Dietary potassium regulates vascular calcification and arterial stiffness

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Vascular calcification is a risk factor that predicts adverse cardiovascular complications of several diseases including atherosclerosis. Reduced dietary potassium intake has been linked to cardiovascular diseases such as hypertension and incidental stroke, although the underlying molecular mechanisms remain largely unknown. Using the ApoE-deficient mouse model, we demonstrated for the first time to our knowledge that reduced dietary potassium (0.3%) promoted atherosclerotic vascular calcification and increased aortic stiffness, compared with normal (0.7%) potassium–fed mice. In contrast, increased dietary potassium (2.1%) attenuated vascular calcification and aortic stiffness. Mechanistically, reduction in the potassium concentration to the lower limit of the physiological range increased intracellular calcium, which activated a cAMP response element–binding protein (CREB) signal that subsequently enhanced autophagy and promoted vascular smooth muscle cell (VSMC) calcification. Inhibition of calcium signals and knockdown of either CREB or ATG7, an autophagy regulator, attenuated VSMC calcification induced by low potassium. Consistently, elevated autophagy and CREB signaling were demonstrated in the calcified arteries from low potassium diet–fed mice as well as aortic arteries exposed to low potassium ex vivo. These studies established a potentially novel causative role of dietary potassium intake in regulating atherosclerotic vascular calcification and stiffness, and uncovered mechanisms that offer opportunities to develop therapeutic strategies to control vascular disease.

Introduction

Vascular pathology comprises not only remodeling of resistance vessels, but also stiffening of compliance vessels. In the Cardiovascular Health Study, a prospective multicenter study of 4,476 men and women at 65 years of age or older, 79.9% developed increased carotid artery intima and media thickness (1). As large arteries stiffen, left ventricular hypertrophy develops, which increases the rate of cardiovascular events (2, 3). Arterial stiffness has become an independent predictor of cardiovascular morbidity and mortality, representing an important health problem for the nation as a whole.

Emerging clinical evidence suggests the importance of vascular calcification in aortic stiffening (4–8), and vascular calcification is a risk factor that predicts adverse cardiovascular outcomes in several diseases, often with overlapping vascular complications such as atherosclerosis, diabetes, and chronic kidney disease (9). Previously considered a passive, unregulated, and degenerative process occurring in the arterial media, vascular calcification has now been demonstrated to be a highly regulated process of osteochondrogenic differentiation of vascular cells (10, 11). We and others have shown that osteogenic differentiation and calcification of vascular smooth muscle cells (VSMCs) contribute to vascular calcification in atherosclerosis (12–14). Calcifying VSMCs produce extracellular matrix proteins that further accelerate the development of arterial calcification (15, 16). Therefore, calcification of VSMCs results in alterations in the molecular and cellular functions of VSMCs, the main cell type that determines the vascular tone, and leads to disturbance of the vasculature and arterial stiffening (17).

Epidemiological studies have suggested a role for potassium in the clinical manifestations of cardiovascular disease. Inverse associations between serum potassium levels and risks of chronic kidney disease (18) and metabolic syndrome (19) have been documented in patients along with a higher propensity to develop cardiovascular calcification. In prospective cohort studies, low serum potassium levels are associated with significantly higher risks of cardiovascular mortality compared with patients who had serum potassium levels greater than 4.0 mmol/l.
concentrations within the normal range (20, 21). Reduced potassium is associated with cardiovascular diseases such as hypertension and chronic heart failure, and appropriate dietary potassium intake improves those pathological conditions (22–24). Therefore, serum potassium levels are associated with cardiovascular health and disease; however, the mechanisms responsible for potassium-regulated cardiovascular complications are not fully understood.

At the cellular level, potassium has been shown to reduce VSMC proliferation (25) and inhibit VSMC migration in vitro (26). In animal models, elevation of extracellular potassium levels resulting from increased dietary potassium intake attenuated balloon injury–induced neointimal formation in rat carotid arteries, possibly through inhibition of proliferation and migration of VSMCs (27). In a population-based study, high levels of dietary potassium have been postulated to impair proliferation of VSMCs, and reduce monocyte adherence to vessel walls, thereby retarding the progression of atherosclerosis (28). Nonetheless, a direct effect of dietary potassium on VSMC osteogenic differentiation and the development of vascular calcification in atherosclerosis has not been established and characterized.

