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Insulin/insulin growth factor signaling is a conserved pathway that regulates lifespan. However, long-lived loss-of-function mutants often produce insulin resistance, slow growth, and impair reproduction. Recently, a gain-of-function mutation in the kinase insert domain (KID) of the *Drosophila* insulin/IGF receptor was seen to dominantly extend lifespan without impairing insulin sensitivity, growth, or reproduction. This substitution occurs within residues conserved in mammalian insulin receptor (IR) and insulin growth factor-1 receptor (IGF-1R). We produced 2 knock-in mouse strains that carry the homologous KID Arg/Cys substitution in murine IR or IGF-1R, and we replicated these genotypes in human cells. Cells with heterodimer receptors of IR or IGF-1R induce receptor phosphorylation and phospho-Akt when stimulated with insulin or IGF. Heterodimer receptors of IR fully induce pERK, but ERK was less phosphorylated in cells with IGF-1R heterodimers. Adults with a single KID allele (producing heterodimer receptors) have normal growth and glucose regulation. At 4 months, these mice variably display hormonal markers that associate with successful aging counteraction, including elevated adiponectin and FGF21, as well as reduced leptin and IGF-1. Livers of IGF-1R females show decreased transcriptome-based biological age, which may point toward delayed aging and warrants an actual lifespan experiment. These data suggest that KID mutants may slow mammalian aging while they avoid the complications of insulin resistance.

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Mutation in IR or IGF1R produces features of long-lived mice while maintaining metabolic health

Ulalume Hernández-Arciga,¹ Jun Kyoung Kim,^{1,2} Jacob L. Fisher,^{1,2} Alexander Tyshkovskiy,^{3,4} Alibek Moldakozhayev,^{3,5,6,7} Catherine Hall,⁸ Souvik Ghosh,^{1,9} Yashvandhini Govindaraj,^{1,2} Ian J. Sipula,^{10,11} Jake Kastroll,^{10,11} Diana Cooke,¹² Jinping Luo,¹³ Jonathan K. Alder,^{14,15} Stacey J. Sukoff Rizzo,¹ Gene P. Ables,¹² Eunhee Choi,⁸ Vadim N. Gladyshev,^{3,7} Michael J. Jurczak,^{10,11} Marc Tatar,¹⁶ and Andrey A. Parkhitko^{1,11}

¹Aging Institute of UPMC and University of Pittsburgh, Pittsburgh, Pennsylvania, USA. ²Dietrich School of Arts & Sciences, University of Pittsburgh, Pennsylvania, USA. ³Division of Genetics, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, Massachusetts, USA. ⁴Belozersky Institute of Physico-Chemical Biology, Moscow State University, Moscow, Russia. ⁵Department of Neurology and Neurosurgery, McGill University, Montreal, Quebec, Canada. ⁶Metabolic Disorders and Complications Program, and Brain Repair and Integrative Neuroscience Program, Research Institute of the McGill University Health Centre, Montreal, Quebec, Canada. ⁷Broad Institute of MIT and Harvard, Cambridge, Massachusetts, USA. ⁸Department of Pathology and Cell Biology, Vagelos College of Physicians and Surgeons, Columbia University, New York, New York, USA. ⁹Department of Human Genetics, School of Public Health, ¹⁰Center for Metabolism and Mitochondrial Medicine, Department of Medicine, and ¹¹Division of Endocrinology and Metabolism, Department of Medicine, University of Pittsburgh, Pittsburgh, Pennsylvania, USA. ¹²Orentreich Foundation for the Advancement of Science Inc., Cold Spring, New York, USA. ¹³Department of Molecular Biology, Cell Biology & Biochemistry, Brown University, Providence, Rhode Island, USA. ¹⁴Pulmonary, Allergy, Critical Care, and Sleep Medicine, University of Pittsburgh, Pittsburgh, Pennsylvania, USA. ¹⁵Dorothy P. and Richard P. Simmons Center for Interstitial Lung Disease, University of Pittsburgh School of Medicine, Pittsburgh, Pennsylvania, USA. ¹⁶Department of Ecology, Evolution and Organismal Biology, and The Center on the Biology of Aging, Brown University, Providence, Rhode Island, USA.

Authorship note: MT and AAP contributed equally to this work.

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Insulin/insulin growth factor signaling is a conserved pathway that regulates lifespan. However, long-lived loss-of-function mutants often produce insulin resistance, slow growth, and impair reproduction. Recently, a gain-of-function mutation in the kinase insert domain (KID) of the *Drosophila* insulin/IGF receptor was seen to dominantly extend lifespan without impairing insulin sensitivity, growth, or reproduction. This substitution occurs within residues conserved in mammalian insulin receptor (IR) and insulin growth factor-1 receptor (IGF-1R). We produced 2 knock-in mouse strains that carry the homologous KID Arg/Cys substitution in murine IR or IGF-1R, and we replicated these genotypes in human cells. Cells with heterodimer receptors of IR or IGF-1R induce receptor phosphorylation and phospho-Akt when stimulated with insulin or IGF. Heterodimer receptors of IR fully induce pERK, but ERK was less phosphorylated in cells with IGF-1R heterodimers. Adults with a single KID allele (producing heterodimer receptors) have normal growth and glucose regulation. At 4 months, these mice variably display hormonal markers that associate with successful aging counteraction, including elevated adiponectin and FGF21, as well as reduced leptin and IGF-1. Livers of IGF-1R females show decreased transcriptome-based biological age, which may point toward delayed aging and warrants an actual lifespan experiment. These data suggest that KID mutants may slow mammalian aging while they avoid the complications of insulin resistance.

Introduction

Insulin/insulin growth factor signaling (IIS) is a conserved nutrient-sensing pathway that regulates lifespan in invertebrate and murine models (1–4). Loss-of-function mutations in the insulin-like receptor of *C. elegans* (*daf-2*) or *Drosophila* (*dInr*) can increase lifespan as much as 2-fold (5–8). In mice, lifespan can be extended nearly 50% in IGF-1R hemizygotes (+/–) in a sex- and strain-dependent manner (9).

However, longevity mutations in these models also produce insulin resistance, slow growth, and impair reproduction (10). This paradox limits our ability to translate such results to humans: slow aging conferred by repressing insulin/IGF signaling may cause metabolic and growth-related disease.

Mature mammals have 2 cell-surface receptors that can bind insulin and IGF ligands: insulin receptor (IR) and insulin-like growth factor-1 receptor (IGF-1R) (11). The insulin and IGF-1Rs function as dimers to interact with insulin and IGF-1, from which they transduce intracellular signaling and subsequently undergo endocytosis (12, 13). Various murine mutations in these receptors have been characterized for their potential effect on aging and to document their deleterious properties. For example, mice homozygous for a mutation (IR^{P1195L}), ortholog of the canonical *daf-2(e1370)* longevity allele of *C. elegans*, die soon after birth from diabetic ketoacidosis, whereas heterozygous mice have normal lifespan but developed insulin resistance with hyperinsulinemia (14, 15). Nelson et al. (16) found that mice hemizygous for a null mutation in the IR had increased lifespan in males but not in females. These mice have congenital insulin resistance, although this complication did not progress with age any faster than in WT. Disruption of IR in adipose tissue alone extends lifespan by 13% (17) and protects mice from age- and obesity-related insulin insensitivity and glucose intolerance (18). These traits, however, may be secondary to how loss of IR impairs adipose development.

Mutations of the IGF-1R also extend lifespan but, again, with some ambiguity and deleterious properties. Female but not male lifespan is increased in mice in the 129/SvPas background with 1 null allele of *Igf1r* (19). This benefit was less apparent, and sometimes became associated with glucose intolerance, when this *Igf1r* genotype was studied in different genetic backgrounds and in standardized laboratory conditions (9, 20). As an alternative to *Igf1r*-null mutants complemented by a WT allele, Lorenzini et al. (21) studied a hypomorph that reduced ligand *igf1* titer. There was little effect on male survival, while female survival was increased in some test locations but not in others. Neoplasm, body mass, and bone density were reduced in all of these *igf1*-deficient mice.

