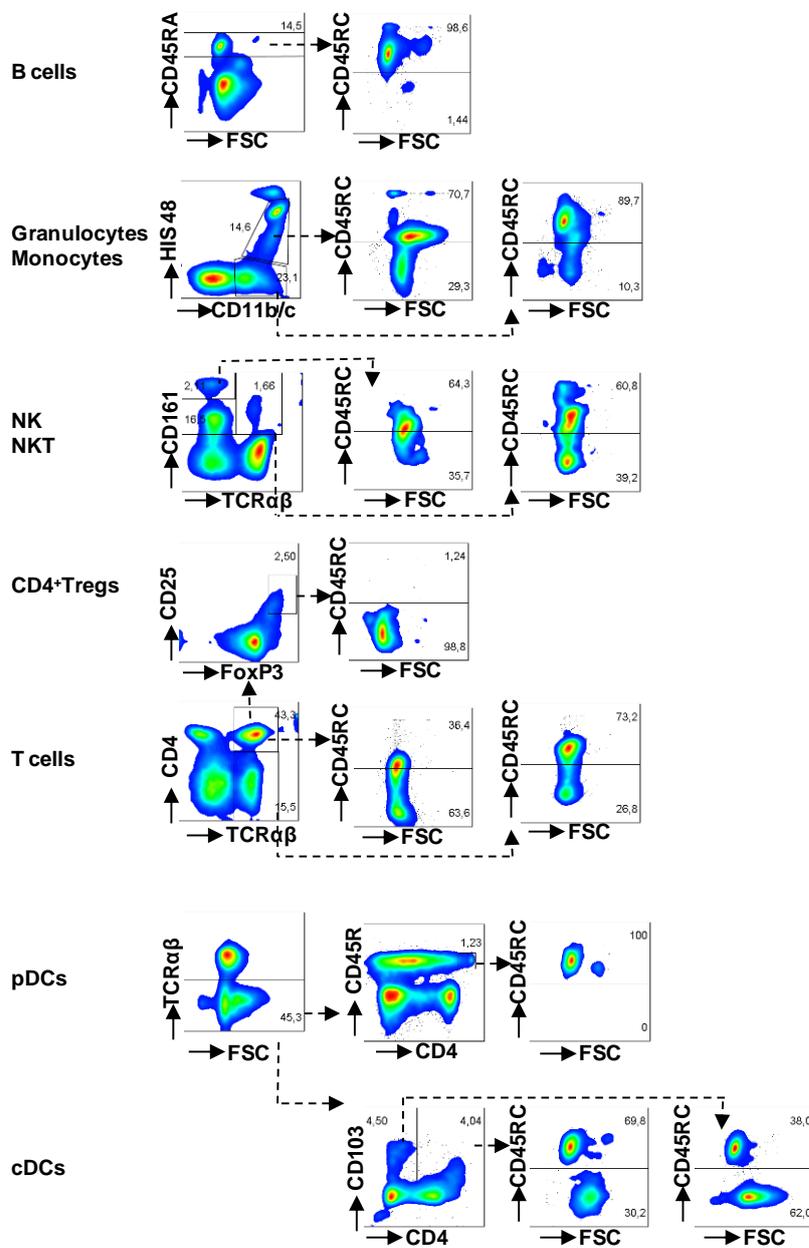
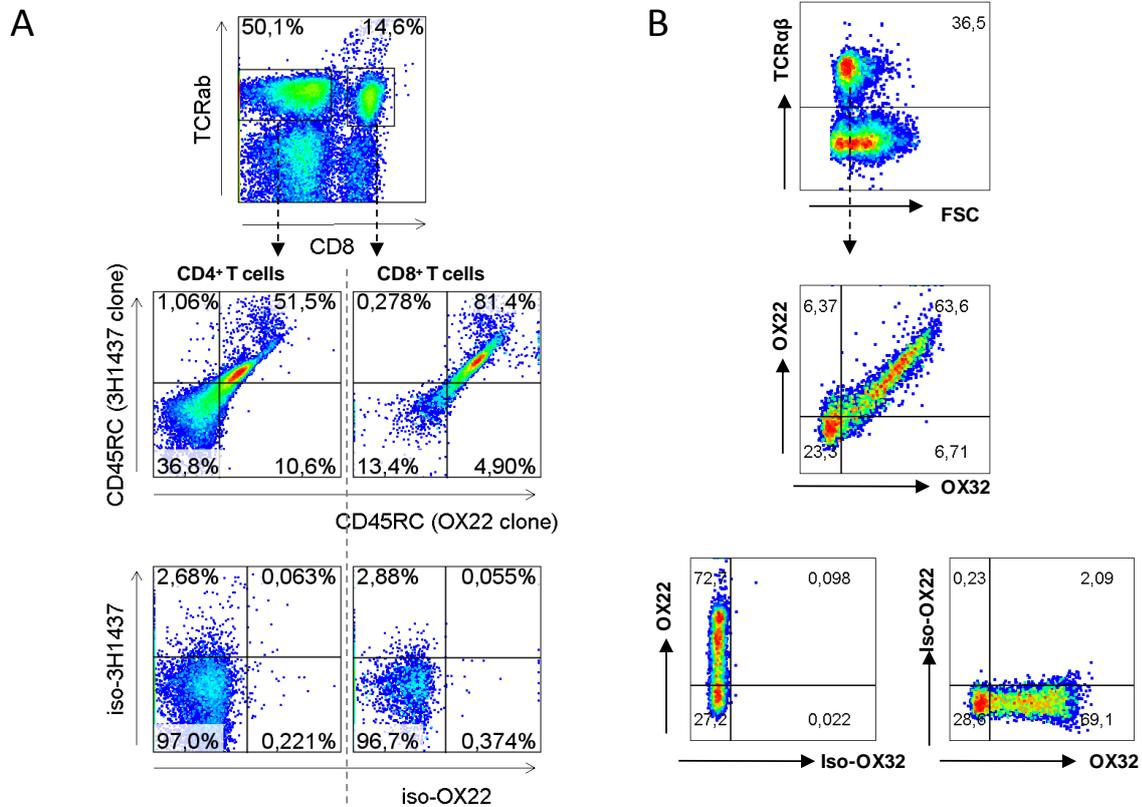