In this study, we have provided the first evidence to our knowledge that supports a causative role of dietary potassium in regulating osteogenic differentiation and calcification of VSMCs in vitro and vascular calcification and stiffness in atherosclerotic animals in vivo. Furthermore, we defined key mechanisms underlying low-potassium-induced VSMC calcification involving elevated intracellular calcium, activated cAMP response element–binding protein (CREB), and increased autophagy. These findings provide molecular insights into the regulation of vascular calcification and stiffness by potassium, which explain the impact of dietary potassium intake on cardiovascular complications, and uncover potentially new targets to develop strategies to prevent and treat vascular complications.

Results
Dietary potassium regulated vascular calcification and aortic stiffness in mice. The effect of dietary potassium on atherosclerotic vascular calcification was characterized in vivo using the high-fat-fed ApoE−/− mouse model (12, 13), with dietary intake of standard (0.7% wt/wt), low (0.3% wt/wt), or high (2.1% wt/wt) potassium, as previously reported (29, 30). Mice fed the 0.3% potassium diet exhibited significant increases in vascular calcification, compared with mice fed the 0.7% potassium diet, whereas the 2.1% potassium diet markedly inhibited vascular calcification (Figure 1, A and B). The effects of dietary potassium on vascular calcification were demonstrated in aortic root sections by Alizarin red staining (Figure 1, A and B), as well as descending aortas by total calcium quantification (Figure 1C). It is worth noting that mice fed the 0.3% potassium diet had lower mean serum potassium levels (3.70 ± 0.21 mEq/l), while mice fed the 2.1% potassium diet had higher serum potassium levels (4.73 ± 0.15 mEq/l), compared with levels (4.27 ± 0.23 mEq/l) observed in mice fed the standard (0.7% potassium) diet (Supplemental Table 1; supplemental material available online with this article; https://doi.org/10.1172/jci.insight.94920DS1).

In line with our observation of elevated calcium content in the descending aortas, echocardiographic analysis revealed that the 0.3% potassium diet induced a significant increase in mean pulse wave velocity (PWV) (Figure 1D), an indicator of aortic stiffness (31), suggesting that impaired aortic compliance is associated with low dietary potassium–induced vascular calcification. In contrast, compared with animals fed the 0.7% potassium diet, animals fed the 2.1% potassium diet exhibited inhibited vascular calcification and concurrently reduced PWV, supporting an important role of dietary potassium in regulating vascular calcification and stiffness.

Potassium regulated vascular calcification ex vivo. To determine if there was a direct effect of extracellular potassium level on calcification of the arteries and VSMCs in their natural milieu, we employed an ex vivo ring culture model that we and others have recently developed for histological and quantitative analysis of arterial calcification (32, 33). Based on normal physiological levels of serum potassium in adult C57BL/6 mice (34–36), we determined the effects of potassium at the lower (3.7 mM, low K⁺), middle (5.4 mM, control), and higher (6.0 mM, high K⁺) end of the physiological range on aortic calcification. Consistent with the in vivo results, we found that low potassium markedly enhanced vascular calcification in the aortic media, as demonstrated by Alizarin red staining (Figure 2A), while high potassium inhibited aortic calcification. Quantification of total calcium content demonstrated a significant increase in calcification in aortic rings cultured in medium containing 3.7 mM potassium, which was inhibited by 6.0 mM potassium (Figure 2B). These results demonstrated a direct effect of potassium on the calcification of the aortic media, supporting the role of low potassium in promoting VSMC calcification.
Lower physiological levels of potassium induced VSMC osteogenic differentiation and calcification. To determine the effects of potassium on VSMC calcification in cell culture, we first assessed the effects of extracellular potassium on VSMC viability using potassium at concentrations up to 108 mM, so as to identify a nontoxic level of potassium. Consistent with previous observations (37), we found that potassium, at concentrations lower than 54 mM, did not affect VSMC viability (Supplemental Figure 1). However, at concentrations from 3.7 to 4.7 mM, potassium markedly enhanced VSMC calcification compared with its concentration at 5.4 mM, as determined by Alizarin red staining of matrix calcium (Figure 3A) and total calcium quantification in cell lysates measured by the Arsenazo III method (Figure 3B); such calcification did not occur when potassium levels were maintained at 5.7 mM and 6.0 mM.

