Partially exhausted tumor-infiltrating lymphocytes predict response to combination immunotherapy

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Introduction

Immune regulation in the tumor microenvironment is a complex and multifaceted phenomenon (1). Chronic antigen exposure in the setting of tumor growth and metastasis leads to anergy and dysfunction (exhaustion) of the adaptive immune response, in part through increased engagement of the programmed death 1 (PD-1) receptor with its ligand (PD-L1) (2). The net result of signaling through this axis is an attenuation of the cytotoxic and cytokine-producing capacity of tumor-infiltrating lymphocytes, leading to ineffective antitumor immune responses (2). Inhibition of the PD-1 pathway has proven to be effective
in many solid and hematological tumors (3–5). Understanding the host and tumor microenvironment factors that mediate response to PD-1 blockade can provide critical insights into the host antitumor immune response (6, 7). These factors include tumor mutation burden (6–8), host microbiome (9, 10), disease stage (11), lactate dehydrogenase (LDH) levels (11), baseline tumor size (11), and the presence of liver metastasis (12–14). These factors have been referred to as the cancer immunogram (15) or the cancer immune set point (16), and include both tumor intrinsic and extrinsic factors (17).

We have recently shown that the fraction of partially exhausted cytotoxic T lymphocytes (peCTLs, CTLA-4hiPD-1hi) strongly correlates with response to anti–PD-1 monotherapy (18). Functionally, these cells may contain tumor-antigen-specific T cells (19, 20), and they demonstrate a partially exhausted phenotype which is hypothesized to be activated by treatment with anti–PD-1 (18, 21). Here, we examine how the relative abundance of peCTLs correlates with specific host factors, as well as response to immune checkpoint monotherapy and combination therapy.

**Results**

A total of 112 unique melanoma tumors were analyzed. While age was not correlated, sex significantly correlated with the percentage of peCTLs, with males having a larger proportion of these cells compared with females (males, 25.95% vs. females 17.5%, \( P = 0.041 \), Mann-Whitney test) (Figure 1, A and C). In contrast, the percentage of tumor-infiltrating regulatory T lymphocytes (Tregs, as defined by CD45+CD3+CD4+Foxp3+CD25+ cells) did not correlate with age or sex (Figure 1, B and D).

Given prior data regarding the reduced response to anti–PD-1 therapy in patients with elevated LDH and liver metastasis (11, 22), we examined these and other disease characteristics, including American Joint Committee on Cancer (AJCC) stage (stage III vs. IV). Disease stage is known to be correlated with survival (23),
while the presence of liver metastasis has been reported to inversely correlate with response to checkpoint inhibitors in melanoma, non–small cell lung cancer, and recently also in bladder cancer (13, 22). While no correlation with disease stage was observed (Figure 2, A and B), the presence of liver metastasis was associated with reduced percentages of tumor-infiltrating peCTLs (Figure 2, C and D). Elevated LDH levels have been reported to correlate with reduced objective response rates (ORRs) to anti–PD-1 monotherapy (4, 11, 24, 25). Thus, we examined whether there was an association between LDH levels and both peCTL and Treg percentages. Neither of these T cell subsets was associated with elevated LDH (Figure 2, E and F).

