

1 Supplemental material

2 *Kidney tissue lysis, protein isolation, and immunoblotting*

3 The following protocol was adapted from Yang et al. ¹. Kidney tissues were snap frozen in liquid
4 nitrogen and stored at -80°C immediately on the mice sacrifice day. Tissue samples were
5 homogenized manually in RIPA lysis buffer containing phosphatase inhibitor (Calbiochem,
6 524625), protease inhibitor (Sigma, P1860), and SDS 10%. The protein concentration was
7 measured by BCA assay (Pierce, 23225). Protein (20 ug) from lysates was loaded on a 10%, 12%
8 and 15% polyacrylamide gel and separated by SDS-PAGE. Proteins were transferred to
9 nitrocellulose membranes (Bio-Rad, 162-0115), blocked in 5% skimmed milk in PBS containing
10 0.1% Tween (PBST), and probed overnight at 4°C with the following primary antibodies: rabbit
11 anti-LC3 at 1:1000 (Novus, NB600-1384); rabbit anti-SQSTM1/p62 at 1:2000 (Abcam, ab91526);
12 goat anti-TIM-1/KIM-1/HAVCR at 1:1000 (Novus Biologicals, AF1817); rabbit anti- α 1 β tubulin at
13 1:2000 (Proteintech, 11-224-1-AP); rabbit anti-GFP at 1:1000 (Cell signaling, 2956S). Following
14 incubation with primary antibodies, the blots were washed, probed with the Anti-Rabbit IgG HRP
15 at 1:8000 (Abcam, ab7074) or Anti-Goat IgG HRP at 1:10000 (R&D system, HAF017) for 30 min at
16 room temperature, and then visualized using the western ECL kit (Bio-Rad, 1705062).
17 Quantification of western blot bands was performed using Image Lab software (Bio-Rad).

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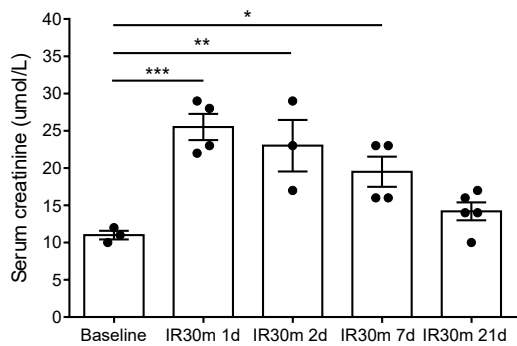
19 **Supplementary Figure Legends**

20 **Figure S1. Renal ischemia-reperfusion injury induces kidney dysfunction and tubular damage.**

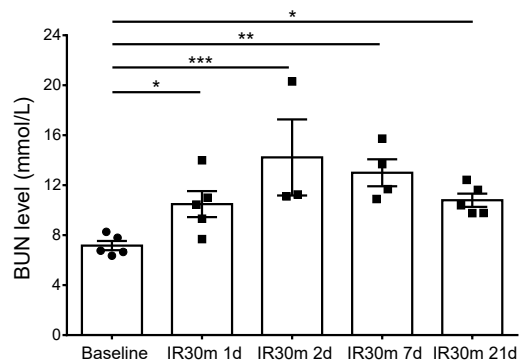
21 **(A)** Bar graph of serum creatinine levels in GFP-LC3 mice post-IRI (n=3-5). **(B)** Bar graph of blood
22 urea nitrogen levels in GFP-LC3 mice post-IRI (n=3-5). **(C)** Bar graph of mean tubular injury scores
23 in H&E-stained kidney sections (n=4-5). **(D)** Bar graph of mean number of PTC congestion in H&E-
24 stained kidney sections (n=4-5). Quantification was performed with samples at baseline, 1, 2, 7,
25 and 21 days post-IRI. For each kidney, ten randomly selected high-power fields (200× original
26 magnification) were evaluated, consisting of five fields from the cortex and five from the
27 corticomedullary junction. Values are mean ± SEM. P values obtained by one-way ANOVA and the
28 Bonferroni post hoc test. *p<0.05, **p<0.01, ***p<0.001, and ****p<0.0001, compared between
29 baseline and each time points.