Picarda, Bézie, Boucault et al., Suppl. Figure 1

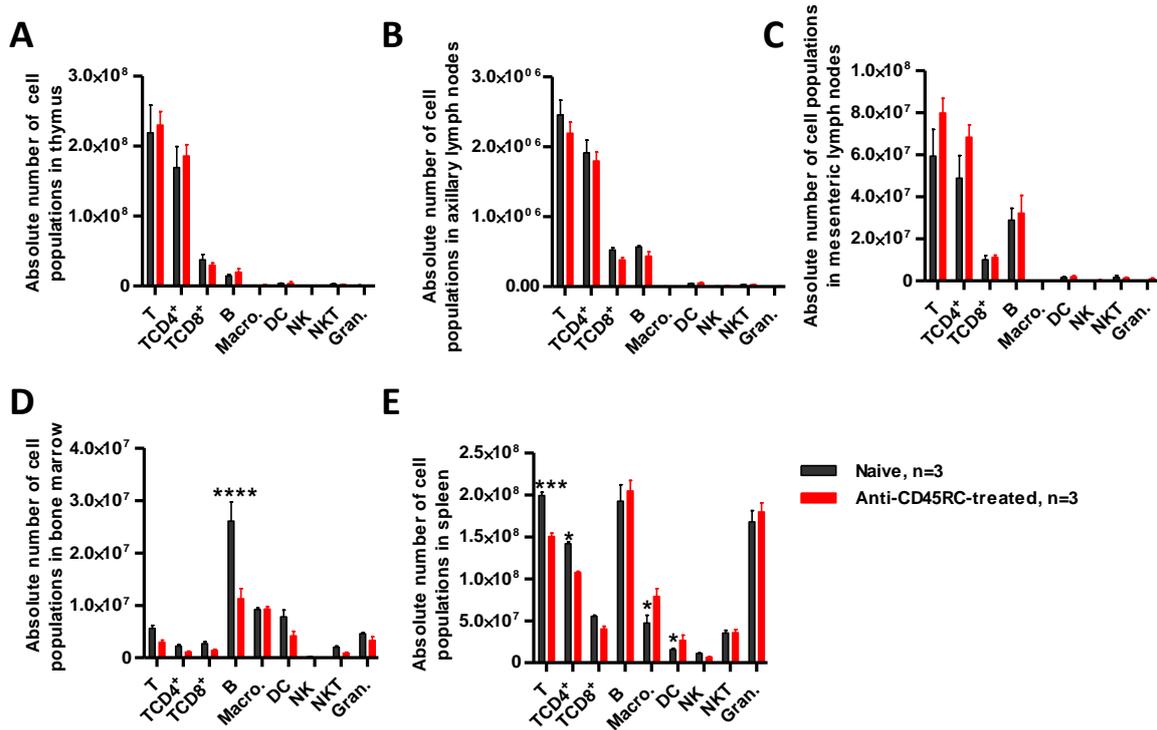


**Supplementary Fig. 1 Ex vivo staining with anti-CD45RC MAb on rat cells. Gating strategy of CD45RC analysis on cell subset from PBMCs from rats. Cells were first gated on morphology, excluding doublets cells and DAPI<sup>+</sup> dead cells. Representative experiment of 6.**