While the relative abundance of tumor-infiltrating peCTLs correlates with response to anti–PD-1 monotherapy (18), it is currently unknown whether this metric predicts response to ipilimumab/nivolumab combination therapy. Thus, we quantified peCTLs and ORRs by Response Evaluation Criteria in Solid Tumors version 1.1 (RECIST v1.1) criteria (26) to ipilimumab/nivolumab combination therapy (n = 29) and compared these with anti–PD-1 monotherapy (n = 64) (Figure 3). Across all patients treated with either single-agent or combination checkpoint therapy, there was a significant correlation between pe-
percentage of peCTLs and ORR (CR+PR). Responders had a peCTL fraction of 30.5% versus 15.4% in non-responders ($P < 0.0001$, Mann-Whitney test). Responses to PD-1 monotherapy were significantly more likely when the peCTL fraction was greater than 20% (65.8% vs. 5%, $P = 0.0001$), while with combination immunotherapy, responses were not significantly different with high or low peCTL fraction (72.7% vs. 38.9%, $P = 0.1281$). We have previously reported that a peCTL fraction of 20% or lower is associated with lack of response to anti–PD-1 monotherapy (18). Using the same metric, only 1 of 18 patients (5.6%) with peCTL 20% or lower responded to anti–PD-1 monotherapy, whereas 7 of 20 patients (35%) with peCTL 20% or lower responded to combination ipilimumab and nivolumab. The relative risk of response in the low-peCTL population was 6.3 (95% CI 1.2–37.7), and this difference was statistically significant ($P = 0.045$, Fisher’s exact test). In contrast, there was no difference in patients with high (>20%) peCTL. In these patients, 70% of patients treated with ipilimumab/nivolumab responded, while 66.7% of patients treated with anti–PD-1 monotherapy responded ($P > 0.9999$, Fisher’s exact test). In the illustrative example shown in Figure 3C, a patient with a peCTL fraction of 12% failed to respond to anti–PD-1 monotherapy (Figure 3C), while a patient with a peCTL fraction of 3.6% responded to ipilimumab/nivolumab combination therapy (Figure 3C), illustrating the lower threshold needed for combination immunotherapy response. There was no significant correlation between ORR and Treg percentage (Figure 4, Treg panel) ($P = 0.0925$, Mann-Whitney test), while peCTL level was highly correlated with response status across all patients (Figure 4, peCTL panel) ($P = 0.0001$, Mann-Whitney test).

Discussion

While PD-1 inhibition represents a major advance in the treatment of melanoma and other cancers, it is well recognized that in most cancers only a minority of patients benefit from this approach (3, 4, 25), and that host and tumor factors play a role in response to these agents (11, 15, 17). Previously, we studied the abundance of tumor-infiltrating peCTLs in freshly isolated melanoma tumors using a novel assay that uses multiparameter flow cytometry and found that the fraction of peCTL/total CD8+ T cells within tumors directly correlated with response to anti–PD-1 monotherapy in a 40-patient cohort (18). Here, we utilized this approach in an expanded cohort of patients to understand the relationship between host factors and the abundance of peCTLs within the tumor microenvironment. Additionally, studying Tregs in the tumor microenvironment using a fractional Treg score (Tregs/total CD4+ T cells) found no correlation between the frequency of these cells with host demographics and disease characteristics. Importantly, female sex and the presence of liver metastasis significantly correlated with reduced peCTLs and reduced ORRs, while AJCC disease stage (stage III vs. IV) and elevated LDH did not. Although the association of elevated LDH with reduced response rate to anti–PD-1 therapy is well documented (4, 14), the lack of association with reduced peCTLs suggests that other immune cells such as NK cells (27) or tumor macrophages (28) may be involved in mediating the effects of LDH on immunotherapy. The effects of liver metastasis have been recently shown to reduce ORR and progression-free survival (PFS) to anti–PD-1 antibodies in melanoma and non–small cell lung cancer (22), and also in urothelial cancer (13) in immunotherapy-treated patients, and remains an area of active investigation. Preclinical data suggest that activated CD8+ T cells can be selectively deleted in the liver (29) and that liver tolerance (30) is operational in allotransplantation. Further studies in other tumor types and animal models may reveal additional information about the mechanism of liver metastasis–induced tolerance (31). The effect of female sex on reduced response rate to PD-1 has been recently reported (22, 32). Additional data are needed to determine the validity and generalizability of this observation.