How insulin and IGF receptors modify aging also depends on when the manipulation is initiated. In worms, depletion of DAF-2 during early adulthood extends lifespan but associates with slowed growth, germline shrinkage, egg retention, and reduced brood size. Survival was extended without these correlated effects when DAF-2 was depleted after reproductive cessation or just in the intestine during early adulthood (22). Similarly, Mao et al. (23) tested if murine aging was slowed by monoclonal antibodies targeting the IGF-1 receptor beginning with 18-month-old mice. This improved female health span and remaining life expectancy, while reducing neoplasms and inflammation. Tissue-limited manipulations of IGF signaling can also produce age-specific effects. Increased cardiomyocyte IGF-1R signaling improves heart function in early life but accelerates cardiac aging and reduces survival in later life. In contrast, low IGF-1R signaling suppresses heart function in young mice but preserves cardiac aging (24).

Drosophila melanogaster, unlike mammals, has a single insulin/IGF-like receptor (*dInr*) and 7 insulin/IGF-like peptide encoding genes (*ilps*). Genetic loss-of-function genotypes of the insulin/IGF signaling system extend lifespan but also slow growth, reduce adult size, impair fecundity and limit the ability of insulin to induce phosphorylation of AKT (pAKT) (5, 8). We recently uncovered a potentially novel mutation in the kinase insert domain (KID) (5, 25) that robustly extends lifespan without these deleterious effects (5). This Arg1466Cys substitution in the KID (denoted *dInr*^{KID} or *dInr*³⁵³) robustly extends lifespan in males and females while the adults are full-sized, highly fecund, and induce pAkt when heterozygous with a WT allele (*dInr*^{KID}/*dInr*^{wt}). Importantly, the arginine residue of the KID in *dInr* is conserved in the KIDs of mammalian IR (mouse IR at Arg1109) and IGF-1R (mouse IGF-1R at Arg1096). Here we ask how an Arg>Cys substitution within the KID of IR and IGF-1R affects young mice. We generated 2 knock-in strains that carry the KID Arg>Cys substitution in either IR (IR^{R1109C}) or IGF-1R (IGF-1R^{R1096C}). We characterize how these substitutions affect growth, neuromuscular function, metabolic health, hormonal profiles, and biological age. In a parallel human cell culture model, we ask how the mammalian KID substitutions affect ligand activation of the receptor and downstream phosphorylation of Akt and ERK. We find that heterozygote mice grow normally and are overall metabolically healthy. To varying degrees, these young mice show nascent endocrine profiles previously associated with slowed aging in several murine models. Based on a transcriptomic clock, livers of IGF-1R heterozygous females have reduced biological age and warrant an actual lifespan experiment. In ligand-stimulated cells, IGF-1R^{R1096C} and IR^{R1109C} form functional heterodimers with a WT protomer that activates pAKT but only partially activates pERK. We conclude that the tyrosine receptor KID domains of murine IR and IGF-1R are potential modulators of aging that avoid complications of receptor loss of function.

Results

Viable offspring with Arg>Cys substitutions in the KID of IR (InsR^{R1109C}) or IGF1R (IGF-1R^{R1096C}). The longevity inducing Arg>Cys substitution within the KID of dInr resides in the Arg-Pro-Glu sequence at the start of the KID (5). In general, KIDs are unstructured and variable domains that connect the N- and C-terminal lobes of receptor tyrosine kinases, including mammalian IR and IGF-1R (Figure 1A) (5, 25). We generated 2 knock-in mouse strains in the C57BL/6J background that carry the homologous KID Arg>Cys substitution in murine IR (InsR^{R1109C}) or IGF-1R (IGF-1R^{R1096C}) (Figure 1B). These mutations are viable as heterozygotes over a WT receptor allele. Parental crosses of WT with heterozygous InsR^{R1109C} produced F₁ progeny as 45.5% homozygote WT and 54.5% heterozygote. Crosses of WT with heterozygous IGF-1R^{R1096C} adults produced 53.1% WT and 46.9% heterozygote offspring. Crosses of heterozygote InsR^{wt}/InsR^{R1109C} parents produced F₁ in proportions of 27.7% WT, 63.8% heterozygote, and 8.5% mutant homozygote. Offspring of 38.9% WT, 52.8% heterozygote, and 8.3% mutant homozygotes were derived from crosses of heterozygote InsR^{wt}/IGF-1R^{R1096C} parents (Figure 1C). We could not generate any InsR^{R1109C}, IGF-1R^{R1096C} double heterozygous pups.

Normal growth of InsR^{R1109C} and IGF-1R^{R1096C} heterozygotes. In *Drosophila*, growth is not retarded in WT/KID heterozygotes (dInR⁺/dInR^{KID}) (5). We therefore assessed how growth through 4 months of age was affected when mice were heterozygous or homozygous for InsR^{R1109C} or IGF-1R^{R1096C} (Figure 2, A–D). Because we derived few F₁ homozygotes, hereafter, we include those observations without statistical analyses. InsR⁺/InsR^{R1109C} heterozygotes gained the same mass as WT siblings, while mutant homozygotes at first gained the same mass as WT but plateaued at a lighter weight (38% decrease in females, 48% decrease in males) (Figure 2E). Estimated growth rate based on a parametric model determined the rates were similar among WT and InsR^{R1109C} heterozygotes (Figure 2, A, B, and E).

Female IGF-1R^{R1096C} heterozygotes accumulated less mass than WT at 4 months (12.7% decrease); male heterozygotes weighed as much as WT (Figure 2, C, D, and J). The mass of IGF-1R^{R1096C} homozygotes was reduced in both sexes (Figure 2, C, D, and J). The growth rate of IGF-1R^{R1096C} heterozygotes equaled that of WT in males but was 0.63-fold reduced in females.

We used NMR to measure how the genotypes affected body composition. InsR^{R1109C} and IGF-1R^{R1096C} heterozygotes had the same percentage lean mass as WT (Figure 2, G and L). Total lean mass was slightly reduced in IGF-1R^{R1096C} heterozygotes of both sexes (Figure 2, F and K). Fat mass (total and percentage) was moderately increased in male InsR^{R1109C} heterozygotes (Figure 2, H and I), while total fat was reduced in female IGF-1R^{R1096C} heterozygotes (Figure 2, M and N). Overall, heterozygotes with an Arg>Cys substitution in the KID of murine IR and IGF1R show few or modest effects on growth.

Normal metabolic rates in heterozygote InsR^{R1109C} and IGF-1R^{R1096C}. We used indirect calorimetry to evaluate how InsR^{R1109C} and IGF-1R^{R1096C} affect whole animal metabolism by measuring energy expenditure (EE), respiratory exchange ratio (RER), total activity, and food uptake in mice at 4 months of age. RER describes the substrates used to generate energy (26–30). The RER of WT heterozygotes with InsR^{R1109C} or IGF-1R^{R1096C} was like that of WT in both sexes (0.8–0.85), indicative of substrates from a mixture of protein, lipid, and carbohydrate (Figure 3, A and B, and Supplemental Figure 1, A–D; supplemental material available online with this article; <https://doi.org/10.1172/jci.insight.189683DS1>). RER was reduced (<0.7) among the available male InsR^{R1109C} homozygotes, indicating these animals used lipids rather than carbohydrates to produce energy as occurs when cells are insulin resistant (31) (Figure 3A and Supplemental Figure 1B). Resting EE (Figure 3, C and D, and Supplemental Figure 1, E–H) total activity (Figure 3, E and F, and Supplemental Figure 1, I–L), and food intake (Figure 3, G and H, and Supplemental Figure 1, M–P) did not differ among WT and InsR^{R1109C} or IGF-1R^{R1096C} heterozygote siblings. Overall, InsR^{R1109C} and IGF-1R^{R1096C} heterozygotes present metabolic profiles indistinguishable from their WT littermates.

