

## **Supplemental Tables and Figures**

**The metalloproteinase-proteoglycans ADAMTS7 and ADAMTS12 provide an innate,  
tendon-specific protective mechanism against heterotopic ossification**

**Timothy Mead et al**

### Supplemental Table 1.

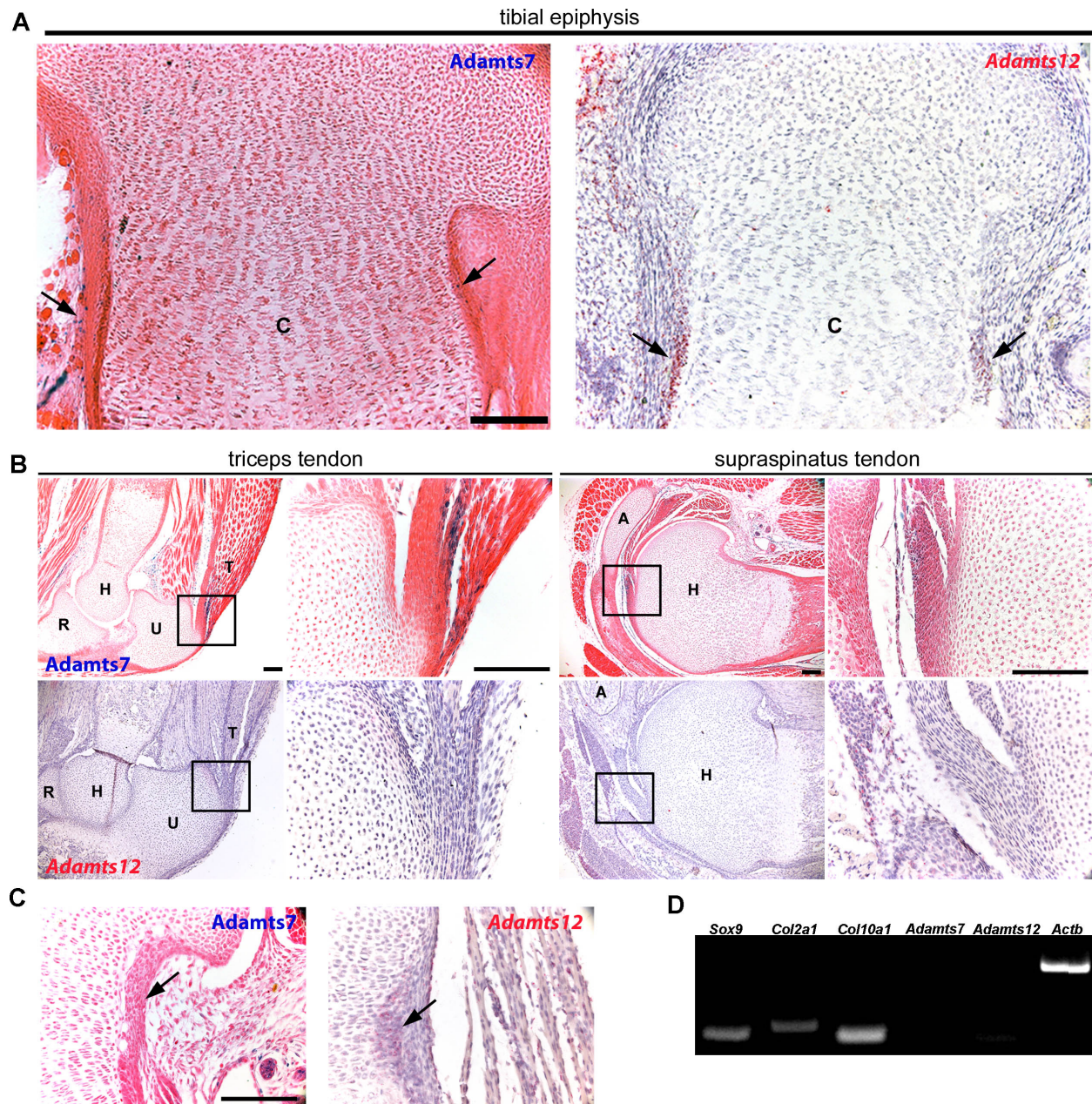
#### Antibodies used for immunofluorescence and western blots.

<b>Antibody</b>	<b>Product #</b>	<b>Source</b>	<b>Dilution</b>
anti-biglycan	LF-159	Dr. Larry Fisher, NIH	1:200
anti-fibromodulin	LF-150	Dr. Larry Fisher, NIH	1:200
anti- decorin	LF-114	Dr. Larry Fisher, NIH	1:200
anti-aggrecan	AB1031	EMD-Millipore	1:500
anti-collagen X	ab58632	Abcam	1:1000
anti-pSMAD159	#13820	Cell Signaling	1:200
anti-GAPDH	MAB374	EMD-Millipore	1:5000
anti-ADAMTS7	AB45044	Abcam	1:100
anti-ADAMTS12	24934-1-AP	Proteintech	1:50