Combined blockade of the PD-1/CTLA-4 checkpoints represents a major advance in immunotherapy, resulting in increased response rates and PFS in melanoma (33). However, this approach also exposes patients to high rates of severe autoimmune toxicities (34), and careful patient selection is key to using this potent combination versus single-agent anti–PD-1 (35). Recent studies have utilized PD-L1 immunohistochemical staining as a biomarker for response, with recent data showing that patients with low tumor PD-L1 expression (<1% or <5%) have better PFS and overall survival (OS) with anti–PD-1 combination therapy than with anti–PD-1 monotherapy (33, 36). In this current study, we sought to improve resolution by stratifying patients by low (≤20%) versus high (>20%) peCTL frequency. Using this cutoff, derived from our previous analysis in patients treated with anti–PD-1 monotherapy (18), we found that low-peCTL patients had increased response rates with combination checkpoint blockade, while high-peCTL patients had equivalent response rates with either combination or monotherapy. Taken together, these results suggest that fewer
tumor-infiltrating peCTLs are required to achieve a response to ipilimumab/nivolumab combination therapy. Since peCTLs express CTLA-4 (18, 21), our data imply that dual blockade is additive or synergistic when peCTLs are infrequent in the tumor microenvironment. However, when peCTLs are abundant the additional stimulus of CTLA-4 blockade may not be necessary for maximal antitumor effect. These results com-

Figure 3. Treatment response to anti–programmed death 1 (PD-1) monotherapy or ipilimumab/nivolumab combination therapy correlated with pretreatment partially exhausted cytotoxic T lymphocytes (peCTLs, CTLA-4 hi-PD-1 hi). (A) Flow cytometric data from metastatic tumors taken and pregated on live CD45+CD3+CD8+ cells. peCTLs as a percentage of total CD8+ T cells is shown on the y axis, and response to either anti–PD-1 monotherapy (n = 64) or ipilimumab/nivolumab combination (n = 29) is designated by color. Patients with partial or complete responses (PR+CR) by Response Evaluation Criteria in Solid Tumors version 1.1 (RECIST v1.1) are shown in green, and patients with stable or progressive disease (SD+PD) are shown in red. (B) Responders and nonresponders to anti–PD-1 monotherapy and combination therapy relative to pretreatment peCTL count. Low peCTL count is 20% or lower, high peCTL count is greater than 20%. For patients with low peCTLs (left), the difference in objective response rate was 35% for ipilimumab plus nivolumab, while it was 5.6% for anti–PD-1 monotherapy (P = 0.045 by Fisher’s exact test). The difference for patients with high peCTLs (middle) was not significantly different: 70% for ipilimumab plus nivolumab and 66.7% for anti–PD-1 monotherapy. For the overall cohort (right), the difference was also not statistically significant: objective response rate was 46.7% for ipilimumab plus nivolumab versus 46.3% for anti–PD-1 monotherapy. NS, not significant. (C) Illustrative examples for 2 patients. Left: A nonresponder to anti–PD-1 monotherapy with 12% peCTLs with progression in the liver (white arrow). Right: A responder to ipilimumab plus nivolumab combination therapy with 3.6% peCTLs shows a response in a large left axillary mass (white arrow).
To complement the PD-L1 subset analysis data from the PD-1/CTLA-4 randomized combination trial discussed above, though studies of peCTLs in other tumor types and in animal models are needed to validate these observations.

There is a complex interplay between the environment, the microbiome, the host immune system, and the tumor (6, 15, 17). The results presented here provide a framework with which we can begin to understand why specific patient subsets have a paucity of tumor-infiltrating peCTLs and thus a lower likelihood of responding to immune checkpoint blockade. Future studies will require the discovery of specific therapies that reinvigorate this population and are tailored toward specific constellations of host factors and peCTL frequency.

**Methods**

**Study design and tumor sample procurement.** Between January 2014 and August 2016, 102 patients with advanced melanoma at UCSF underwent biopsy for tumor-infiltrating lymphocyte profiling. Biopsies of accessible melanoma tumors were obtained after patients provided informed consent under the UCSF Committee on Human Research Protocol 138510. Biopsy samples were obtained with a 16- or 18-gauge needle, or a 4-mm punch tool. Fresh tumor samples were immediately placed on ice and transported to the laboratory for dissociation and analysis. Of these, 64 patients underwent treatment with monotherapy PD-1, 29 patients underwent treatment with ipilimumab plus nivolumab, and 9 patients with other regimens (anti–PD-1/indoleamine deoxygenase combination).