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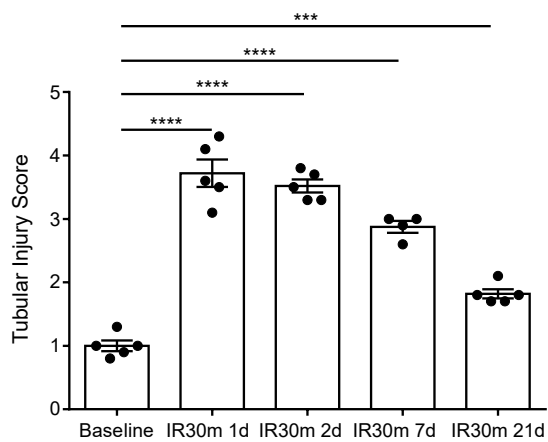
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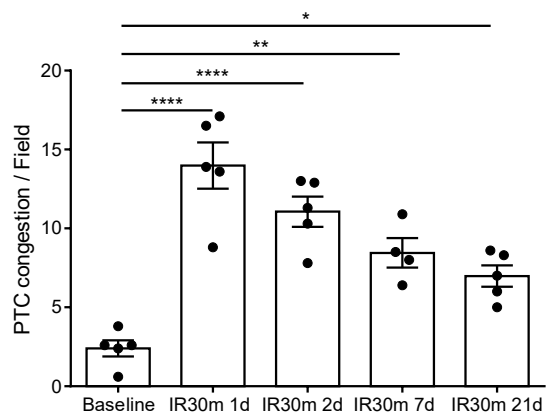
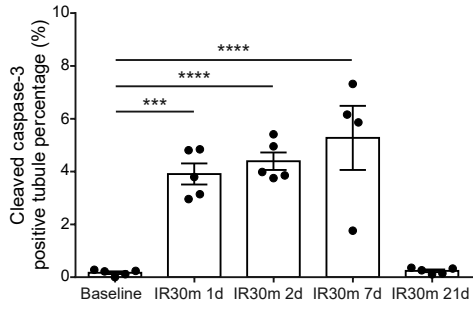
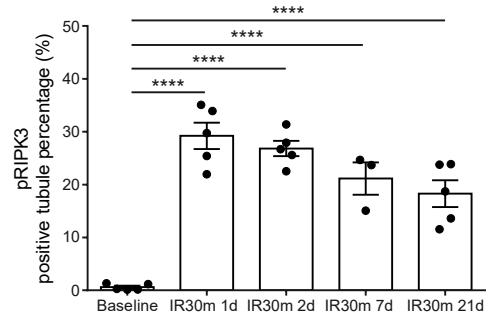


Figure S2. Renal ischemia-reperfusion injury induces unique patterns of necroptosis and caspase-3-dependent apoptosis in tubules and peritubular capillaries. (A) Bar graph of cleaved caspase-3 IHC quantification in renal tubules (n=4-5). (B) Bar graph of pRIPK3 IHC quantification in renal tubules (n=3-5). (C) Bar graph of cleaved caspase-3 IHC quantification in renal PTC (n=4-5). (D) Bar graph of pRIPK3 IHC quantification in renal PTC (n=3-5). Quantification was performed with samples at baseline and 1, 2, 7, and 21 days post-IRI. For each kidney, ten randomly selected high-power fields (200× original magnification) were evaluated, consisting of five fields from the cortex and five from the corticomedullary junction. Values are mean ± standard error of the means (SEM). P values obtained by one-way ANOVA and the Bonferroni post hoc test. **p<0.01, ***p<0.001, and ****p<0.0001, compared between baseline and each time point.

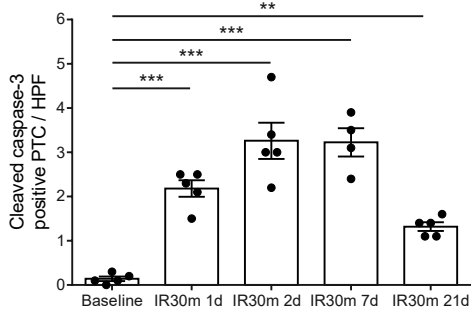
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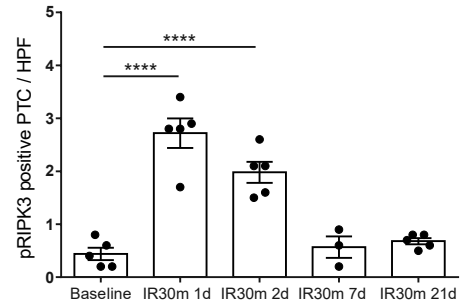
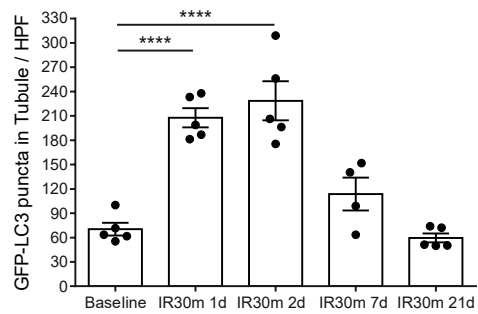