We and others have demonstrated that VSMC calcification resembles osteogenic differentiation of bone cells (38, 39); therefore, we determined the effect of potassium on the osteogenic differentiation of VSMCs, as indicated by the changes in the expression of bone markers and smooth muscle markers (13, 38). Increased expression of Runx2, a key osteogenic transcription factor that we have determined to be an important regulator of VSMC calcification (13, 38), was found to be upregulated by low potassium at both protein and mRNA levels (Figure 3, C and D). Consistently, the expression of Runx2-regulated osteogenic markers, including osteocalcin (OC) and alkaline phosphatase (ALP), were induced (Figure 3D), while the SMC marker genes, α-smooth muscle actin (α-SMA) and smooth muscle protein 22 α (SM22α), were markedly reduced concurrently (Figure 3, C and E). These data indicated a direct effect of low potassium...
on promoting VSMC osteogenic differentiation and calcification, via increasing osteogenic markers and decreasing SMC markers.

Activation of intracellular calcium signaling mediated low-potassium-induced VSMC calcification. Potassium deficiency has been shown to increase intracellular calcium in VSMCs (40). As increased intracellular calcium flux has been linked to VSMC calcification (41), we examined whether low potassium may induce VSMC calcification via activation of calcium signaling. We found that potassium at a control condition of 5.4 mM did not affect calcium flux (Figure 4A, dotted line). In contrast, elevation of intracellular calcium was evident in VSMCs within minutes after exposure to a low potassium concentration of 3.7 mM; the increase was sustained over the 30-minute duration (Figure 4A). Consistently, low potassium induced rapid and heightened activation of several known downstream mediators that include protein kinase C (PKC), and calcium-activated CREB (Figure 4B), but did not affect extracellular signal–regulated kinases (ERKs) signaling.

As potassium can be transported via 4 types of potassium channels (42), we determined the roles of these channels in mediating low-potassium-induced VSMC calcification. We found that inhibition of ATP-sensitive potassium channels (K_{ATP}) by glibenclamide, voltage-dependent potassium channels (Kv) by 4-aminopyridine, and calcium-activated potassium channels (K_{Ca}) by TRAM34 and charybdotoxin had no effects (Supplemental Figure 2), while inhibition of inward-rectifier potassium channels (K_{IR}) by barium (43, 44) abolished low-potassium-induced VSMC calcification (Figure 4C). Furthermore, inhibition of potassium-activated calcium signaling by calcium channel inhibitors, verapamil and nifedipine, attenuated low-potassium-induced vascular calcification (Figure 4C). Altogether, these results showed that low-potassium-induced elevation of intracellular calcium signaling mediated its effects on VSMC calcification.

Activation of calcium signaling-mediated CREB was required for low-potassium-induced VSMC calcification. The effects of these inhibitors of potassium or calcium channels on calcium signaling–activated downstream signal CREB were further determined. Consistent with the inhibitory effects on low-potassium-induced VSMC calcification, the low-potassium-induced activation of CREB in VSMCs was abolished by the K_{IR} inhibitor, barium, as well as the calcium channel inhibitors, verapamil and nifedipine (Figure 4D). Furthermore, knockdown of CREB using lentivirus-mediated short hairpin RNA (shCREB) blocked low-potassium-induced VSMC calcification and Runx2 upregulation (Figure 4, E and F), supporting the requirement of intracellular calcium–activated CREB signaling in mediating low-potassium-induced osteogenic differentiation and calcification of VSMCs.