Picarda, Bézie, Boucault et al., Suppl. Figure 2

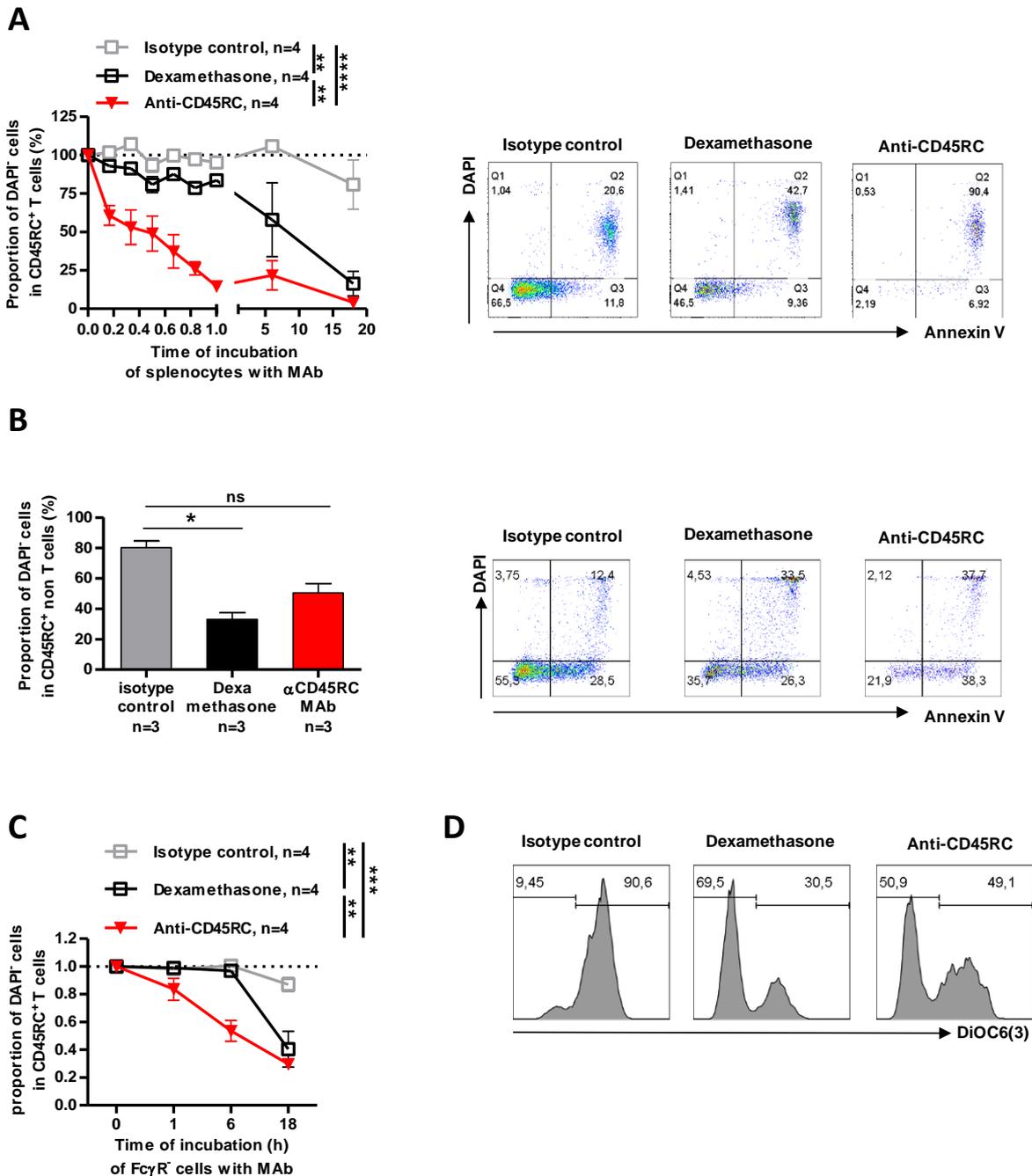


**Supplementary Fig. 2 Flow cytometry staining using distinct anti-CD45RC MAbs for CD45RC<sup>+</sup> cells tracking.** PBMCs were recovered from naive animals and analyzed for CD45RC staining using **three** distinct Abs recognizing a different epitope, OX22, OX32 and 3H1437 clones. Cells were gated on living CD4<sup>+</sup> or CD8<sup>+</sup> T cells for OX22 and 3H1437 staining (**A**) or living T cells for OX22 and OX32 staining (**B**), stained successively and analyzed by flow cytometry. A representative experiment of 2 individuals.



**Supplementary Fig. 3 Short-term anti-CD45RC treatment resulted in minimal modification of leucocytes composition in non-blood compartments of long term tolerant recipients.** (A) Thymus, (B) Mesenteric and (C) axillary lymph nodes, (D) bone marrow, and (E) spleen were analyzed for cell subsets composition (T cells, B cells, macrophages, DCs, NK cells, NKT cells and granulocytes) 120 days after transplantation and anti-CD45RC treatment, and compared with naive rats. n=3 for all groups. Friedman test, Dunn's post test. \*p<0.05; \*\*\*p<0.001; \*\*\*\*p<0.0001.