Figure S3. Autophagy dynamics in tubules and peritubular capillaries following renal ischemia-reperfusion injury. (A) Bar graph of GFP-LC3 puncta quantification in renal tubules (n=5). (B) Bar graph of SQSTM1/p62 IHC quantification in renal tubules (n=4-5). (C) Bar graph of autophagic puncta quantification in renal PTC (n=3-5). (D) Bar graph of SQSTM1/p62 IHC quantification in renal PTC (n=4-5). All quantification was performed with samples at baseline, 1, 2, 7, and 21 days post-IRI. For each kidney, ten randomly selected high-power fields (200× original magnification) were evaluated, consisting of five fields from the cortex and five from the corticomedullary junction. Values are mean ± standard error of the means (SEM). P values obtained by one-way ANOVA and the Bonferroni post hoc test. **p<0.01, ***p<0.001, and ****p<0.0001, compared between baseline and each time point.

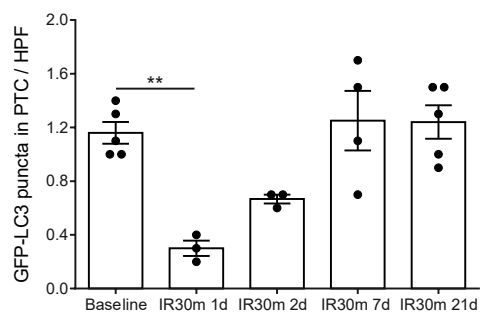
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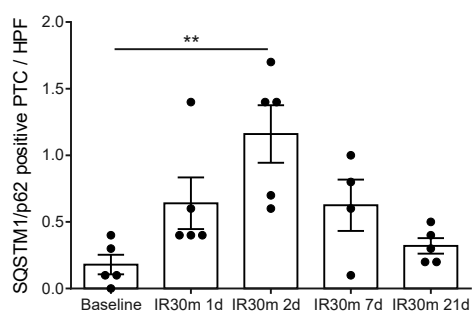
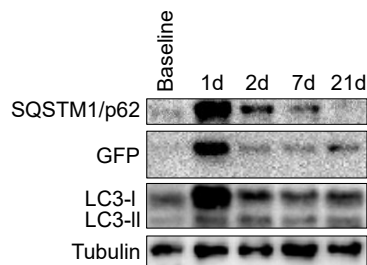
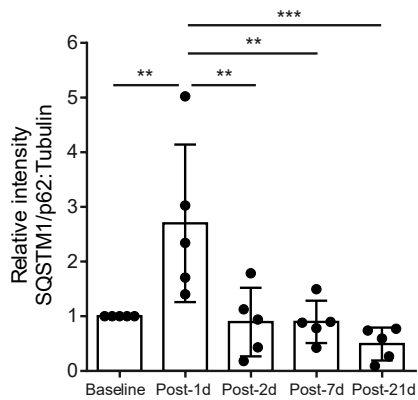


Figure S4. Western blot of autophagy-related proteins in GFP-LC3 mice kidney following renal ischemia-reperfusion injury. (A) Representative western blots of autophagy markers SQSTM1/p62, LC3-I, LC3-II, and GFP fragment and tubulin in GFP-LC3 mouse kidney. (B)–(D) Western blot quantification of SQSTM1/p62 (B), GFP fragment (C), LC3-II/LC3-I (D). Quantification was compared between 1-day post-IRI and each time point (n=5). Values are mean $\pm$ standard deviation (SD). P values obtained by one-way ANOVA and the Bonferroni post hoc test. *p<0.05, **p<0.01, and ***p<0.001 compared between 1-day post-IRI and each time point.

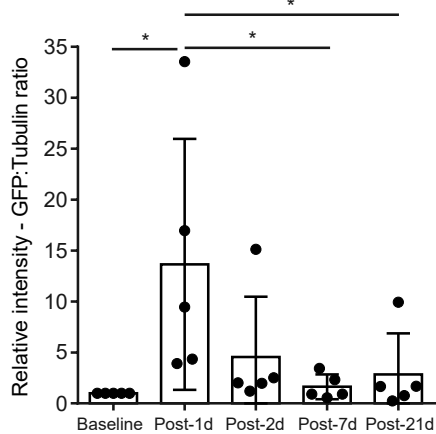
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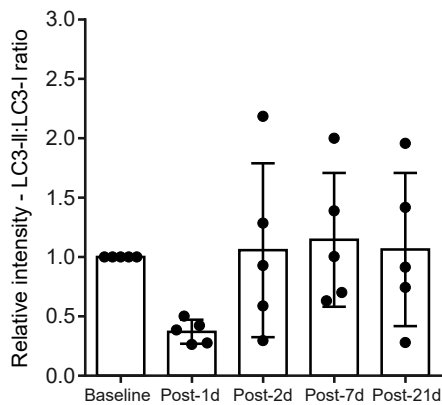