Normal glycated hemoglobin in heterozygote InsR^{R1109C} and IGF-1R^{R1096C}. To assess glucose homeostasis in InsR^{R1109C} and IGF-1R^{R1096C} heterozygotes, we measured glucose tolerance, plasma insulin, and glycated hemoglobin. Plasma glucose and glycated hemoglobin were normal in male and female InsR^{R1109C} heterozygotes, although mice within the glucose tolerance test (GTT) had elevated fasting insulin (Figure 4, A–H, and Supplemental Figure 2, A–D). As expected, available homozygotes presented elevated plasma glucose, insulin, and glycated hemoglobin, consistent with a state of insulin resistance (Figure 4, A–H, and Supplemental Figure 2, A–D). Male and female IGF-1R^{R1096C} heterozygotes presented normal glucose tolerance (GTT), glycated hemoglobin, and insulin titer except for fasted females where plasma insulin was reduced relative to WT (Figure 4, I–P, and Supplemental Figure 2, E–H).

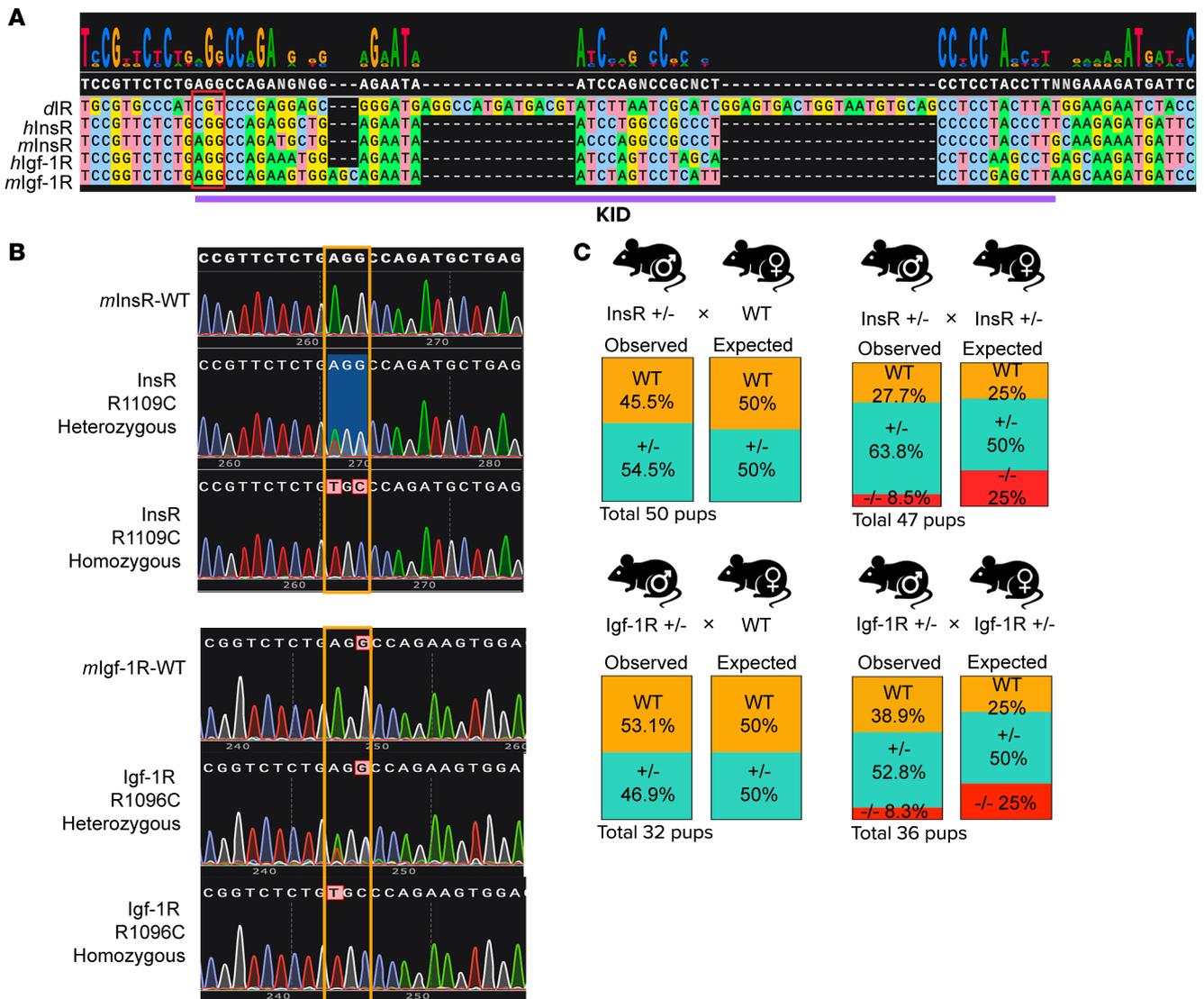


Figure 1. Knock-in mouse strains carrying the homologous dInR^{KID} Arg/Cys substitution in the KID of IR ($\text{InsR}^{\text{R1109C}}$) or IGF1R ($\text{IGF-1R}^{\text{R1096C}}$). (A) Sequence alignment of the KID domain for *Drosophila*, human, and mouse insulin receptor (IR) and insulin growth factor-1 receptor (IGF-1R). (B) DNA sequence chromatogram showing 1 peak for either AGG (WT) or TGC (homozygous) mutant and 2 peaks for heterozygous mutants. (C) Expected versus observed genotype ratios for offspring of WT × heterozygote and heterozygote × heterozygote.

Overall, when heterozygous, the murine KID mutation in $\text{InsR}^{\text{R1109C}}$ results in moderate, compensatory hyperinsulinemia, while a heterozygote with the KID mutation $\text{IGF-1R}^{\text{R1096C}}$ has little effect on energy metabolism and glucose homeostasis.

Improved neuromuscular function in heterozygotes of $\text{InsR}^{\text{R1109C}}$ and $\text{IGF-1R}^{\text{R1096C}}$. We assessed neuromuscular function of young mutant mice through rotarod agility, open field exploratory activity, and spontaneous wheel running. The ability to maintain balance on the accelerating rotarod did not differ between $\text{InsR}^{\text{R1109C}}$ or $\text{IGF-1R}^{\text{R1096C}}$ heterozygotes relative to their age- and sex-matched littermate WT controls (Figure 5, A and B). Motor coordination was intact in homozygous IGF-1R mice relative to heterozygotes and WT, although InsR homozygous appeared to be strongly impaired (based on our limited sample size) (Figure 5, A and B). In the open field test, which measures spontaneous exploratory activity, $\text{IGF-1R}^{\text{R1096C}}$ heterozygotes traveled the same distance as age- and sex-matched WT littermates. Interestingly, female $\text{InsR}^{\text{R1109C}}$ heterozygotes but not males traveled less than age- and sex-matched WT littermates (Figure 5, C and D). Rearing behavior, a typical exploratory behavior in mice that is measured by cumulative vertical activity in the open field, was increased in $\text{IGF-1R}^{\text{R1096C}}$ male heterozygotes but not female $\text{IGF-1R}^{\text{R1096C}}$ heterozygotes relative to age- and sex-matched

