significant, and data are shown as mean ± SEM.

**Study approval.** Protocols were approved by the Johns Hopkins Animal Care and Use Committee.

**Author contributions**

YL, JS, and PAC designed experiments; YL, JS, and SDF performed experiments; YL, JS, SDF, DV, and PAC troubleshooting experiments; YL, JS, SDF, JW, DV, and PAC analyzed and interpreted data; JW performed statistical analyses; YL and PAC wrote the first draft of manuscript; YL, JS, SDF, JW, DV, and PAC edited the manuscript.

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