Figure S5. Autophagy inhibition enhances kidney dysfunction following renal ischemia-reperfusion injury. Serum creatinine levels up to 21 days with PBS or CHQ post renal-IRI (n=5-8). Quantification was performed by comparing the PBS and CHQ groups at baseline, 1, 2, 7, and 21 days post-IRI. Values are mean $\pm$ standard error of the means (SEM). P values obtained by one-way ANOVA and the Bonferroni post hoc test. ***p<0.001.

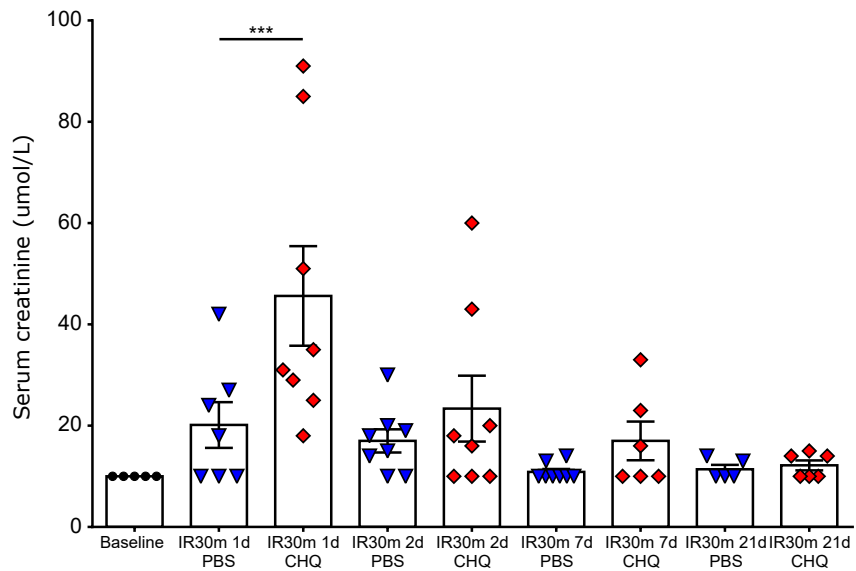
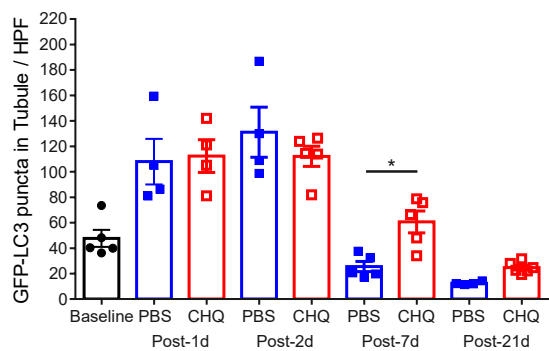


Figure S6. Autophagy inhibition enhances autophagosome formation following renal ischemia-reperfusion injury. (A) Bar graph of GFP-LC3 puncta quantification in renal tubules from PBS and CHQ-injected mice (n=4-8). (B) Bar graph of autophagic puncta quantification in renal PTC from PBS and CHQ-injected mice (n=4-8). Quantification was performed with samples at baseline and 1, 2, 7, and 21 days post-IRI. For each kidney, ten randomly selected high-power fields (200× original magnification) were evaluated, consisting of five fields from the cortex and five from the corticomedullary junction. Values are mean ± standard error of the means (SEM). P values obtained by one-way ANOVA and the Bonferroni post hoc test. *p<0.05 and ****p<0.0001, compared between PBS and CHQ at the same time points.