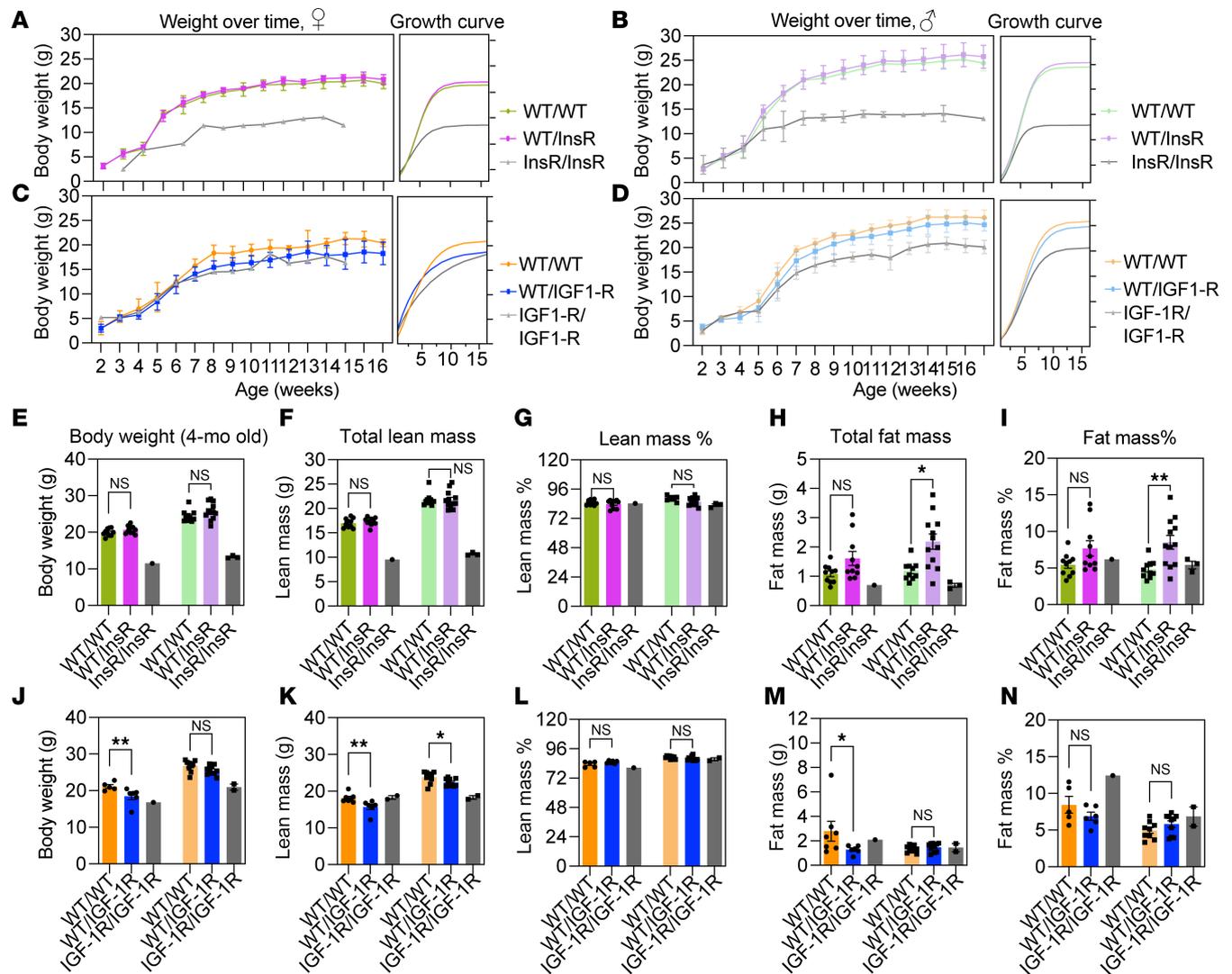


Figure 2. Effect of *InsR*^{R1109C} and *IGF-1R*^{R1096C} substitutions on growth rate. (A–D) Weekly mass (left panel) and logistic growth curves (right panel) in (A) *InsR*^{R1109C} female, (B) *InsR*^{R1109C} male, (C) *IGF-1R*^{R1096C} female, and (D) *IGF-1R*^{R1096C} male mice. (E and J) Female and male body weight in 4-month-old (E) *InsR*^{R1109C} and (J) *IGF-1R*^{R1096C} mice. (F and K) Female and male total lean mass in (F) *InsR*^{R1109C} and (K) *IGF-1R*^{R1096C} mice. (G and L) Female and male percent lean mass normalized to body weight in (G) *InsR*^{R1109C} and (L) *IGF-1R*^{R1096C} mice. (H and M) Female and male total fat mass weight in (H) *InsR*^{R1109C} and (M) *IGF-1R*^{R1096C} mice. (I and N) Female and male percent fat mass normalized to body weight in (I) *InsR*^{R1109C} and (N) *IGF-1R*^{R1096C} mice. WT and heterozygotes were available for statistical comparison (females WT/WT and WT/*InsR* *n* = 10; males WT/WT *n* = 10 and WT/*InsR* *n* = 12; females WT/WT *n* = 5 and WT/*IGF-1R* *n* = 6; males WT/WT *n* = 9 and WT/*IGF-1R* *n* = 10). Homozygote mutants were rare. Data are shown as mean ± SD, Student's *t* test. **P* < 0.05; ***P* < 0.01.

WT littermate controls (Figure 5F). Cumulative vertical activity in the open field was unaltered in *InsR*^{R1109C} heterozygotes relative to age- and sex-matched WT littermate controls (Figure 5E). Thigmotaxis behavior, measured by time spent at the perimeter of the open field, is an indicator of anxiety-like behavior and was similar across all sexes and genotypes (Figure 5, G and H). *InsR*^{R1109C} and *IGF-1R*^{R1096C} homozygotes had reduced cumulative distance traveled and rearing (based on limited observations) (Figure 5, C–F). Voluntary home cage wheel running assesses spontaneous physical activity. As expected, all patients were significantly more active during the dark cycle relative to the light cycle. *InsR*^{R1109C} heterozygote males and females ran the same total distance. Male and female *InsR*^{R1109C} heterozygotes were similar to WT with the exception of time spent running on the second night in males (Figure 5, I, J, M, and N). *IGF-1R*^{R1096C} heterozygotes and WT were similar in both total distance and time spent running (Figure 5, K, L, O, and P).

Overall, young *InsR*^{R1109C} heterozygotes perform slightly better as males in wheel running (males) but less so as females. *IGF-1R*^{R1096C} heterozygote males have modestly elevated activity in open field as measured by rearing behavior.