**Supplemental Table 2.****Quantitative Real-Time PCR primer pairs**

<i>Gapdh</i>	Forward: 5'-TGGAGAAACCTGCCAAGTATGA-3'
	Reverse: 5'-CTGTTGAAGTCGCAGGAGACA-3'
<i>Adamts7</i>	Forward: 5'-GGAGTGAGGACCCAGATAAGTA-3'
	Reverse: 5'-CGTGCATAGGTGAAGGTAGTG-3'
<i>Adamts12</i>	Forward: 5'-CCAAAGGTGCGAGGGATATAAG-3'
	Reverse: 5'-ACCCTCCGTTGAGGTAGTATT-3'
<i>Scx</i>	Forward: 5'-GCACCTTCTGCCTCAGCAAC-3'
	Reverse: 5'-TTCTGTCACGGTCTTTGCTCA-3'
<i>Mkx</i>	Forward: 5'-ACAATCCACACACAGGGCCG-3'
	Reverse: 5'-GGTCTGCCGCCAGCTTTTATC-3'
<i>Tnm</i>	Forward: 5'-CTACAGCAATGGCGAGAAGAAGAAG-3'
	Reverse: 5'-GACCTACAAAGTAGATGCCAGTGTATC-3'
<i>Coll1a1</i>	Forward: 5'-GTCCGAGGTCCTAATGGAGATGC-3'
	Reverse: 5'-GGTCCAGGGAATCCGATGT-3'
<i>Col3a1</i>	Forward: 5'-GAGGAATGGGTGGCTATCCT-3'
	Reverse: 5'-GGTATCCAGGAGAACCAGGAG-3'
<i>Acan</i>	Forward: 5'-CTGTCTATCTGCACGCCAACC-3'
	Reverse: 5'-CCTCTTCACCACCCACTCCGA-3'
<i>Coll10a1</i>	Forward: 5'-CCAGGACACAATACTTCATCCCATAACC-3'
	Reverse: 5'-CCAGGAATGCCTTGTTCTCCTCTTAC-3'
<i>Runx2</i>	Forward: 5'-CCACAGAGCTATTAAAGTGACAGTG-3'