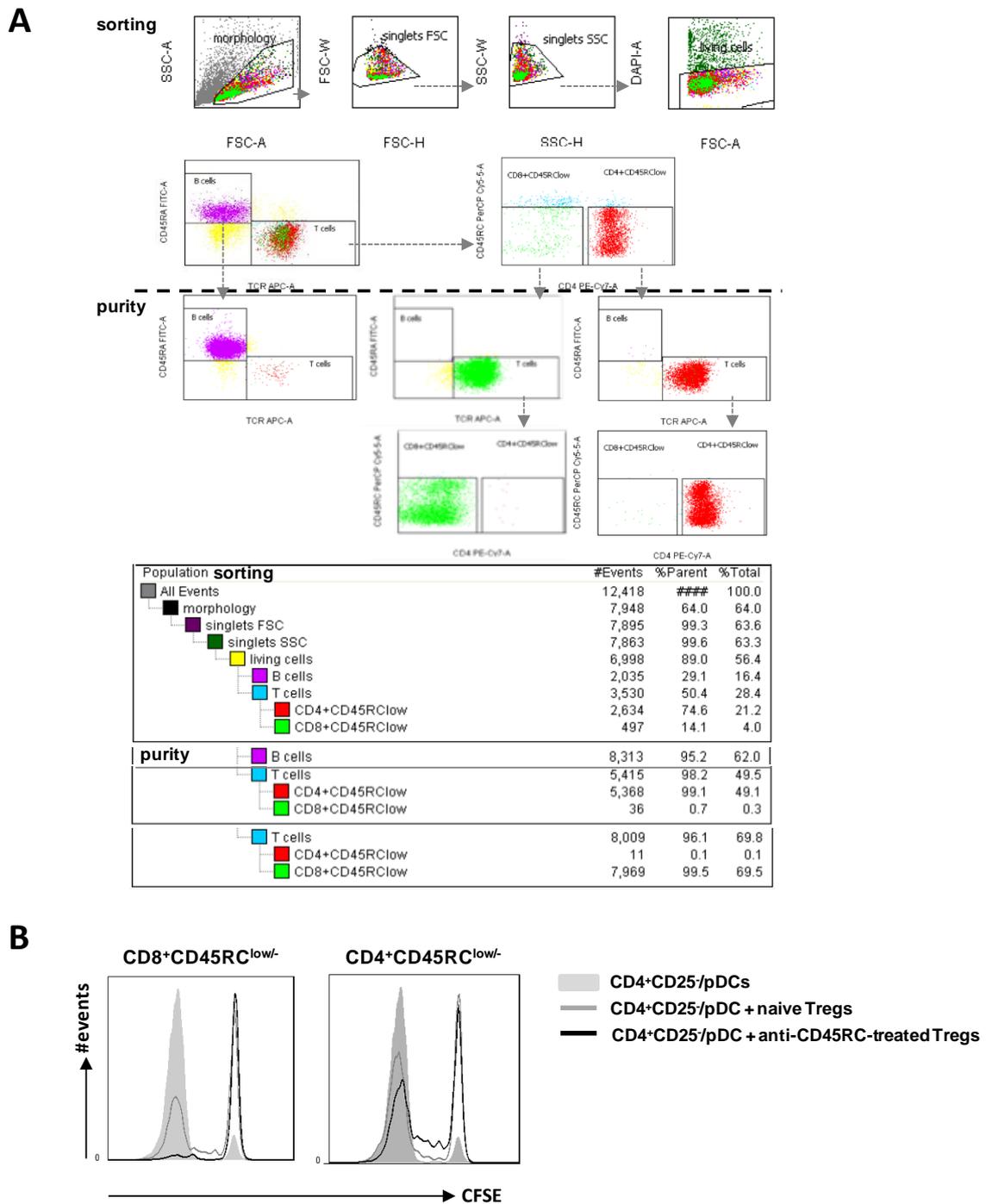
Picarda, Bézie, Boucault et al., Suppl. Figure 4



**Supplementary Fig. 4 Efficient apoptosis induction following CD45RC targeting critically contributes to tolerance induction to cardiac allograft in rat.** Total splenocytes were incubated with anti-CD45RC MAb or isotypic controls in presence of heat-inactivated rat serum (i.e. absence of complement). Dexamethasone was used as positive controls. CD45RC<sup>+</sup> T cells for 10 min to 18h (A) or non-T cells for 18h (B) were analyzed by flow cytometry for DAPI labeling. Right : representative experiment at 1h. A: Two Way ANOVA

RM test. \*\* $p < 0.01$ , \*\*\*\* $p < 0.0001$ . B: Friedman and Dunn's post test \* $p < 0.05$ . (C) Splenocytes depleted from  $Fc\gamma R^+$  cells were incubated with anti-CD45RC MAb or isotype control in presence of heat-inactivated frozen rat serum for 10 min to 18h. Dexamethasone was used as positive controls. CD45RC<sup>+</sup> T cells were analyzed by flow cytometry for DAPI labeling. Two Way ANOVA RM test. \*\* $p < 0.01$ , \*\*\*\* $p < 0.001$ . (A-C) Results are expressed as relative proportion of positive cells among CD45RC<sup>+</sup> T cells  $\pm$  SEM. n=4 for all groups. (D) Total splenocytes were incubated with anti-CD45RC MAb or isotypic controls in presence of heat-inactivated frozen rat serum for 1 to 18h. Dexamethasone was used as positive controls. CD45RC<sup>+</sup> T cells were analyzed by flow cytometry for DiOC6(3) labeling. One representative experiment. Two Way ANOVA RM test. \*\* $p < 0.01$ , \*\*\*\* $p < 0.001$ .