**Study approval.** Melanoma tumor biopsies were performed after written informed consent was obtained from patients under the UCSF Committee on Human Research Protocol 138510.

**Treatment outcome groups and efficacy analysis.** Two treatment outcome groups, responders and nonresponders, were defined using radiologic imaging following anti–PD-1 monotherapy or ipilimumab/nivolumab combination treatment. Responders included patients with tumor target lesions that met RECIST v1.1 criteria for complete response (≥99% reduction in the target lesions) or partial response (≥30% reduction in target lesions). Nonresponders included patients with tumor target lesions that met RECIST v1.1 criteria for progression (≥20% increase in the target lesions) or stable disease (<30% reduction or <20% increase in tumor target lesions). Efficacy and immunological data available as of August 2016 were included in all the analyses. The efficacy analysis was limited to best overall response (BOR), defined as the best tumor response according to RECIST v1.1 criteria from the start of treatment to the time of disease progression or death. The laboratory team was blinded to all demographic and clinical data (including treatment response) throughout all studies.

**Flow cytometric analyses.** Multiparameter flow cytometry was performed on pretreatment samples obtained from metastatic tumors as previously described. Freshly isolated samples were minced and digested overnight with buffer consisting of collagenase type 4 (Worthington, 4188), DNase (Sigma-Aldrich, DN25-1G), 10% FBS, 1% HEPES, and 1% penicillin/streptavidin in RPMI medium. Single-cell
suspensions were double filtered, centrifuged, and counted. For intracellular cytokine analysis, digested tumor cell suspensions were stimulated with PMA/ionomycin for 4 hours as previously described (37). Approximately $2 \times 10^6$ cells were stained with multiple fluorochrome-conjugated monoclonal antibodies. The following antibodies were used (eBioscience, unless otherwise stated): anti-hCD3 (UCHT1), anti-hCD8 (RPA-T8), anti-hCD45 (HI30), anti-CD4 (SK3), anti-Foxp3 (PCH101), anti–hCTLA-4 (14D3), anti–PD-1 (EH12.2H7; Biolegend), anti–HLA-DR (LN3), anti–PD-L1 (MIH1), and LIVE/DEAD Fixable Aqua Dead Cell Stain (Life Technologies). Data were acquired by an LSRFortessa (BD Biosciences) and analyzed using FlowJo software (Tree Star, Inc.).

**Flow cytometry standardization and gating strategy.** All the samples were fresh and acquired by the Fortessa at different time points. To standardize voltages over time, Sphero Ultra Rainbow beads (Spherotech) were used to calibrate and normalize to baseline intensity. Gates were determined using both isotype control antibody staining and an internal negative control cell population (i.e., PD-1 and CTLA-4 expression on CD3– cells).

**Statistics.** Patients with tumors exhibiting a threshold of less than 20% CTLA-4 hiPD-1hi CD8+ T cells were noted to have infrequent responses in initial testing. This threshold was used in subsequent correlative analyses. The Mann-Whitney test was used to compare peCTL and Treg levels across paired demographic and disease variables assuming nonparametric distributions. Fisher’s exact test was used to analyze contingency tables. A $P$ value of less than or equal to 0.05 was considered significant. In all figures quantifying flow cytometric data, the mean value ± SEM is depicted. Statistical analysis was done using Prism software (GraphPad).

**Author contributions**

AID, KL, KKT, and MDR designed the research studies. AID, KL, KKT, KM, MLP, and PS conducted experiments. AID, KL, KKT, KM, J Liu, LSL, MP, and LC acquired data. AID, MDR, KL, KKT, KM, J Lee, MP, PS, AN, LC, and APA analyzed data. AID, JH, KL, KKT, MDR, MA, and MFK wrote the manuscript.

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