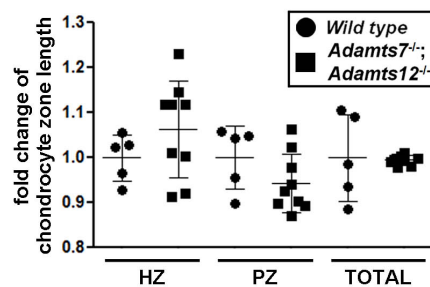
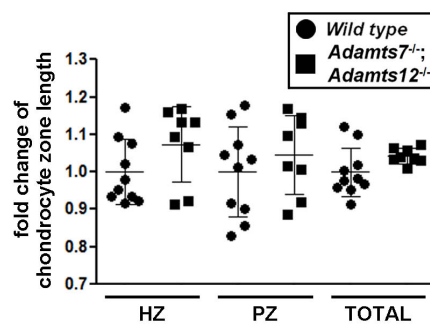
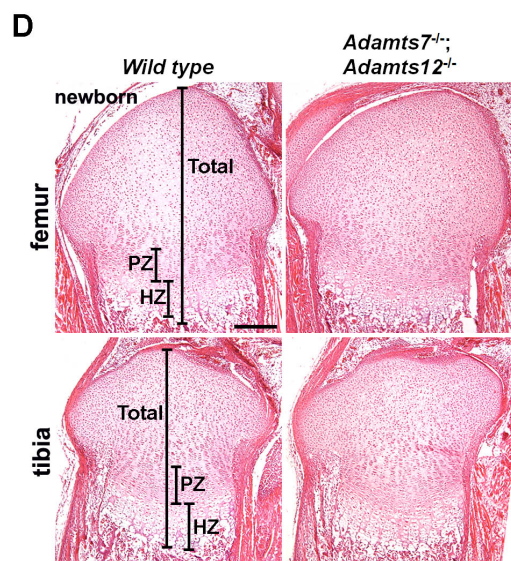
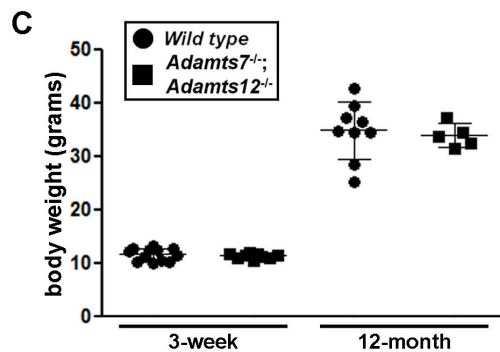
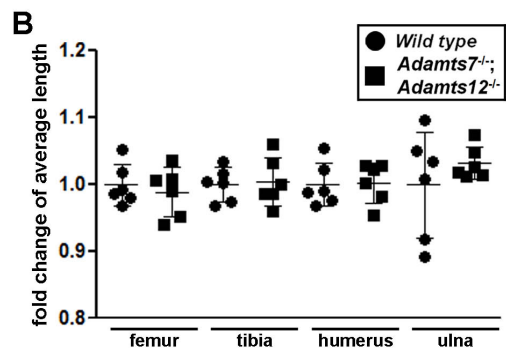
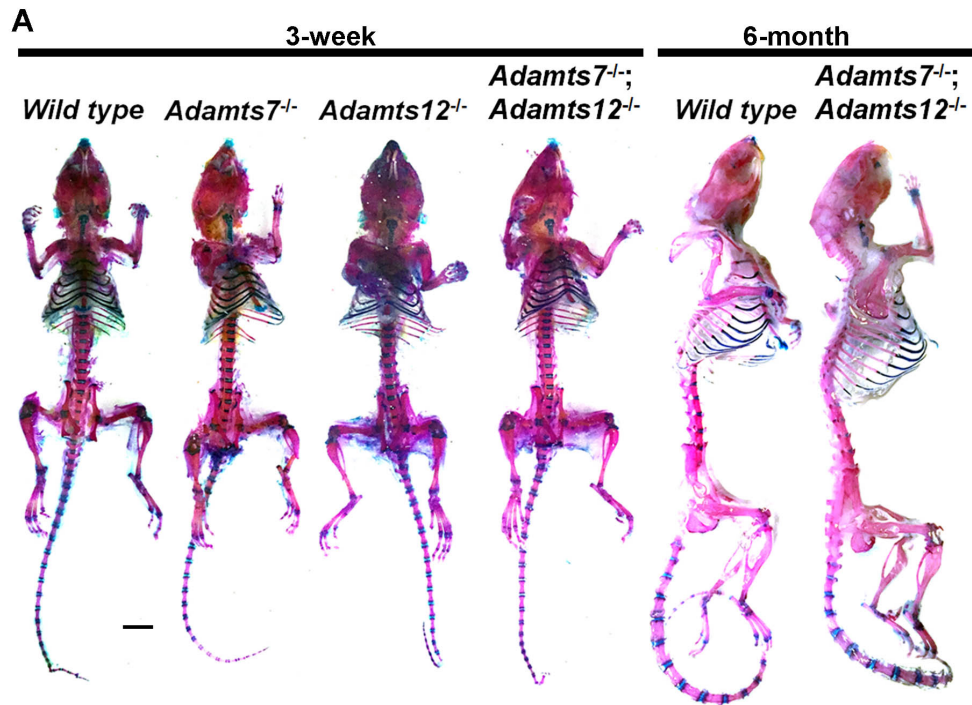
	Reverse: 5'-AACAAACTAGGTTTAGAGTCATCAAGC-3'
<i>Sp7</i>	Forward: 5'-CTCTCCATCTGCCTGACTCC-3'
	Reverse: 5'-CCAAATTTGCTGCAGGCT-3'
<i>Bgn</i>	Forward: 5'-ATTGCCCTACCCAGAACTTGAC-3'
	Reverse: 5'-GCAGAGTATGAACCCTTTCCTG-3'
<i>Fmod</i>	Forward: 5'-CAAGGCAACAGGATCAATGAG-3'
	Reverse: 5'-CTGCAGCTTGGAGAAGTTCA-3'
<i>Dcn</i>	Forward: 5'-GACTCCACGACAATGAGATCACC-3'
	Reverse: 5'-GTTGCCATCCAGATGCAGTTC-3'
<i>ADAMTS7</i>	Forward: 5'-CTTCTGCGAGGACATGGATAAT-3'
	Reverse: 5'-CCCCTGAGACACCACTTATTC-3'
<i>ADAMTS12</i>	Forward: 5'-TGGGAAACAGTGGCAAGATAG-3'
	Reverse: 5'-TGCTCAAGGATTGGGAAGTG-3'
<i>BGN</i>	Forward: 5'-AACTAGTCAGCCTGCGCCT-3'
	Reverse: 5'-GTCCCAGAAGCCTCTCTGCT-3'
<i>FMOD</i>	Forward: 5'-AGCAGCCTCCTTGAGCTAGA-3'
	Reverse: 5'-CAGAAGCTGCTGATGGAGAA-3'
<i>DCN</i>	Forward: 5'-AATGCCATCTTCGAGTGGTC-3'
	Reverse: 5'-AGCAATGCGGATGTAGGAGA-3'
<i>GAPDH</i>	Forward: 5'-AGCCTCAAGATCATCAGCAATG-3'
	Reverse: 5'-CTTCCACGATACCAAAGTTGTCAT-3'



**Supplemental Figure 1. *Adamts7* and *Adamts12* are not expressed in growth plate cartilage or forelimb tendons.** (A) 18.5 day-old embryonic tibia shows lack of *Adamts7* ( $\beta$ -gal staining, blue nuclei, eosin counterstain is red) and *Adamts12* expression (RNA in situ hybridization (red signal)) in the growth plate cartilage (C), but both are expressed in the perichondrial groove of Ranvier (arrows). The data are representative of n=5. (B) 18.5 day-old embryo forelimbs show sparse *Adamts7* ( $\beta$ -gal staining, blue nuclei, eosin counterstain is red) and *Adamts12* expression (RNA in situ hybridization (red signal)) respectively in triceps and supraspinatus tendons. The data are representative of n=6. R; radius; H, humerus; U, ulna; T, triceps; A, acromion. (C) 18.5 day-old mouse embryo humeral head shows *Adamts7* and *Adamts12* expression in the perichondrial groove of Ranvier (arrows). The data are representative of n=4. (D) RT-PCR of