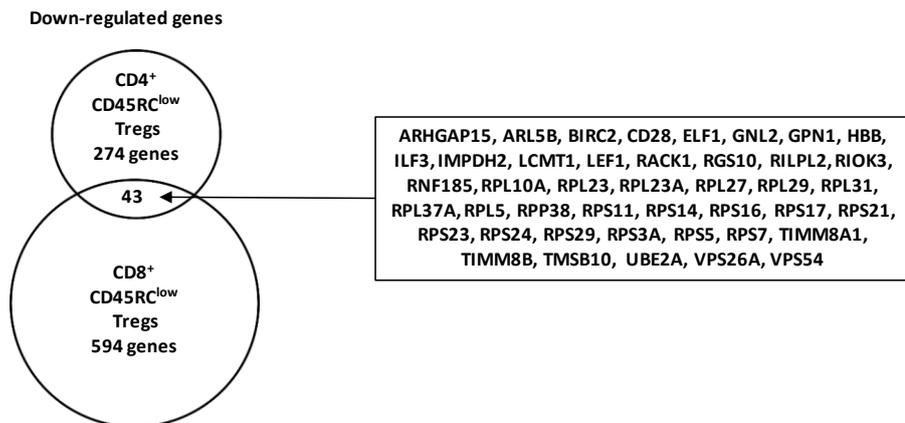
Picarda, Bézine, Boucault et al., Suppl. Figure 5



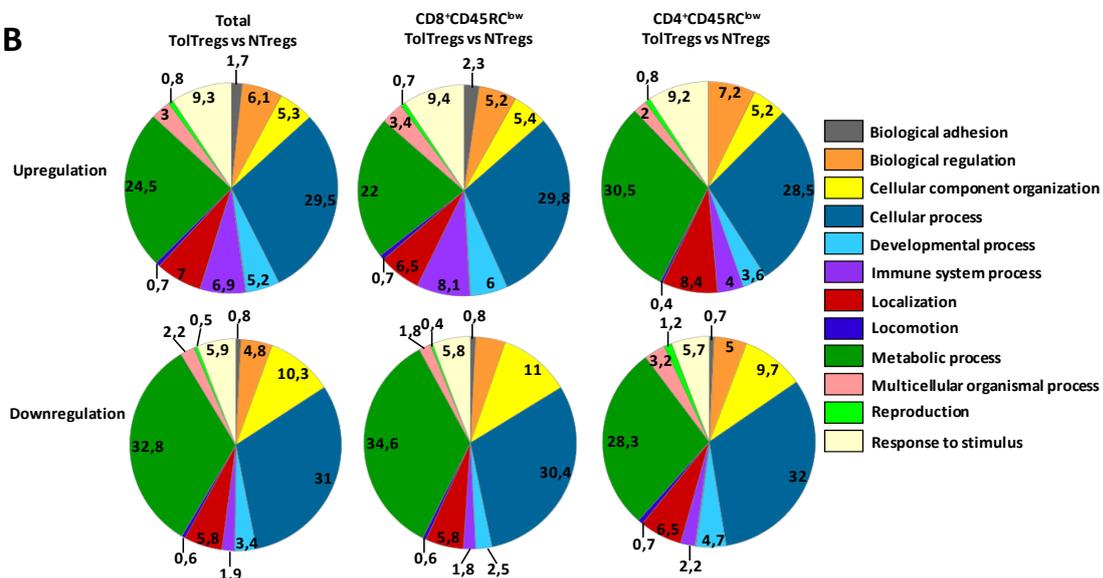
**Supplementary Fig. 5** CD8<sup>+</sup> and CD4<sup>+</sup>CD45RC<sup>low/-</sup> T cells were assessed for suppressive activity in ex vivo culture assays and for in vivo adoptive transfers. **(A)** Gating strategy and purity of FACS Aria cell sorting of B cells and CD4<sup>+</sup>CD45RC<sup>low</sup> and CD8<sup>+</sup>CD45RC<sup>low</sup> T cells from anti-CD45RC treated rats. Purity was greater than 95%. Representative experiment. **(B)** Representative histograms of 3 experiment of suppressive assay where CFSE-labeled

LeW.1A CD4<sup>+</sup>CD25<sup>-</sup> T cells were stimulated with allogeneic LEW.1W pDCs in absence (filled light grey) or presence of CD8<sup>+</sup>CD45RC<sup>low/-</sup> (left) or CD4<sup>+</sup>CD45RC<sup>low/-</sup> (right) Tregs from LEW.1A anti-CD45RC-treated long-term tolerant recipients (black line) or naive rats (grey line) at ratio effector : suppressor 1:1. Proliferation was analyzed by gating on DAPI CPD450<sup>-</sup>CD4<sup>+</sup>T cells after 6 days of culture.