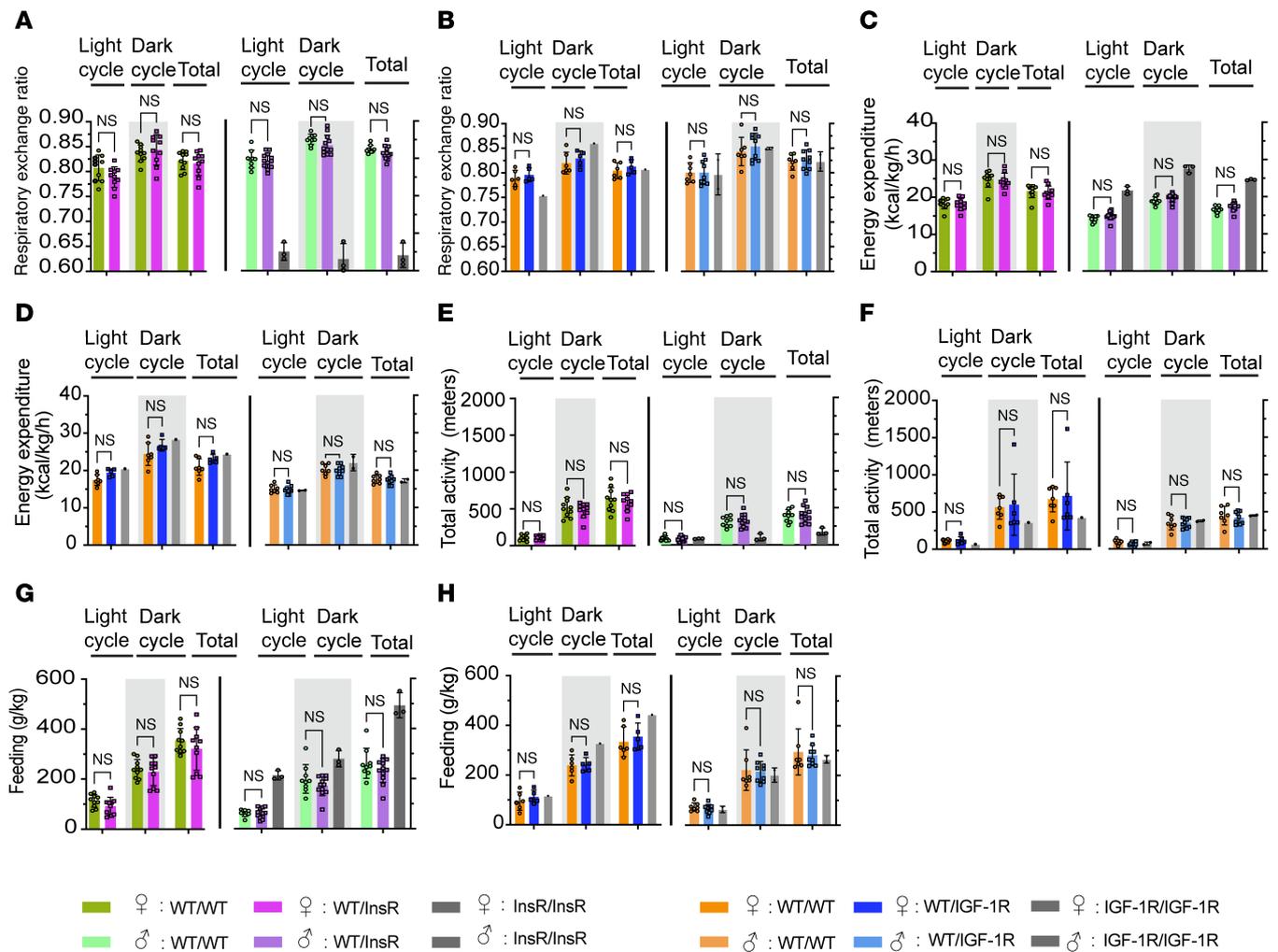


Figure 3. Metabolic rate of *InsR*^{R1109C} and *IGF-1R*^{R1096C} heterozygous mice. (A and B) Respiratory exchange ratio (RER) in (A) *InsR*^{R1109C} and (B) *IGF-1R*^{R1096C} mice, 24-hour average (per kg of lean mass). (C and D) Energy expenditure (EE), 24 hr average (per kg of lean mass) in (C) *InsR*^{R1109C} and (D) *IGF-1R*^{R1096C}. (E and F) Total activity 24-hour average in (E) *InsR*^{R1109C} and (F) *IGF-1R*^{R1096C}. (G and H) Feeding, 24-hour average (per kg of lean mass) in (G) *InsR*^{R1109C} and (H) *IGF-1R*^{R1096C}. WT and heterozygous mice were available for statistical comparison (females WT/WT and WT/*InsR* $n = 10$; males WT/WT $n = 10$ and WT/*InsR* $n = 12$; females WT/WT $n = 5$ and WT/*IGF-1R* $n = 6$; males WT/WT $n = 9$ and WT/*IGF-1R* $n = 10$). Homozygous mutant mice were rare. Data are shown as mean \pm SD, Student's t test.

*Aging-associated hormones of *InsR*^{R1109C} and *IGF-1R*^{R1096C} heterozygotes.* We measured plasma titers of FGF21, IGF-1, adiponectin, leptin, and GDF15 to assess how *InsR*^{R1109C} and *IGF-1R*^{R1096C} affect hormones that are sometimes associated with successful aging. We quantified plasma cholesterol and triglycerides to assess metabolic health. In all assays, plasma was collected from 4-month-old mice in the morning after 6 hours fasting.

FGF21 was increased in heterozygote *InsR*^{R1109C} males but decreased in females, while FGF21 titer in *IGF-1R*^{R1096C} heterozygous mice was equal to that of WT (Figure 6, A and B). Adiponectin was increased in both male and female *InsR*^{R1109C} heterozygotes but did not differ between WT and heterozygotes of *IGF-1R*^{R1096C} (Figure 6, C and D). IGF-1 levels were not different among *InsR*^{R1109C} heterozygotes and WTs in either sex, but the hormone was significantly decreased in *IGF-1R*^{R1096C} male heterozygotes (Figure 6, E and F). The satiety hormone leptin was elevated in male *InsR*^{R1109C} heterozygotes, even though these mice have normal weight and food intake. In contrast, leptin was significantly decreased in *IGF-1R*^{R1096C} female heterozygotes but not in males (Figure 6, G and H). The titer of GDF15 typically increases with chronological age and stress in WT mice (32–34). Here, the titer at 4 months was less in *IGF-1R*^{R1096C} heterozygote females than in WT but not in males. GDF15 did not significantly differ between *InsR*^{R1109C} heterozygotes and WT (Figure 6, I and J). Plasma triglycerides were significantly decreased in *IGF-1R*^{R1096C} female heterozygotes but not in males or in *InsR*^{R1109C} heterozygotes (Figure 6, K and L). We found no significant differences in plasma total cholesterol among genotype or sex (Figure 6, M and N).