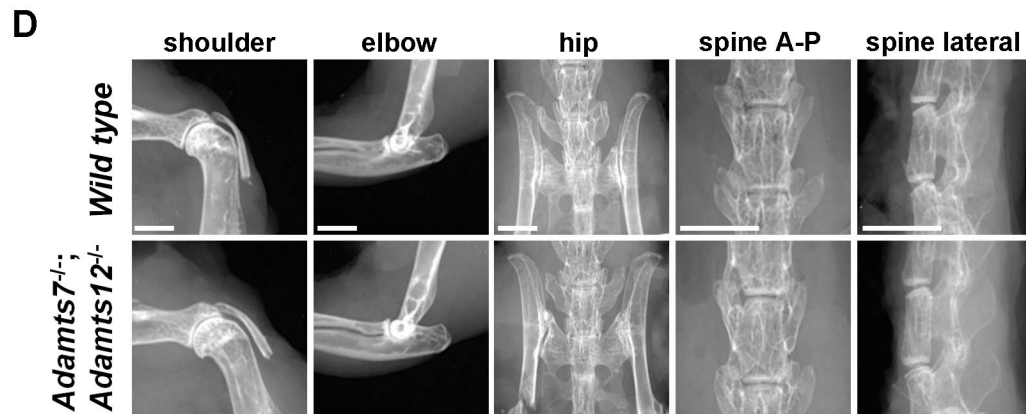
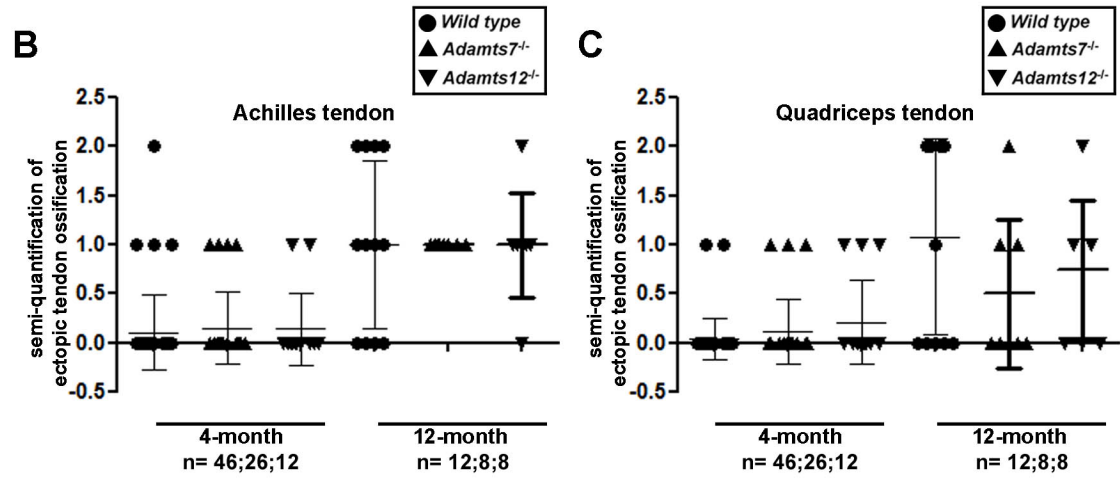
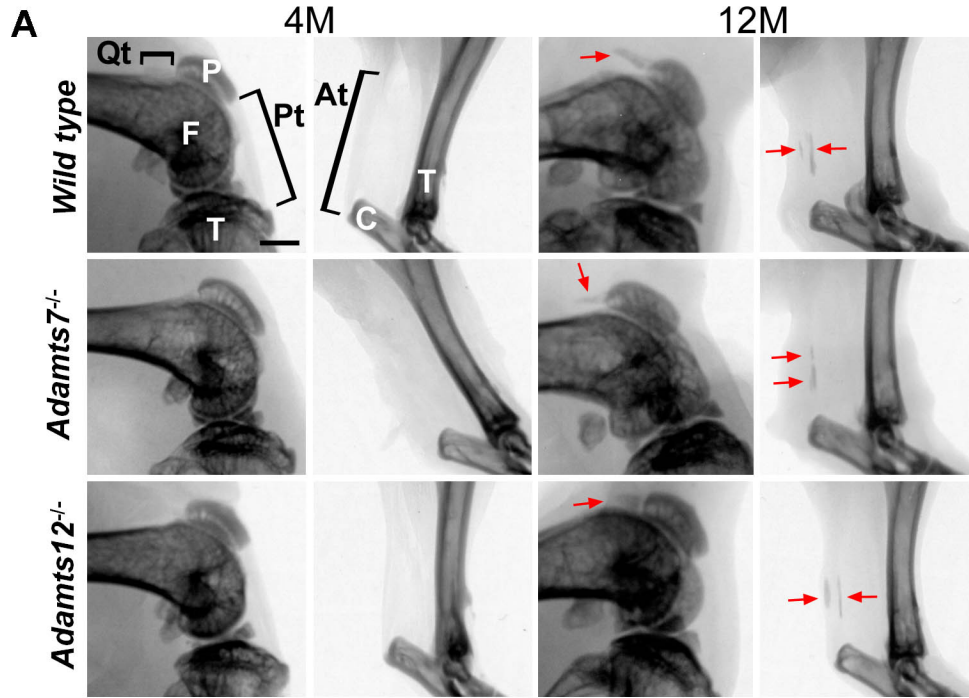
18.5 day-old embryo distal femoral cartilage reveals expression of chondrocyte markers *Sox9*, *Col2a1* and *Col10a1*, but neither *Adamts7* nor *Adamts12*. n=3. Scale bars: 100µm.