**A**

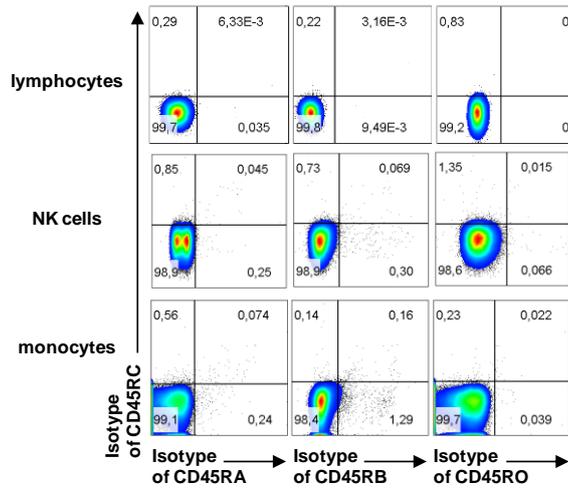


**B**

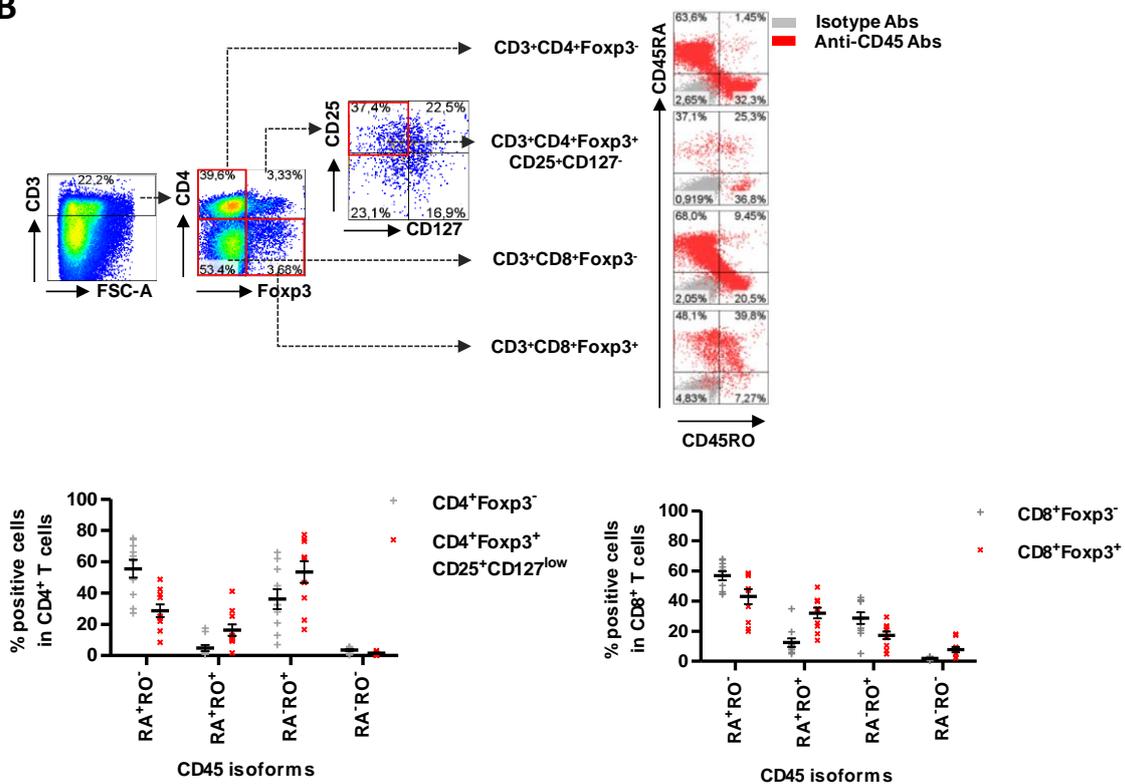


**Supplementary Fig. 6 RNA sequencing of Tregs from anti-CD45RC treated tolerant recipient as compared to natural Tregs. (A)** Venn diagrams were drawn based on downregulated genes from RNA-seq data sets. Circles depict unique and shared genes downregulated by CD4<sup>+</sup> and CD8<sup>+</sup> CD45RC<sup>low</sup> Tregs from anti-CD45RC MAb treated vs untreated rats. The squares represent a subset of the list of downregulated genes corresponding to each part of the circles. **(B)** PANTHER gene ontology analysis of functional annotations associated with upregulated or downregulated genes unique or shared by CD4<sup>+</sup> and CD8<sup>+</sup> CD45RC<sup>low</sup> Tregs from CD45RC MAb treated vs untreated rats. Each color represents different functions. Results are expressed in percentage of modified genes expression.