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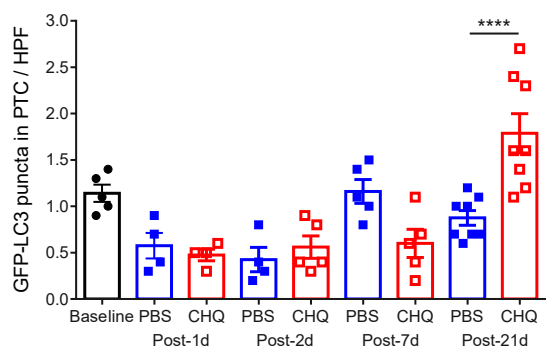
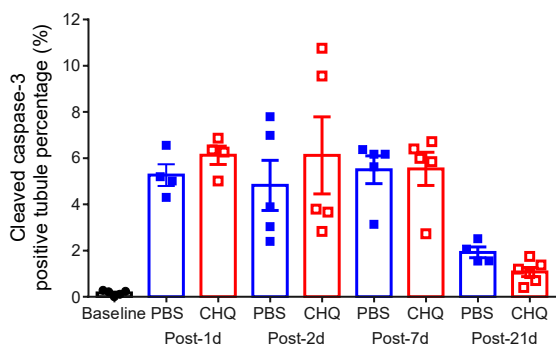
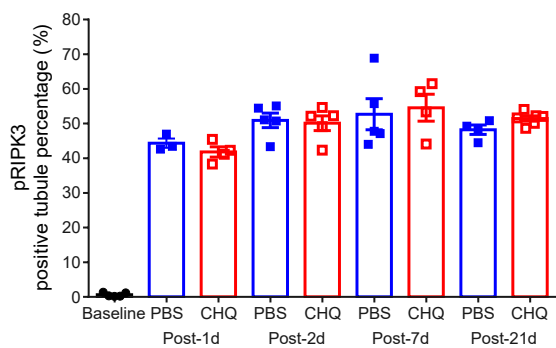


Figure S7. Autophagy inhibition enhances peritubular capillary apoptosis but not tubular necroptosis following renal ischemia-reperfusion injury. (A) Bar graph of cleaved caspase-3 IHC quantification in renal tubules from PBS and CHQ-injected mice (n=4-5). (B) Bar graph of pRIPK3 IHC quantification in renal tubules from PBS and CHQ-injected mice (n=4-5). (C) Bar graph of cleaved caspase-3 IHC quantification in renal PTC from PBS and CHQ-injected mice (n=4-5). (D) Bar graph of pRIPK3 IHC quantification in renal PTC from PBS and CHQ-injected mice (n=4-5). Quantifications was performed with samples at baseline and 1, 2, 7, and 21 days post-IRI. For each kidney, ten randomly selected high-power fields (200× original magnification) were evaluated, consisting of five fields from the cortex and five from the corticomedullary junction. Values are mean ± standard error of the means (SEM). P values obtained by one-way ANOVA and the Bonferroni post hoc test. *p<0.05 and **p<0.01, compared between PBS and CHQ at the same time points.

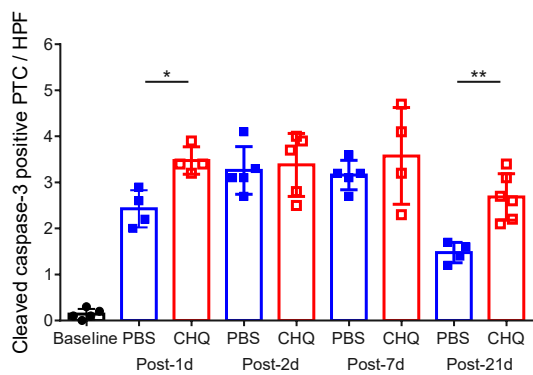
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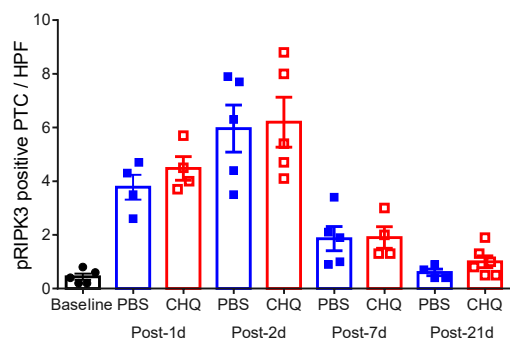
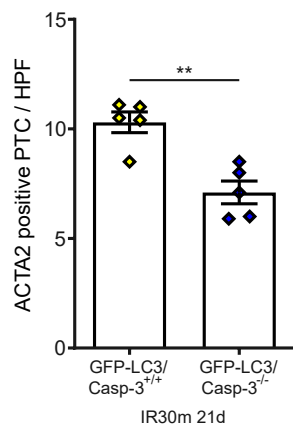
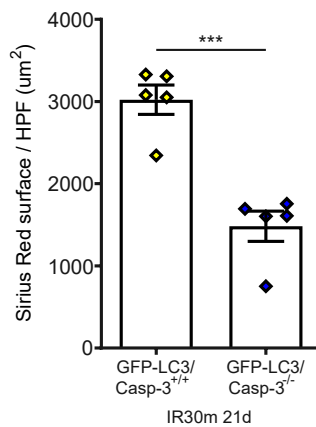