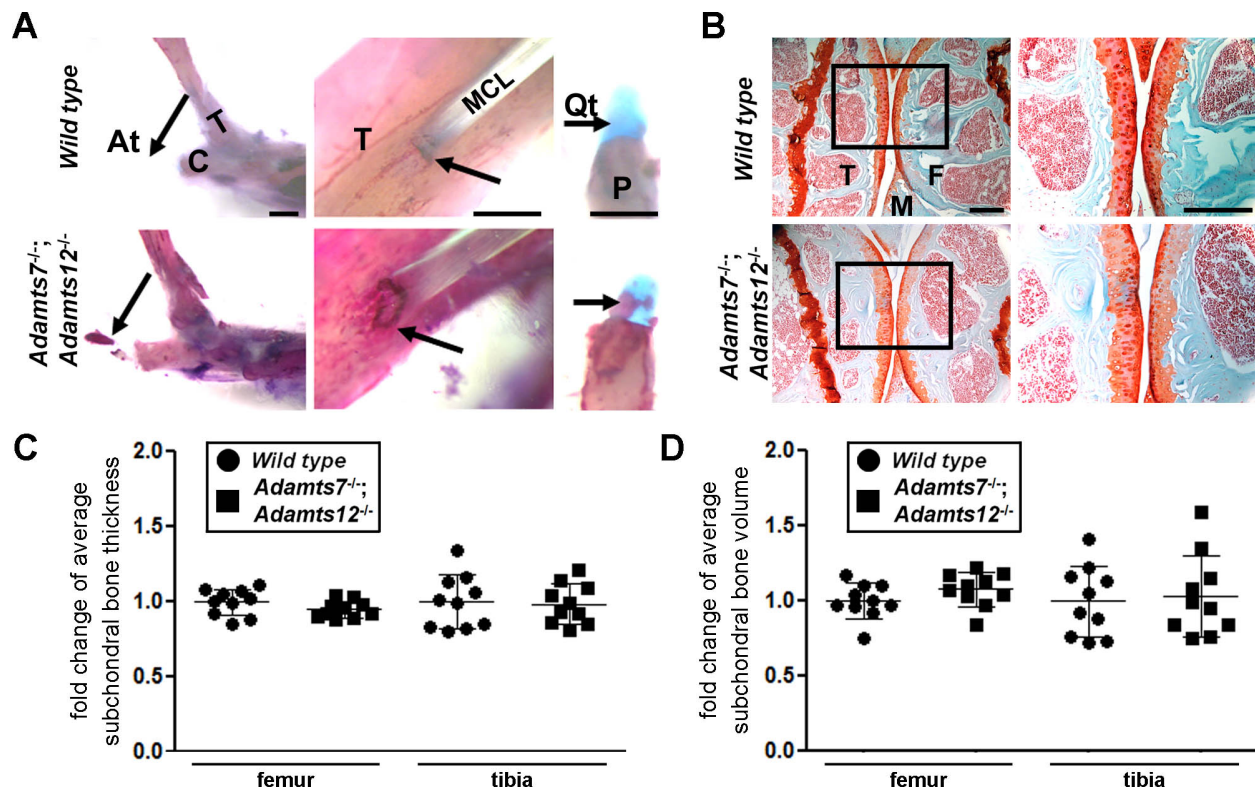


**Supplemental Figure 2. *Adamts7<sup>-/-</sup>;Adamts12<sup>-/-</sup>* mice have normal skeletal patterning and growth.** (A) Alizarin red-Alcian blue stained skeletal preparations of 3 week-old wild type, *Adamts7<sup>-/-</sup>*, *Adamts12<sup>-/-</sup>* and *Adamts7<sup>-/-</sup>;Adamts12<sup>-/-</sup>* mice reveals no skeletal patterning, structural or growth abnormalities. Data are representative of n=5. At 6 months of age, Alizarin red-Alcian blue stained skeletal preparations show that *Adamts7<sup>-/-</sup>;Adamts12<sup>-/-</sup>* mice have normal skeletal dimensions and maturity. Data are representative of n=6. (B) No difference was observed in the femoral, tibial, humeral and ulnar