**A**



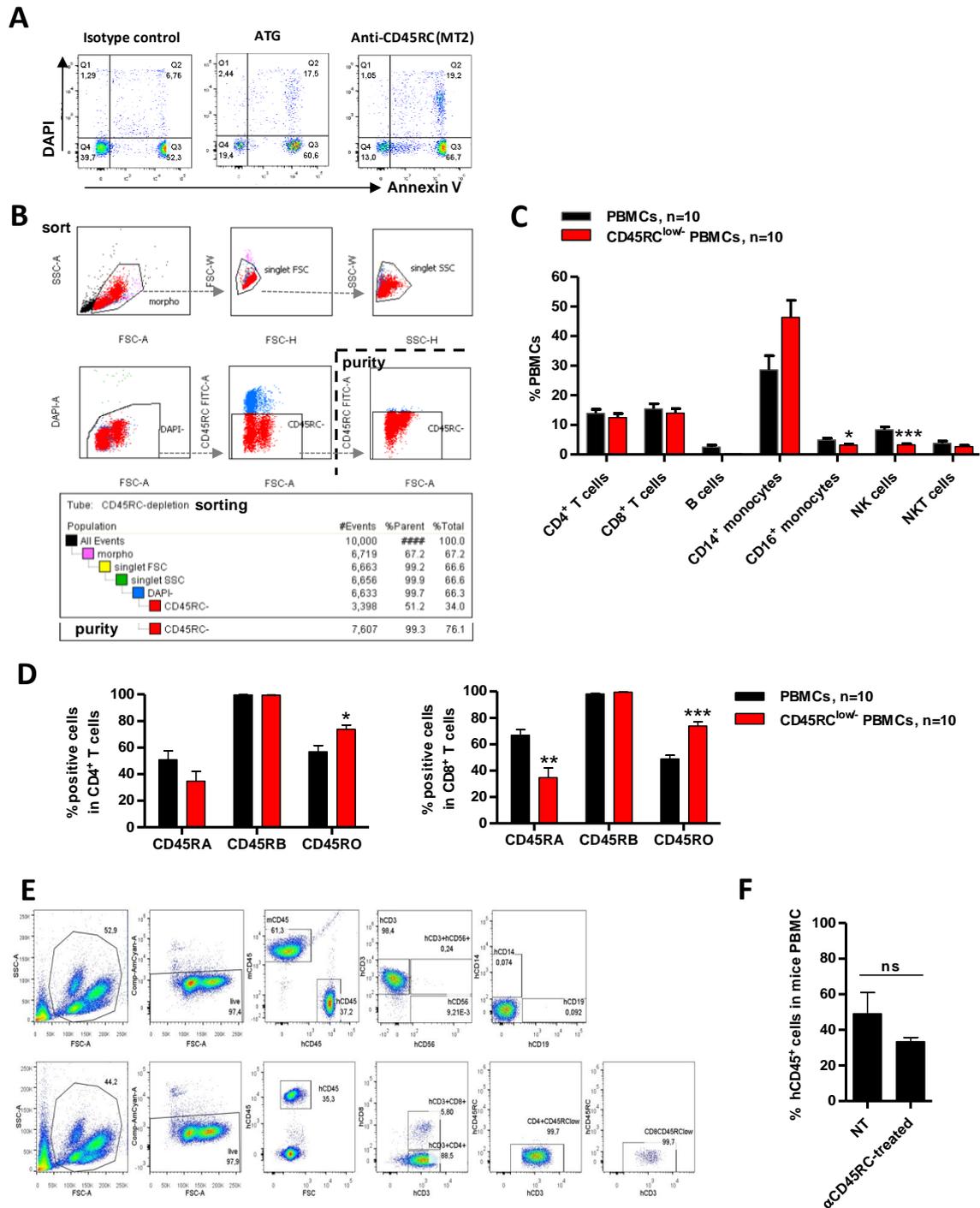
**B**



**Supplementary Fig. 7 Flow cytometry staining with CD45 isoforms isotype matched controls MAbs. (A)** Lymphocytes, NK cells and monocytes were analyzed for isotype control staining by flow cytometry. Representative experiment of 10 individuals. **(B)** Co-expression pattern analysis of CD45RA and CD45RO on CD4<sup>+</sup> and CD8<sup>+</sup> subsets among T cells. A representative dot plot analysis of 10 healthy volunteers and mean $\pm$ -SEM of

CD45RA and CD45RO positive cells among CD4<sup>+</sup>CD25<sup>+</sup>CD127<sup>-</sup> Tregs, CD8<sup>+</sup>Foxp3<sup>+</sup> Tregs, Foxp3<sup>-</sup>CD4<sup>+</sup> and Foxp3<sup>-</sup>CD8<sup>+</sup> T cells.