Figure 2. Potassium regulated vascular calcification ex vivo. Aortic rings prepared from wild-type mice were exposed to osteogenic media with control (5.4 mM), low potassium (3.7 mM, Low K+), or high potassium (6.0 mM, High K+) for 3 weeks. (A) Aortic calcification, determined in consecutive sections of the aortic rings by Alizarin red staining (middle panels). H&E staining (left panels) was performed for histology. Higher-magnification images of the boxed areas in the middle panels are shown in the right panels. Representative images from 4 independent experiments performed in duplicate are shown. Scale bars: 200 μm (left and middle panels) and 50 μm (right panels). (B) Total calcium content of aortic rings in each group determined by Arsenazo III in separated sets of experiments. Results shown are normalized by total protein amount. Bar values are means ± SD (n = 4, *P < 0.01). Statistical analysis was performed by 1-way ANOVA followed by a Student-Newman-Keuls test.
As activation of CREB signaling was linked to increased autophagy in human melanoma cells and liver tissues (45, 46) and elevation of autophagy was demonstrated during osteoblast mineralization (47), we determined whether CREB activation may affect autophagy in VSMCs during low-potassium-induced calcification. We found that low potassium induced the microtubule-associated protein 1 light chain 3 in the conjugated form (LC3 II) (Figure 4F), an indicator of autophagy activation (48). CREB knockdown, however, markedly inhibited low-potassium-induced elevation of LC3 II (Figure 4F). Consistently, inhibition of KIR or calcium channels that were upstream of CREB by barium, verapamil, and nifedipine also attenuated low-potassium-induced expression of LC3 II (Figure 4G), implying that intracellular calcium–activated CREB mediates low-potassium-induced VSMC calcification via the regulation of autophagy.

Low potassium promoted VSMC calcification through autophagy. To determine the role of autophagy in low-potassium-induced VSMC calcification, we first assessed the effects of potassium levels on the expression of LC3 II. Increased expression of LC3 II (Figure 4F) was demonstrated in VSMCs exposed to potassium at concentrations that induced VSMC calcification (Figure 3A). Quantitative analysis further confirmed an increased LC3 II/I ratio by low potassium (Figure 5B), indicating that low potassium activated autophagy in VSMCs. In addition, a time-dependent effect of low potassium, at 3.7 mM, in increasing the LC3 II/I ratio and autophagy activation was demonstrated (Supplemental Figure 3).