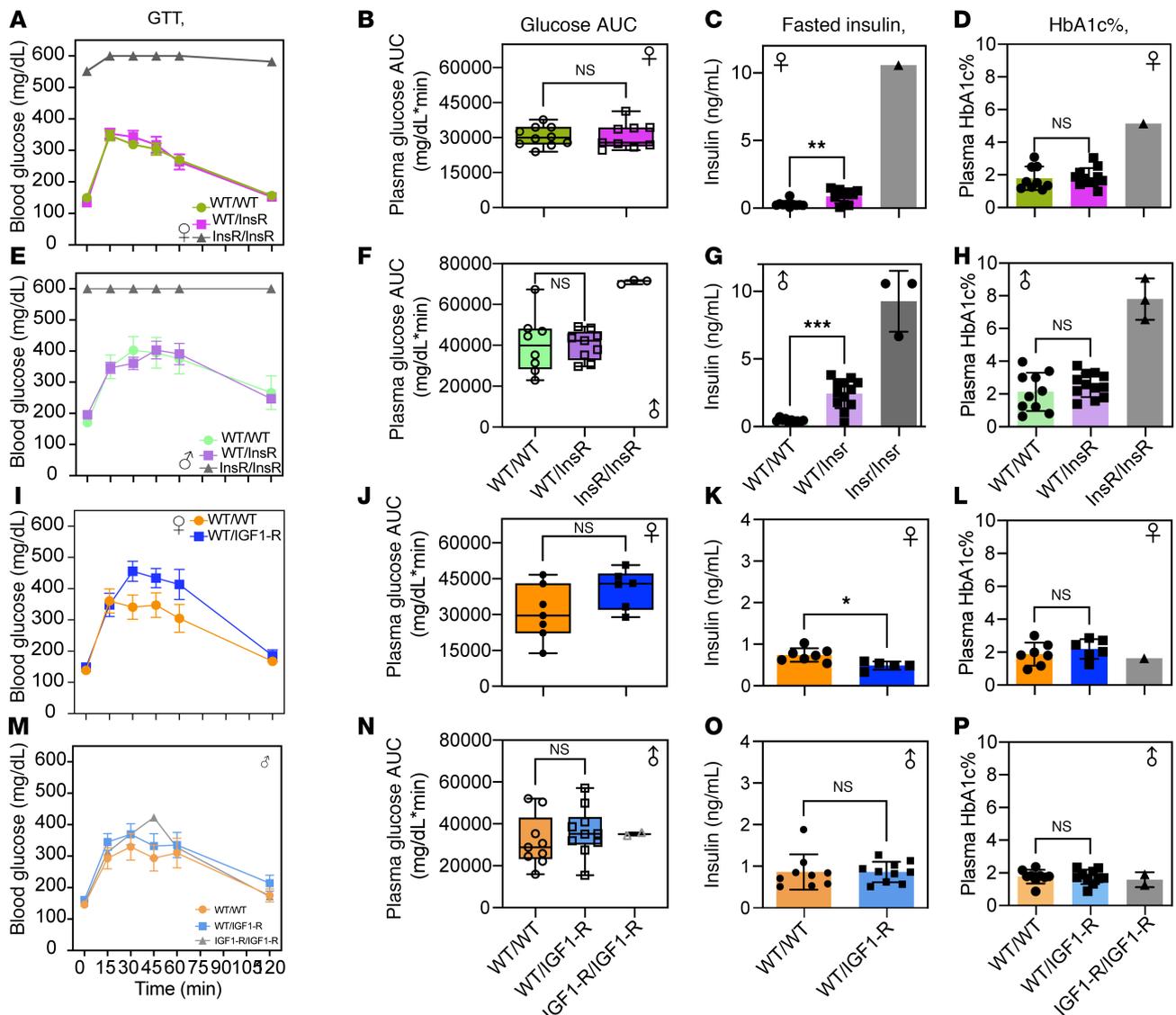


Figure 4. Glycated hemoglobin in *InsR^{R1109C}* and *IGF-1R^{R1096C}* heterozygous mice. (A, E, I, and M) Blood glucose level measured during glucose tolerance test (GTT) using 1.5g/kg glucose dose in (A) female *InsR^{R1109C}*, (E) male *InsR^{R1109C}*, (I) female *IGF-1R^{R1096C}*, and (M) male *IGF-1R^{R1096C}*. (B, F, J, and N) Area under the curve (AUC) for glucose in (B) female *InsR^{R1109C}*, (F) male *InsR^{R1109C}*, (J) female *IGF-1R^{R1096C}*, and (N) male *IGF-1R^{R1096C}*. (C, G, K, and O) Fasted plasma insulin levels in (C) female *InsR^{R1109C}*, (G) male *InsR^{R1109C}*, (K) female *IGF-1R^{R1096C}*, and (O) male *IGF-1R^{R1096C}*. (D, H, L, and P) Glycated hemoglobin A1c percentage in (D) female *InsR^{R1109C}*, (H) male *InsR^{R1109C}*, (L) female *IGF-1R^{R1096C}*, and (P) male *IGF-1R^{R1096C}*. WT and heterozygous were available for statistical comparison (females WT/WT $n = 9-10$ and WT/*InsR* $n = 9-11$; males WT/WT $n = 8-10$ and WT/*InsR* $n = 9-12$; females WT/WT $n = 7$ and WT/*IGF-1R* $n = 6$; males WT/WT $n = 9-10$ and WT/*IGF-1R* $n = 10-11$). Homozygous mutant mice were rare. Data are shown as mean \pm SD, Student's *t* test. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

Overall, young *InsR^{R1109C}* and *IGF-1R^{R1096C}* heterozygous adults present some hormonal changes that are seen in established models of retarded murine aging (elevated FGF21, elevated adiponectin, decreased IGF-1). These patterns, however, are quite variable among genotypes and sex dependent, as seen for elevated FGF21 and leptin in *InsR^{R1109C}* males as well as reduced GDF15 and plasma triglycerides in *IGF-1R^{R1096C}* females.

InsR^{R1109C} and *IGF-1R^{R1096C}* heterodimer receptors have partial altered function. We determined how the KID mutation in IR or IGF-1R affects receptor function, measured as ligand-stimulated receptor kinase activity in human cell culture and by downstream signaling in murine tissue and in human cell culture.

We transfected IR/IGF-1R double-knockout 293FT cells with human IGF-1R^{WT} alone, IGF-1R^{R1096C} alone (corresponding to mouse IGF-1R R1096C, hereafter referred to as R1096C), or a 1:1 mixture of both plasmids to produce heterodimer receptors (Figure 7, A and B). To assess relative expression of these receptor types, we cotransfected Myc-tagged R1096C with untagged IGF-1R^{WT}. IGF-1 stimulation robustly induced autophosphorylation of IGF-1R^{WT} and phosphorylation of AKT and ERK. In contrast, R1096C homodimers exhibited