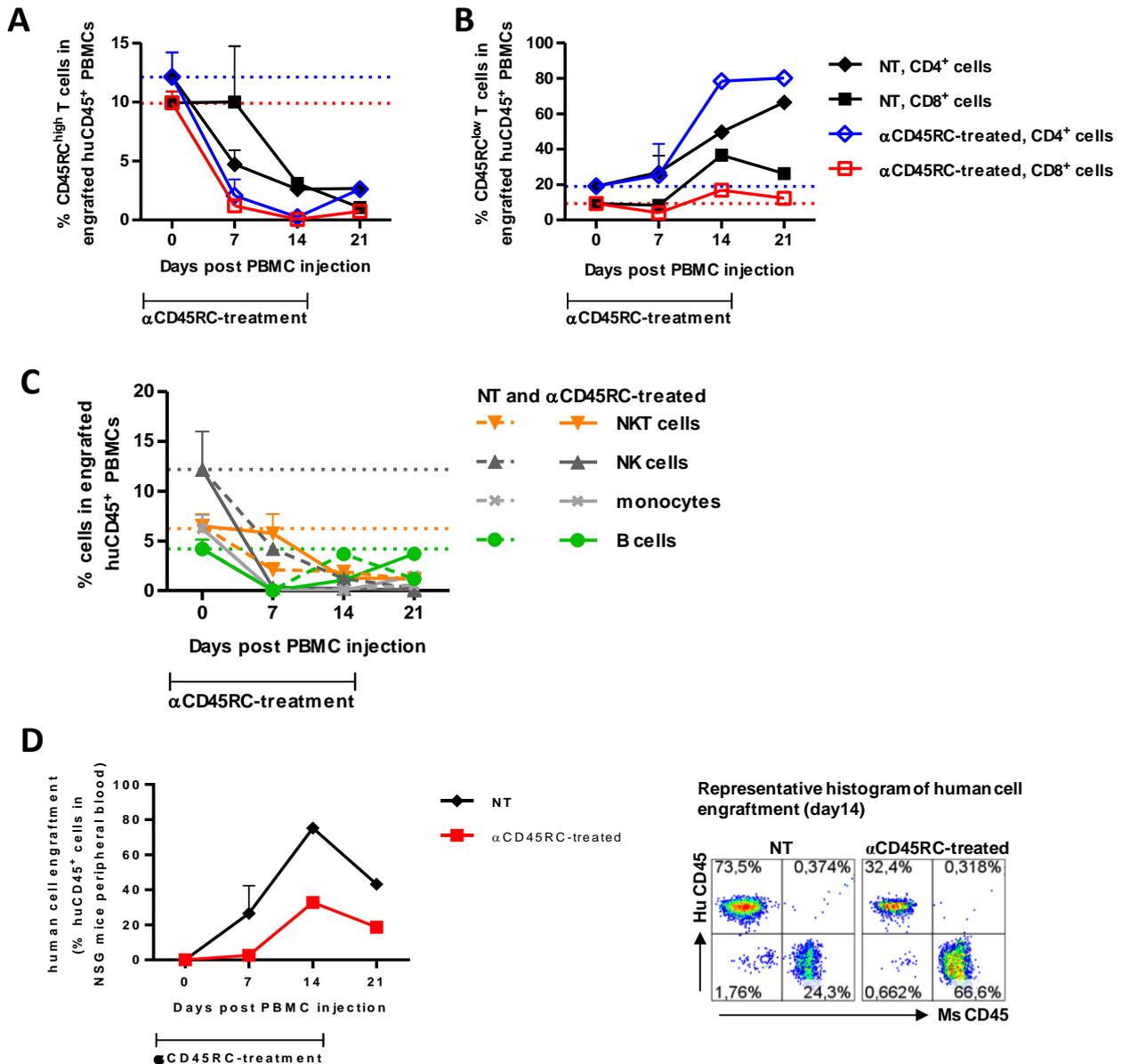
Picarda, Bézie, Boucault et al., Suppl. Figure 8



**Supplementary Fig. 8. In vitro and in vivo assessment of anti-CD45RC MAb treatment on human cells. (A).** PBMCs from healthy human volunteers were incubated with anti-CD45RC (MT2 clone), isotype control (mouse IgG1), or ATG and analyzed for DAPI/Annexin V staining in CD3<sup>+</sup> cells. Representative experiment of 3. **(B)** Gating strategy and purity of FACS Aria cell sorting of CD45RC negative PBMCs from healthy volunteers

for adoptive transfer. Purity was greater than 99%. Representative experiment of 6. **Proportion of immune cells remaining following cell sorting among total PBMCs (C) and proportion of isoforms of CD45 (D) in 10 healthy volunteers. (E)** Representative staining of 3 mice. **(F)** Long surviving mice injected with CD45RC-depleted PBMCs were analyzed for human CD45<sup>+</sup> cell engraftment in mice blood.

Picarda, Bézie, Boucault et al., Suppl. Figure 9



**Supplementary Fig. 9 In vivo assessment of anti-CD45RC MAb treatment on human cells.**

Mice injected with anti-CD45RC MAb or not and human PBMCs were analyzed for the percentage of CD45RC<sup>high</sup> (A) and CD45RC<sup>low</sup> (B) T cells; NKT, NK, B cells or monocytes (C) in engrafted huCD45<sup>+</sup> PBMCs; and for total human CD45<sup>+</sup> cell engraftment in mice blood (D). Results are expressed as mean $\pm$ SEM of cell proportion in human PBMCs (A-C) or mice blood cells (D).