Figure S8. Caspase-3 deficiency reduces fibrosis and improves microvascular density following renal ischemia-reperfusion injury. (A) Quantification of alpha smooth muscle actin (ACTA2) positive peritubular capillary (PTC) immunohistochemistry in the kidney from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} at 21 days post-IRI (n=5). (B) Quantification of Sirius Red staining in the kidney from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} at 21 days post-IRI (n=5). (C) Quantification of PLVAP IHC in the kidney from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} at 21 days post-IRI (n=5). For each kidney, ten randomly selected high-power fields (200× original magnification) were evaluated, consisting of five fields from the cortex and five from the corticomedullary junction. Values are mean ± standard error of the means (SEM). P values obtained by unpaired t- test. *p<0.05 and **p<0.01, ***p<0.001 compared between GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} at 21 days post-IRI.

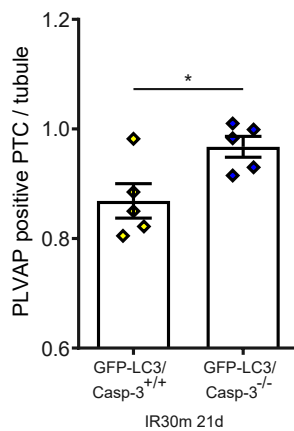
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101 **Figure S9. Autophagy disruption and caspase-3 deficiency do not affect renal dysfunction and**
102 **kidney injury following renal ischemia-reperfusion injury. (A)** Serum creatinine levels at baseline,
103 2 days, and 21 days with PBS or CHQ post renal-IRI (n=4-5). **(B)** Representative immunoblots of
104 kidney lysate following renal IRI. **(C)** Quantification of immunoblots (n=5). Values are mean $\pm$
105 standard error of the means (SEM). P values obtained by one-way ANOVA and the Bonferroni post
106 hoc test. *p<0.05 and **p<0.01.