The essential role of autophagy in mediating low-potassium-induced VSMC calcification was initially determined utilizing 3-methyladenine (3-MA), a pharmacological inhibitor of autophagy. Pretreatment of VSMCs with 3-MA dramatically reduced low-potassium-induced elevation of the LC3 II/I ratio, and concurrently blocked VSMC calcification (Figure 5, C and D). Moreover, we generated stable VSMCs with shRNA knockdown of ATG7, a key regulator of autophagy formation. Similar to the observations with the autophagy inhibitor, the ATG7 knockdown blocked VSMC calcification and inhibited low-potassium-induced elevation of the LC3 II/I ratio (Figure 5, E and F). These results support a definitive role of low-potassium-induced autophagy in mediating its effects on promoting VSMC calcification.
Potassium regulated the activation of CREB and autophagy in vascular calcification ex vivo and in vivo. The roles of activation of CREB and autophagy signals in low-potassium-induced vascular calcification were further determined in the ex vivo ring culture model and in vivo mouse models. As shown in Figure 6A, low-potassium-induced aortic calcification ex vivo was associated with increased activation of intracellular calcium signals, including PKC, CREB, and elevation of LC3 II as well as upregulation of the osteogenic transcription factor Runx2 (Figure 6A). In contrast, high potassium markedly inhibited the activation of PKC and CREB and inhibited LC3 II and Runx2 (Figure 6A). Moreover, increased activation of PKC, CREB, and elevation of LC3 II and Runx2 were demonstrated in arteries from low dietary potassium–fed mice (Figure 6B). Consistently, higher dietary potassium inhibited PKC and CREB activation, and decreased LC3 II and Runx2 (Figure 6B). Altogether, these results further supported an important role for potassium in regulating vascular calcification via effects on calcium signaling, CREB, and autophagy in VSMCs (Figure 6C).
Reduced dietary potassium intake has been linked to the pathogenesis of a variety of human diseases, including atherosclerosis, diabetes, and chronic kidney disease; all of these disease share common vascular complications, such as vascular calcification. Using in vitro cell culture, an ex vivo tissue model, and in vivo atherogenic animals, we have demonstrated that low potassium intake induces VSMC osteogenic differentiation and calcification and promotes atherosclerotic vascular calcification. Concurrently with increased vascular calcification, we have found that PWV, an indicator of arterial stiffness, was increased in animals fed a low-potassium diet (Figure 1D), supporting a direct link between low dietary potassium–promoted atherosclerotic calcification and arterial stiffness. These observations are consistent with previous studies demonstrating that vascular calcification may increase arterial stiffness through the calcification of elastic fibers (49, 50). In sharp contrast, increased dietary potassium inhibited atherosclerotic calcification and improved vascular stiffness in the ApoE−/− mice. Importantly, the increase in vascular calcification and stiffness caused by the reduced-potassium diet occurred with only a small reduction in mean serum potassium concentration, compared with the standard potassium–content diet (3.70 ± 0.21 versus 4.27 ± 0.23 mEq/l; \( P < 0.05 \)). Similarly, increased potassium diets resulted in a small increase in serum potassium concentration (4.73 ± 0.15 mEq/l; \( P < 0.05 \)) and was sufficient to reduce vascular calcification in atherosclerosis. Of note, lower magnesium levels have been reported to be associated with the presence of vascular calcification in clinics (51), as well as in calcification of cultured VSMCs (52). We did not find significant differences in the serum magnesium levels in the low-potassium-fed animals compared with normal- or high-potassium-fed animals (Supplemental Table 1). Therefore, results from these studies support a direct effect of potassium on the calcification of VSMCs, leading to vascular calcification and stiffness. These findings are consistent with previous observations that reduced dietary intake of potassium is associated with increased risk of vascular diseases, such as stroke and hypertension, which were improved by appropriate dietary potassium intake (22–24, 53). These studies uncovered a potentially novel causative link between low potassium and VSMC osteogenic differentiation and calcification in atherosclerosis.

Our studies have added mechanistic insights into our current understanding of the regulation of vascular health and disease by dietary potassium. During the development of atherosclerosis, potassium may regulate multiple cellular processes, as it can affect VSMC proliferation (37), or interplay with sodium homeostasis and a number of neurohormonal pathways that regulate blood pressure (54). Our findings highlight the importance of dietary potassium–regulated VSMC calcification in atherosclerosis.
The in vitro studies in particular demonstrated a direct effect of extracellular potassium levels on VSMC calcification. Although previous studies have shown that potassium reduced VSMC proliferation in vitro (25), the effect of low potassium on promoting VSMC calcification was independent of its effects on VSMC viability (Supplemental Figure 1), suggesting a previously unknown mechanism. Among the 4 known potassium channels on VSMCs, we identified that KIR specifically mediated the effects of low-potassium-induced VSMC calcification. Upregulation of KIR has been reported during osteogenic differentiation of mesenchymal stromal cells; however, the role of KIR in the osteogenic process has not been defined (55). We found that low potassium initiated VSMC calcification through activation of intracellular calcium flux. As KIR was linked to potassium-induced changes in membrane potential of rat arteries (43), it is possible that KIR-mediated membrane depolarization leads to the elevation of intracellular calcium via voltage-gated calcium channels (VGCCs) in VSMCs. In a meta-analysis combining the results of 2 genome-wide association studies, upregulation of CACNA1C, encoding a subunit of the VGCC, was associated with patients with calcific aortic valve disease (56), suggesting a role of VGCCs in vascular calcification. Furthermore, the VGCC blocker nifedipine was found to inhibit coronary arterial calcification in patients with hypertension (57). Consistently, we found that inhibitors of the KIR or VGCCs blocked low-potassium-induced VSMC calcification, which was associated with inhibition of low-potassium-induced calcium signals.