length in 6 month-old mice. n=6. (C) Comparable body weights of wild type and *Adamts7<sup>-/-</sup>;Adamts12<sup>-/-</sup>* mice at 3 weeks and 12 months of age. n=12 at 3 weeks, n=9 at 12 months. (D) Representative H&E-stained sections of newborn femur and tibia showing no change in growth plate dimensions. HZ, hypertrophic zone; PZ, pre-hypertrophic zone; Total, total growth plate. Femur: n=10, 8, respectively. Tibia: n=5, 9, respectively. Scale bars: 1cm in A; 200 $\mu$ m in D. Error bars represent  $\pm$  SEM. Significance was determined by the Student *t*-test.

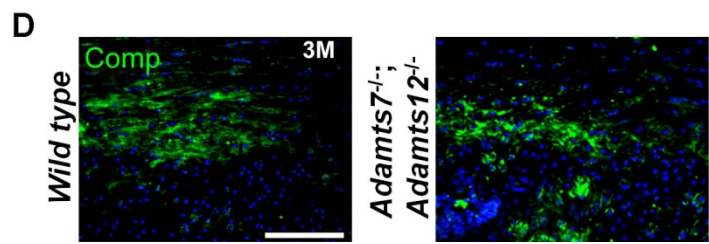
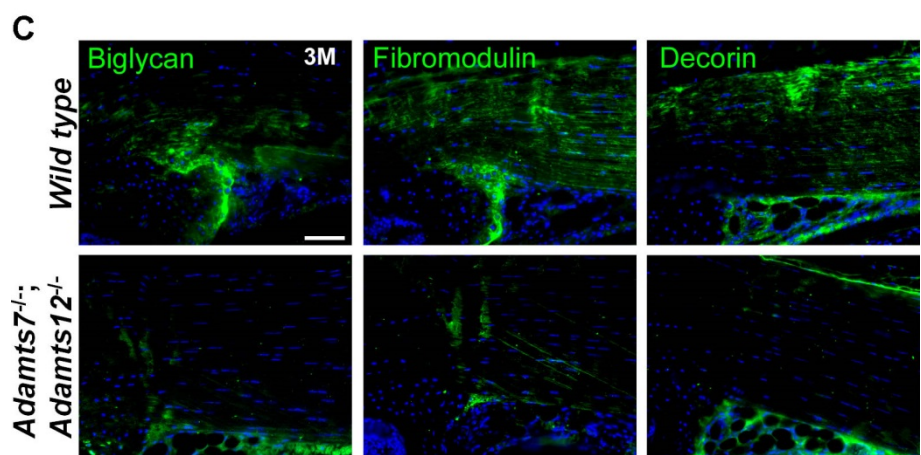
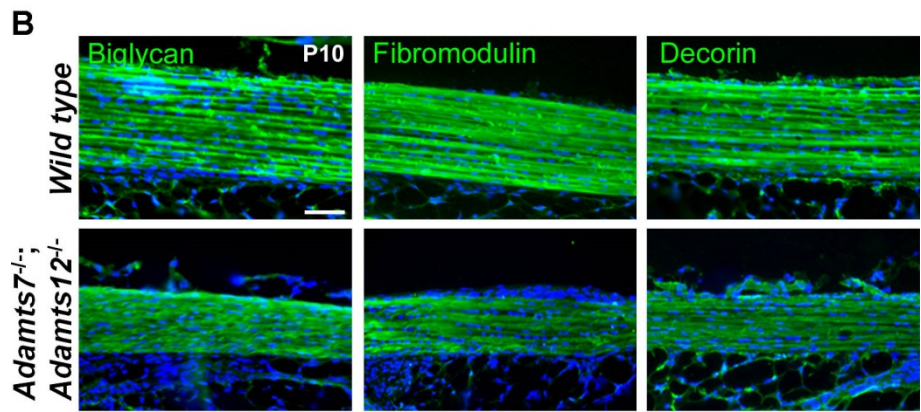
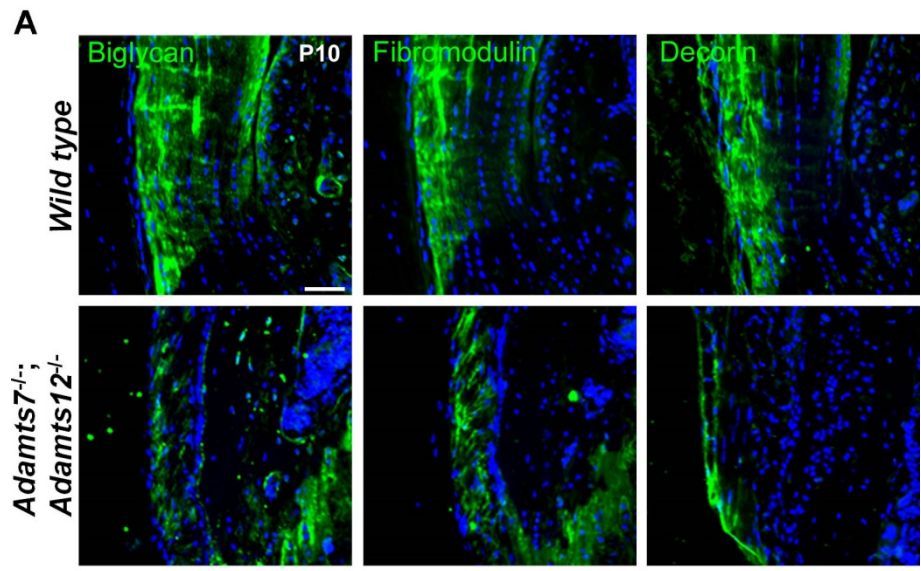