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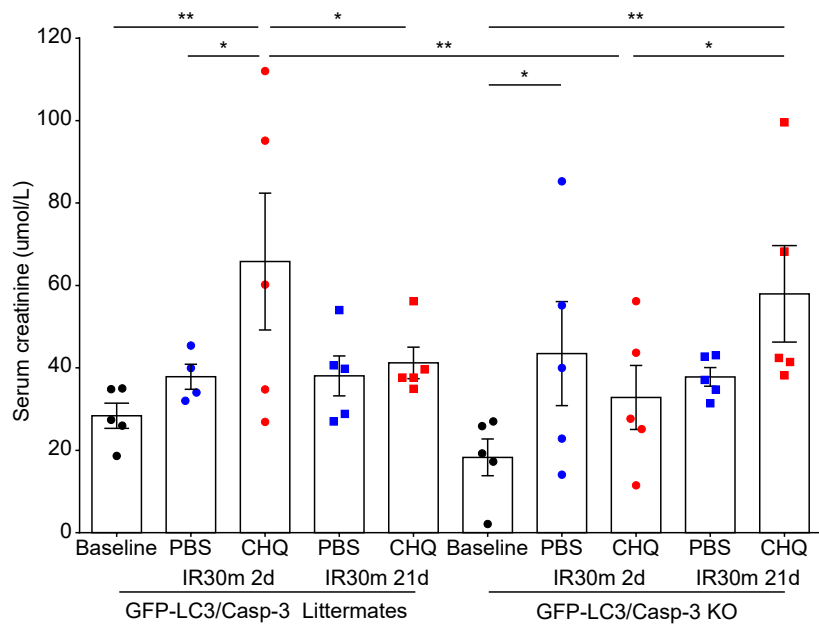
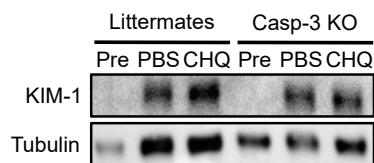
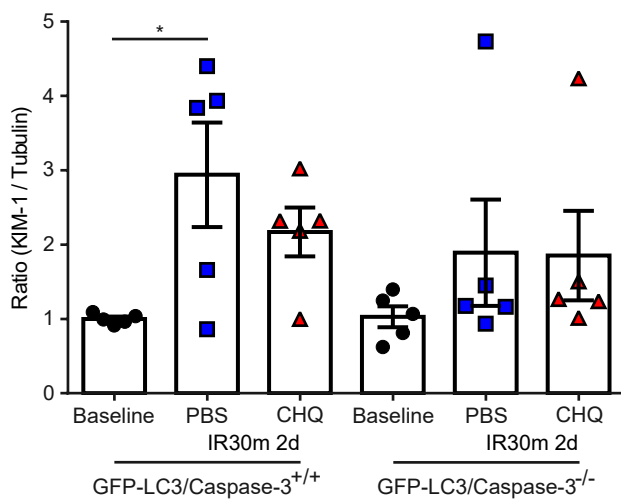
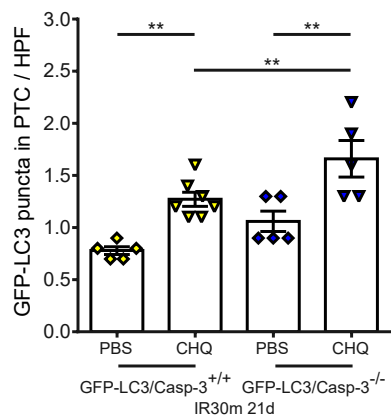
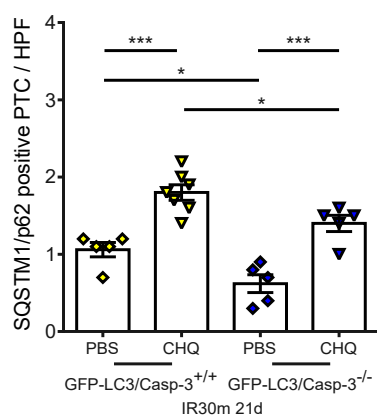
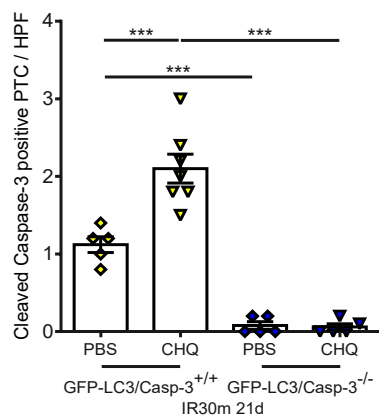
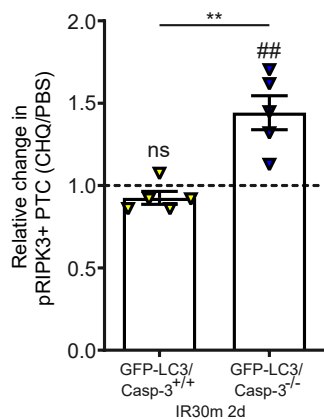
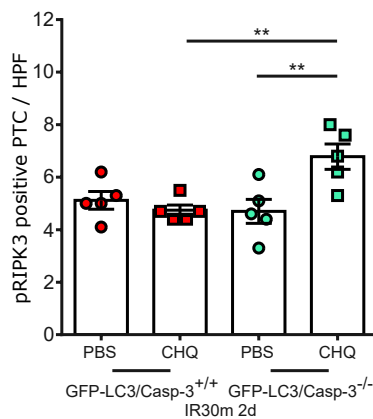
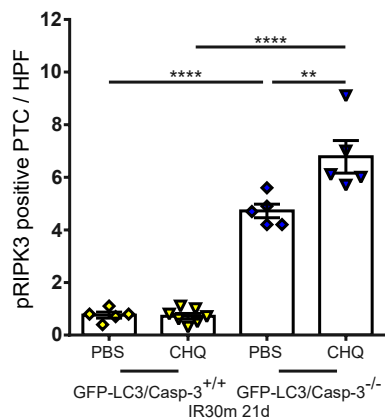
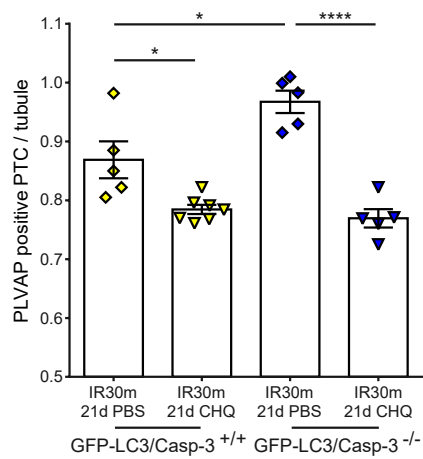
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Figure S10. Autophagy disruption and caspase-3 deficiency worsen microvascular rarefaction, fibrosis, and leukocyte infiltration following renal ischemia-reperfusion injury. (A) Bar graph of autophagic puncta quantification in renal PTC at 21 days post-IRI from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} injected with PBS or chloroquine (CHQ) (n=5-7). (B) Bar graph of SQSTM1/p62 quantification in renal PTC at 21 days post-IRI from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} injected with PBS or chloroquine (CHQ) (n=5-7). (C) Bar graph of cleaved caspase-3 IHC quantification in PTC at 21 days post-IRI from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} injected with PBS or chloroquine (CHQ) (n=5-7). (D) Relative change in phosphorylated RIPK3 (pRIPK3) IHC in PTC at 2 days post-IRI (n=5). (E) Bar graph of pRIPK3 IHC quantification in PTC at 2 days post-IRI from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} injected with PBS or chloroquine (CHQ) (n=4-5). (F) Bar graph of pRIPK3 IHC quantification in PTC at 21 days post-IRI from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} injected with PBS or chloroquine (CHQ) (n=5-7). (G) Bar graph of PLVAP quantification in renal PTC at 21 days post-IRI from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} injected with PBS or chloroquine (CHQ) (n=5-7). (H) Bar graph of ACTA2 quantification in renal PTC at 21 days post-IRI from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} injected with PBS or chloroquine (CHQ) (n=5-7). (I) Relative change in Sirius red staining in kidney at 21 days post-IRI from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} injected with PBS or chloroquine (CHQ) (n=5-7). (J) Bar graph of Sirius red staining in kidney at 21 days post-IRI from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} injected with PBS or chloroquine (CHQ) (n=5-7). (K) Bar graph of CD45-positive area to total tissue area in kidney at 21 days post-IRI from GFP-LC3/Caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} injected with PBS or chloroquine (CHQ) (n=5-7). Randomly selected ten high-power fields images (Original