We observed a prolonged increment of intracellular calcium in low-potassium-treated VSMCs (Figure 4A), which was associated with increased VSMC calcification. Intracellular calcium influx resulting from depolarization-induced VGCC opening has been associated with SMC differentiation (58). However, a long-term increase in intracellular calcium is linked to osteogenic differentiation of SMCs, and inhibition of intracellular calcium intake by KCa blockers attenuates calcium phosphate–induced VSMC calcification (59). In addition, inhibition of L-type calcium channels by verapamil has been shown to reduce VSMC calcification (60). Accordingly, differences in the duration of intracellular calcium flux may lead to diverse downstream molecular signals that differentially regulate VSMC physiology and pathology. Unlike physiologic intracellular calcium influx that is associated with excitation-contraction of SMCs, sustained increase in intracellular calcium may lead to VSMC pathology, such as calcification. Our results with the KIR or calcium channel inhibitors have further supported the role of calcium signaling in mediating low-potassium-induced VSMC calcification.

Low-potassium-induced elevation of intracellular calcium led to the activation of CREB signaling, which was required for low-potassium-induced Runx2 upregulation and osteogenic differentiation and activation of autophagy in VSMCs.

Figure 6. Potassium regulated the activation of CREB and autophagy in vascular calcification ex vivo and in vivo.

(A) Effects of low potassium on the activation of calcium-activated cAMP response element–binding protein (CREB) and autophagy signals ex vivo. Aortic rings from wild-type mice were exposed to osteogenic media with control (5.4 mM), low potassium (3.7 mM, Low K+), or high potassium (6.0 mM, High K+) for 3 weeks. Western blot analysis was performed to determine the phosphorylation of protein kinase C (PKC), CREB, and the expression of microtubule-associated protein 1 light chain 3 (LC3) and runt-related transcription factor 2 (Runx2). Representative blots from 3 independent experiments are shown. (B) Effects of dietary potassium on the activation of CREB and autophagy signals in vivo. Western blot analysis was performed to determine the phosphorylation of PKC, CREB and the expression of LC3 and Runx2 in descending aortas of control, low, or high potassium diet–fed mice. Representative Western blot analyses of protein lysates extracted from the descending aortas from at least 3 mice in each group are shown. (C) Schematic illustration of vascular calcification regulated by potassium. Low potassium activates calcium signaling that promotes activation of CREB, which results in chronically excessive autophagy in vascular smooth muscle cells (VSMCs), leading to the development of calcification. High potassium reverses vascular calcification.
ApoE–/– tion were determined using the ApoE-deficient (ApoE–/–). Effects of dietary potassium on pathogenesis of atherosclerotic vascular calcification. Experimental animals

Methods

In summary, we have determined a causative link between reduced dietary potassium and vascular calcification in atherosclerosis and uncovered the underlying pathogenic mechanisms that integrate enhanced intracellular calcium influx, activated CREB signaling, and elevation of autophagy. These findings provide molecular insights into the previously unappreciated regulation of vascular calcification and stiffness by low potassium intake and emphasize the need to consider dietary intake of potassium in the prevention of vascular complications of atherosclerosis.

Methods

Experimental animals. Effects of dietary potassium on pathogenesis of atherosclerotic vascular calcification were determined using the ApoE-deficient (ApoE–/–) mouse model (12, 13). Eight-week-old male ApoE–/– mice (The Jackson Laboratory) were fed a high-fat, high-cholesterol diet (HFD, Harlan Teklad diet TD88137) containing normal potassium (0.7% wt/wt KCl), low potassium (0.3% wt/wt KCl), or high potassium (2.1% wt/wt KCl) for 30 weeks as previously described (29, 30). Both food and fluid intake were given ad libitum. The Institutional Animal Care and Use Committee of the University of Alabama at Birmingham approved all experimental protocols.