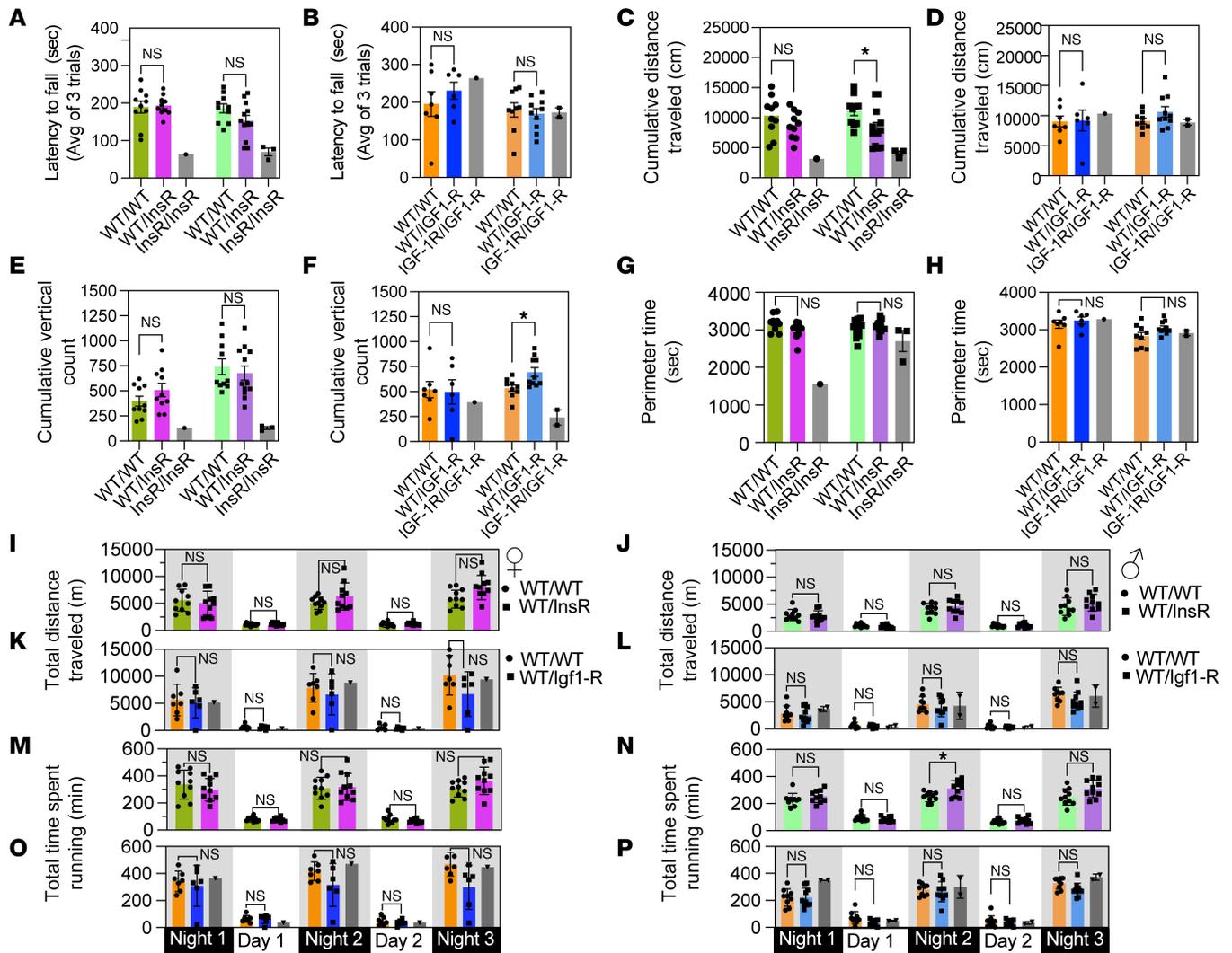


Figure 5. Neuromuscular function in *InsR^{R109C}* and *IGF-1R^{R1096C}* heterozygous mice. (A and B) Latency to rotarod test fall in female and male (A) *InsR^{R109C}* and (B) *IGF-1R^{R1096C}*. (C and D) Cumulative distance traveled in the open field test in female and male (C) *InsR^{R109C}* and (D) *IGF-1R^{R1096C}*. (E and F) Cumulative Vertical Activity the open field test in female and male (E) *InsR^{R109C}* and (F) *IGF-1R^{R1096C}*. (G and H) Time spent in margin areas in the open field test in female and male (G) *InsR^{R109C}* and (H) *IGF-1R^{R1096C}*. (I, J, K, and L) Total distance traveled in wheel-running in (I) female *InsR^{R109C}*, (J) male *InsR^{R109C}*, (K) female *IGF-1R^{R1096C}*, and (L) male *IGF-1R^{R1096C}*. (M, N, O, and P) Total time spent wheel-running in (M) female *InsR^{R109C}*, (N) male *InsR^{R109C}*, (O) female *IGF-1R^{R1096C}*, and (P) male *IGF-1R^{R1096C}*. WT and heterozygous were available for statistical comparison (females WT/WT and WT/InsR $n = 10$; males WT/WT $n = 10$ and WT/InsR $n = 10$ –12; females WT/WT $n = 7$ and WT/IGF-1R $n = 6$; males WT/WT $n = 9$ and WT/IGF-1R $n = 9$ –10). Homozygous mutant mice were rare. Data are represented as means \pm SD, Student's t test. * $P < 0.05$.

markedly reduced autophosphorylation. Despite this defect, IGF-1 ligand still induced AKT phosphorylation to approximately 80% of the level observed in WT-expressing cells, whereas there is only weak phosphorylation of ERK (Figure 7, A and B). Assuming equal expression, cotransfection of IGF-1R^{WT} and R1096C is expected to generate 25% WT:WT homodimers, 50% WT:R1096C heterodimers, and 25% R1096C:R1096C homodimers. If the heterodimer is inactive, IGF-1-induced autophosphorylation should resemble that of cells expressing R1096C alone. However, cells coexpressing WT and R1096C showed IGF-1R autophosphorylation at ~50% of WT levels and pERK activation at ~70% of those in WT-expressing cells, suggesting that the WT:R1096C heterodimer produces partial functional activity (Figure 7, A and B).

We performed similar experiments using human IR^{WT} and an IR^{R1107C} mutant (corresponding to mouse IR-A R1109C, hereafter referred to as R1109C) (Figure 7, C and D). R1109C homodimers showed greatly reduced autophosphorylation and impaired pERK, while pAKT remained comparable with IR^{WT} homodimer cells. Coexpression of IR^{WT} significantly restored autophosphorylation and pERK signaling in the presence of R1109C. Together, these results support the conclusion that both IGF-1R WT:R1096C and IR WT:R1109C heterodimers are functional whereby they retain kinase activity and partial ability to engage downstream signaling pathways.

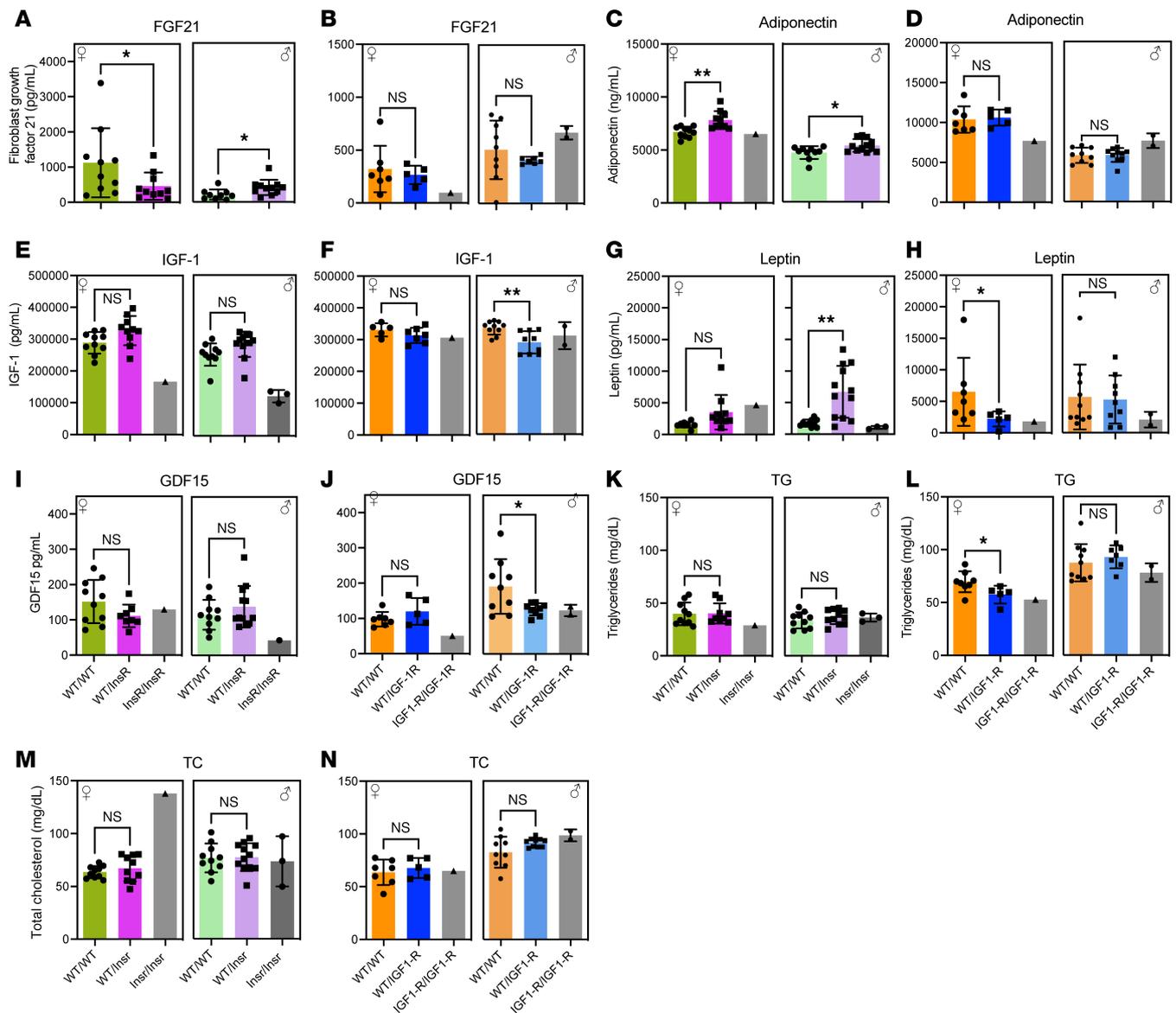


Figure 6. Hormonal and metabolite profiles in *InsR*^{R1109C} and *IGF-1R*^{R1096C} heterozygous mice. (A–L) Hormone plasma levels in female and male mice from *InsR*^{R1109C} and *IGF-1R*^{R1096C} mice respectively, of FGF21 (A and B), Adiponectin (C and D), IGF-1 (E and F), Leptin (G and H), GDF15 (I and J), Triglycerides (TG) (K and L), and Total cholesterol (TC) (M and N). WT and heterozygous were available for statistical comparison (females WT/WT and WT/*InsR* *n* = 10; males WT/WT *n* = 9–10 and WT/*InsR* *n* = 11–12; females WT/WT *n* = 7 and WT/*IGF-1R* *n* = 5–6; males WT/WT *n* = 10 and WT/*IGF-1R* *n* = 9–10). Homozygous mutant mice were rare. Data are shown as mean ± SD, Student's *t* test. **P* < 0.05; ***P* < 0.01.