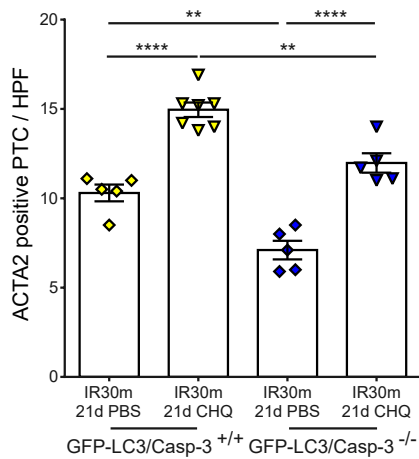
magnification X200) were evaluated from GFP-LC3/caspase-3^{-/-} and GFP-LC3/Caspase-3^{+/+} kidneys from mice injected with PBS or CHQ at 2 and 21 days post-IRI. For each kidney, ten randomly selected high-power fields (200× original magnification) were evaluated, consisting of five fields from the cortex and five from the corticomedullary junction. Values are mean ± SEM. P values obtained by one-way ANOVA and the Bonferroni post hoc test. *p<0.05, **p<0.01, ***p<0.001, and ****p<0.0001.

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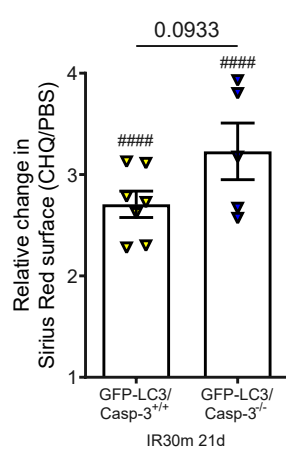
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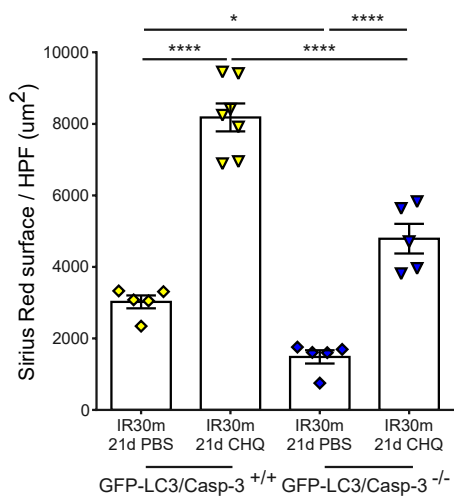
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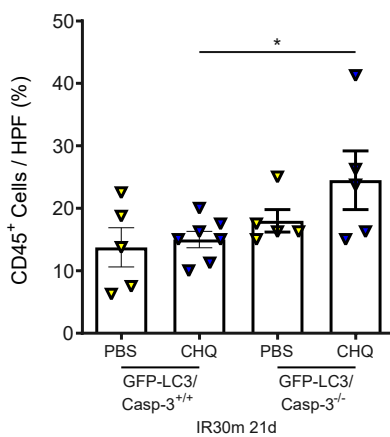
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138 **Supplementary References**

- 139 1. Yang B, Lan S, Dieudé M, et al. Caspase-3 Is a Pivotal Regulator of Microvascular
140 Rarefaction and Renal Fibrosis after Ischemia-Reperfusion Injury. *J Am Soc Nephrol*. Jul
141 2018;29(7):1900-1916. doi:10.1681/asn.2017050581

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