Echocardiography and measurement of PWV. At the experimental endpoint, echocardiography was performed with the high-resolution imaging system VEVO 2100 (Visual Sonics) to determine PWV as previously...
described (76). PWV was calculated as the distance divided by the time interval between the pulse wave at the aortic arch and the abdominal aorta (m/s).

**Analysis of serum biochemical parameters.** Mouse blood was collected at euthanization, and serum was separated from nonheparinized blood and assayed for various minerals, enzymes, and compounds (Supplemental Table 1). All the biochemical parameters were determined by Molecular Diagnostic Services, Inc. using a Beckman Coulter AU680 analyzer.

**Tissue harvesting, processing, and analysis.** Heart and aortic tissues were collected, processed, and analyzed as previously reported (12, 13, 32, 76). Descending aorta was used for total calcium measurement by the Arsenazo III method (Stanbio Laboratory), or protein expression and phosphorylation by Western blot analysis. A series of consecutive 7-μm aortic root sections were used for hematoxylin and eosin (H&E) staining for histology and Alizarin red staining (Sigma-Aldrich) for calcification. Stained sections were examined using a Leica M165 FC microscope, and the percentage of positively stained area in each section was quantified using ImageJ software (NIH). All quantitative analyses were performed blindly by at least 2 investigators.

**Ex vivo aortic calcification.** Aortic rings were obtained from descending aortas of wild-type C57BL/6 mice (The Jackson Laboratory) as we previously described (32). Calcification of aortic rings was performed as previously reported (32), using 3-mm mouse aortic rings cultured in osteogenic media containing 1% FBS with low (3.7 mM), normal (5.4 mM), or high (6.0 mM) potassium for up to 3 weeks. Choline chloride, which has been used to compensate for the deficit in the extracellular ionic concentration (77–79) and does not show any effects on vascular calcification (80), was added to culture media to avoid the changes in osmolarity. Total calcium content in the aortic rings was quantified by the Arsenazo III method (Stanbio Laboratory), or protein expression and phosphorylation by Western blot analysis. As previously reported, using 3-mm mouse aortic rings cultured in osteogenic media containing 1% FBS (Stanbio Laboratory), 10 R-G109a), p-ERK (Cell Signaling, 4370), p-PKC (Cell Signaling, 9375), p-CREB (Cell Signaling, 9198), PC3 (MBL, M186-3), and total CREB (Cell Signaling, 9104), and ATG7 (Cell Signaling, 8558); and detect-
Intracellular calcium measurement. Intracellular calcium was measured using a Fluo4 NW calcium assay kit (Molecular Probes) according to the manufacturer's instructions. Briefly, VSMCs seeded on 96-well black clear-bottom plates (Corning Costar) were preloaded with Fluo4 NW fluorescent dye (1 μM) for 30 minutes, and then exposed to growth media containing low (3.7 mM) or normal potassium (5.4 mM). Fluorescence signals were recorded for 30 minutes at 1-minute intervals using the fluorescence microplate reader with excitation at 485 nm and emission at 525 nm (Synergy 2, BioTek). Intracellular calcium was calculated by subtracting the basal fluorescence intensity from the total fluorescence intensity.

shRNA knockdown in VSMCs. Lentiviral vector carrying shRNA specific for murine CREB, ATG7, or control scrambled shRNA (shScr) was purchased from Open Biosystems and packaged into lentiviral particles as previously described (12). Lentiviral transduction was performed by incubating VSMCs with lentivirus in growth media supplemented with 8 μg/ml polybrene, and stable transfectants were selected with puromycin (2 μg/ml) for 2 weeks.

Statistics. All the data are expressed as means ± SD. For multiple group comparison, 1-way analysis of variance (ANOVA) followed by a Student-Newman-Keuls test was performed. A P value less than 0.05 was considered statistically significant.

Study approval. All animal experiments were performed according to protocols approved by Institutional Animal Care and Use Committee at the University of Alabama at Birmingham.

Author contributions
HW and YC designed the study. YS, CHB, YY, and WEB performed the experiments and analyzed the data. YS, CHB, HW, and YC wrote the manuscript. LJD, PWS, and AA participated in study designing and modification, and edited the manuscript.

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