**Supplemental Figure 3. Absence of ectopic ossification in *Adamts7<sup>-/-</sup>* and *Adamts12<sup>-/-</sup>* mice.** (A) Lateral radiographs of 4 month- and 12 month-old hindlimbs reveal no radiopacities in *Adamts7<sup>-/-</sup>*, *Adamts12<sup>-/-</sup>* and wild type tendons at 4 months. At 12 months, wild type, *Adamts7<sup>-/-</sup>* and *Adamts12<sup>-/-</sup>* mice have minimal radiopacity (red arrows) within the quadriceps and Achilles tendon. P, patella; F, femur; T, tibia; C, calcaneus, At, Achilles tendon; Pt patellar tendon. (B-C) No progression of radiopacity in *Adamts7<sup>-/-</sup>* and *Adamts12<sup>-/-</sup>* Achilles and quadriceps tendons as compared to control. (D) No soft tissue radiopacities are evident in forelimb, pelvis and spine tendons in 18 month-old wild type and *Adamts7<sup>-/-</sup>*; *Adamts12<sup>-/-</sup>* mice. Data are representative of n=6. Scale bars: 1 mm. Error bars represent  $\pm$  SEM. Significance was determined by the Student *t*-test.

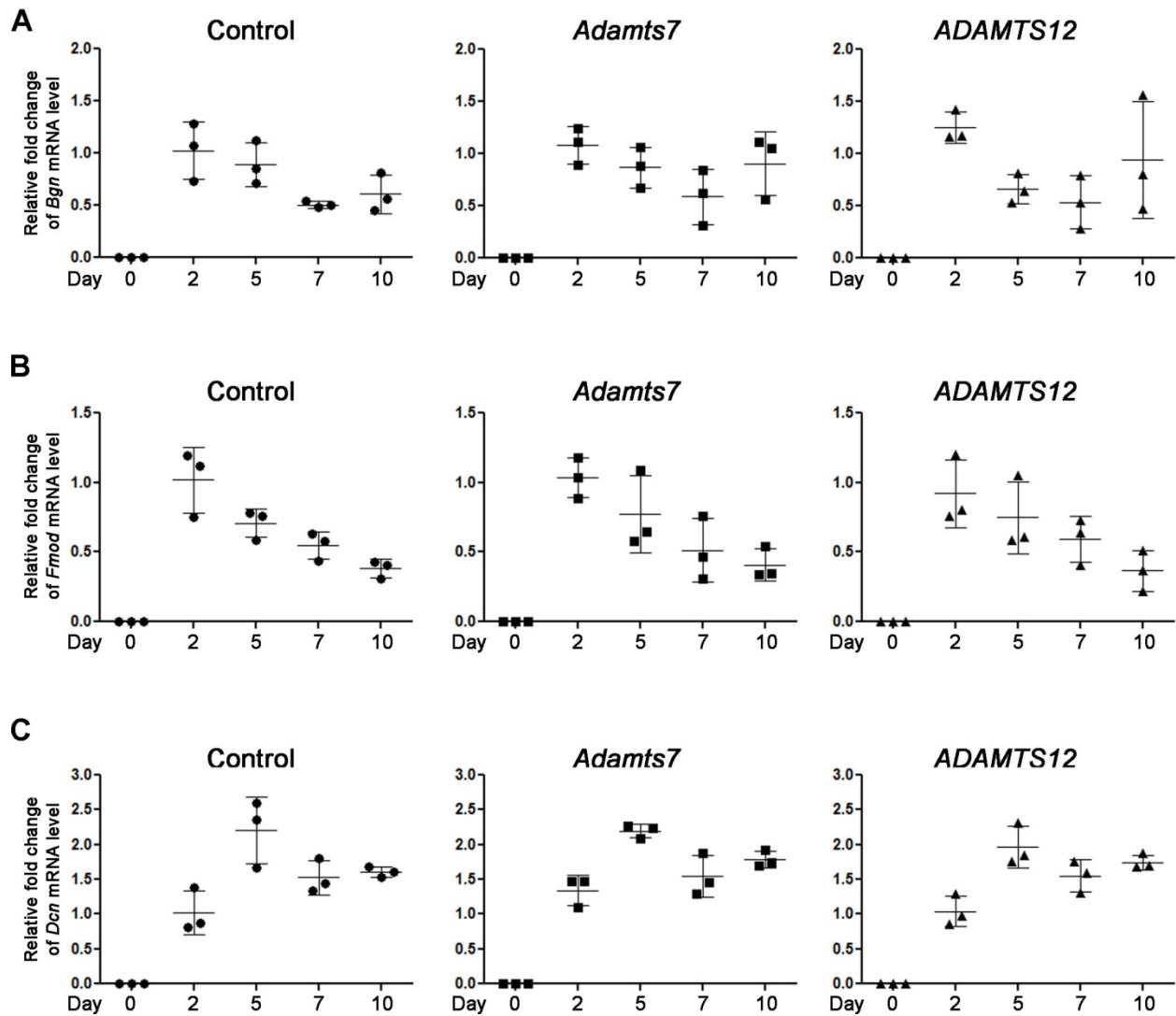


**Supplemental Figure 4. Heterotopic ossification, no arthritic change and normal subchondral bone in *Adamts7<sup>-/-</sup>;*Adamts12<sup>-/-</sup>* hindlimbs. (A) Alizarin red-alcian blue stained skeleton preparations of 6 month-old wild type and *Adamts7<sup>-/-</sup>;*Adamts12<sup>-/-</sup>* hindlimbs reveal ectopic ossification in Achilles tendon (At), medial collateral ligament (MCL) and quadriceps tendon (qt). Data are representative of n=8. (B) Safranin O stained 18 month-old *Adamts7<sup>-/-</sup>;*Adamts12<sup>-/-</sup>* hindlimbs shows normal articular cartilage on the femoral and tibial surfaces. M, meniscus; F, femur; T, tibia; P, patella. Images are representative of n=5. (C-D) No change in subchondral bone thickness (C) or volume (D) of 18 month-old wild type and *Adamts7<sup>-/-</sup>;*Adamts12<sup>-/-</sup>* femur and tibia. n=10. Scale bars: 1 mm in A; 200µm in B. Error bars represent ± SEM. Significance was determined by the Student *t*-test.******

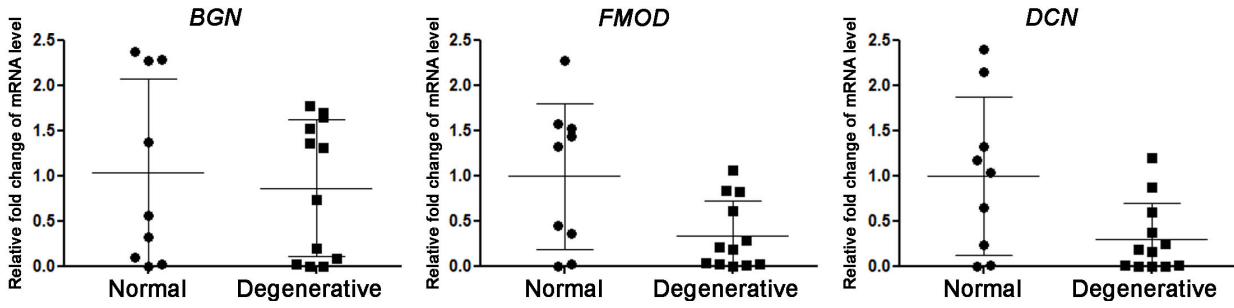


**Supplemental Figure 5. Reduction of small leucine-rich proteoglycans in *Adamts7<sup>-/-</sup>*; *Adamts12<sup>-/-</sup>* patellar tendons. (A-B)** Reduced staining of biglycan, decorin and fibromodulin (green) in 10 day-old patellar tendons near the insertion site **(A)** and mid-substance **(B)**. Sections were counterstained with DAPI (blue). Data are representative of n=3. **(C)** Reduced biglycan, decorin and fibromodulin (green) staining in 3-month-old (3M) *Adamts7<sup>-/-</sup>*; *Adamts12<sup>-/-</sup>* patellar tendons. Data are representative of n=3. **(D)** No change in cartilage oligomeric matrix protein (Comp) (green) staining in 3-month-old (3M) *Adamts7<sup>-/-</sup>*; *Adamts12<sup>-/-</sup>* quadriceps tendons. Sections were counterstained with DAPI (blue). Data are representative of n=3. Scale bars: 100µm.





**Supplemental Figure 6. No change in small leucine-rich proteoglycan mRNA in 10T1/2 pellet cultures.** (A-C) qRT-PCR reveals no significant reduction of *Bgn*, *Fmod* or *Dcn* mRNA in differentiated 10T1/2 cells subjected to chondrogenic culture conditions as compared to control. n=3. Error bars represent  $\pm$  SEM. Significance was determined by a Student *t*-test.



**Supplemental Figure 7. No change in small leucine-rich proteoglycan mRNA in degenerative adult human biceps tendons.** qRT-PCR reveals no significant change of *BGN*, *FMOD* or *DCN* mRNA in degenerative samples. n=9, 12 respectively. Error bars represent  $\pm$  SEM. Significance was determined by a Student *t*-test.