We likewise assessed signal transduction pathways (pERK/ERK and pAKT/AKT) from liver tissue of *InsR*^{R1109C} and *IGF-1R*^{R1096C} heterozygote mice. We also measured pACC/ACC and pS6/S6 to describe altered AMPK and mTORC1 activity, which are characteristic of some long-lived mice (35–39).

pErk/Erk levels were significantly higher in livers of male and female *InsR*^{R1109C} heterozygotes (Supplemental Figure 3, A–D) but were not affected in *IGF-1R*^{R1096C} heterozygotes (Supplemental Figure 3, A–D). pACC/ACC levels were normal in *InsR*^{R1109C} females but significantly decreased in livers of male *InsR*^{R1109C} heterozygotes (Supplemental Figure 3, A–D). In contrast, pACC/ACC was elevated in male and female *IGF-1R*^{R1096C} heterozygotes (Supplemental Figure 3, A–D). pAKT/AKT was normal in *InsR*^{R1109C} females and males (Supplemental Figure 3, A–D) but significantly decreased in *IGF-1R*^{R1096C} females and increased in males (Supplemental Figure 3, A–D); pS6/S6 was significantly lower in *InsR*^{R1109C} females but normal in *InsR*^{R1109C} males (Supplemental Figure 3, A–D). pS6/S6 was reduced in female *IGF-1R*^{R1096C} heterozygotes but was upregulated in male *IGF-1R*^{R1096C} heterozygotes (Supplemental Figure 3, A–D).

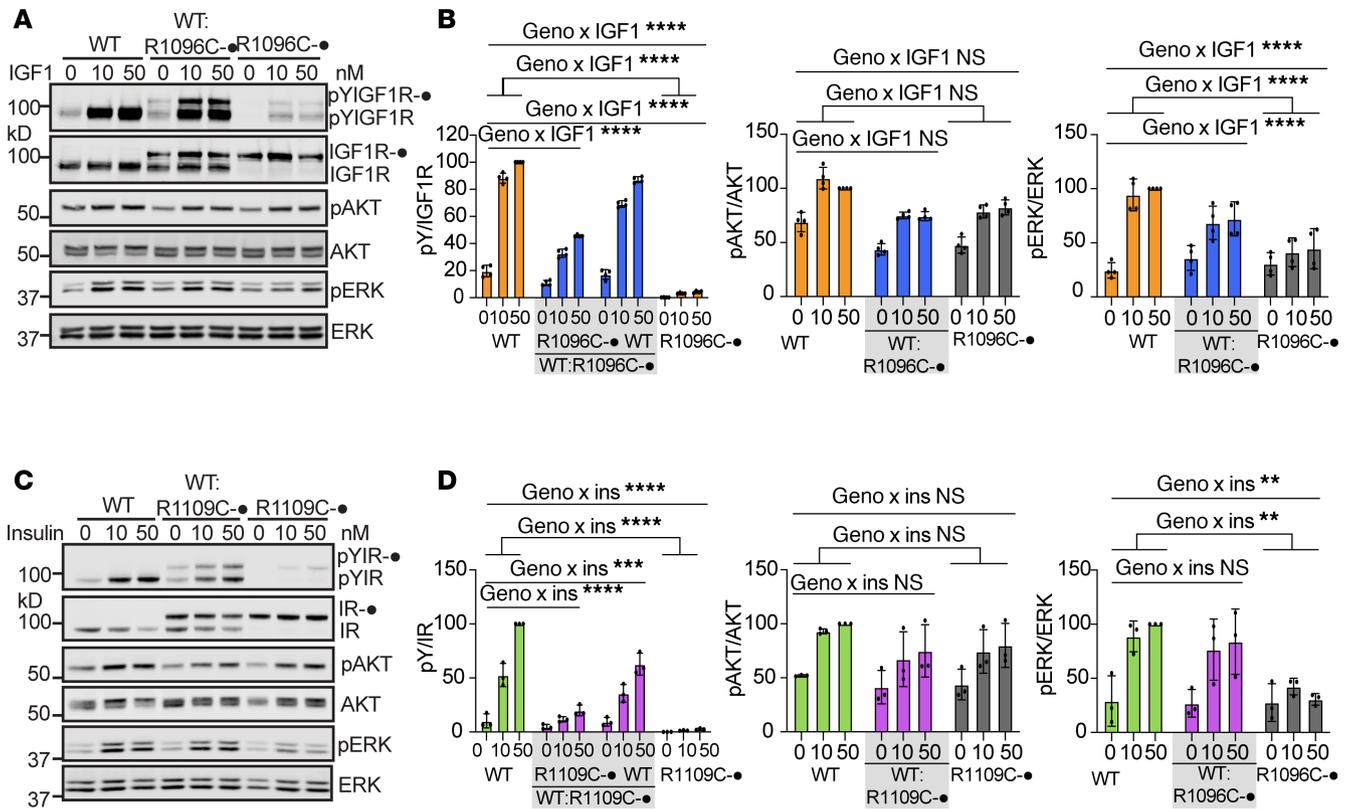


Figure 7. Receptor function in transfected 293FT cells with IGF-1R WT:R1096C heterodimers and IR WT:R1109C heterodimers. (A) IGF-1R signaling in response to the indicated concentrations of IGF-1 for 10 minutes in IR/IGF-1R knockout 293FT cells expressing IGF-1R WT and mutant receptors. **(B)** Quantitative analysis of the Western blot data. Levels of IGF-1R autophosphorylation were normalized to total IGF-1R levels and presented as intensities relative to those in IGF-1R WT cells treated with 50 nM IGF1. Data are shown as mean \pm SD. Significance evaluated by 2-way ANOVA with interaction among $n = 4$ independent experiments, $***P < 0.0001$. **(C)** IR signaling in response to concentrations of insulin for 10 minutes in IR/IGF-1R knockout 293FT cells expressing IR WT and mutant receptors. **(D)** Quantitative analysis of the Western blot data. Levels of IR autophosphorylation were normalized to total IR levels and presented as intensities relative to those in IR WT cells treated with 50 nM insulin. Data are shown as mean \pm SD. Significance evaluated by 2-way ANOVA with interaction among $n = 3$ independent experiments, $**P < 0.01$; $***P < 0.001$; $****P < 0.0001$.

Overall, the activity of TOR, AMPK, and MAPK signaling in mouse tissue was partially altered by $InsR^{R1109C}$ and $IGF-1R^{R1096C}$ when these mutants complement a WT receptor allele. Some of these changes (Table 1) are seen in other models of extended murine longevity, while other observations here are ambiguous, such as where male $IGF-1R^{R1096C}$ increase both AMPK activity and TOR activity.

Decreased biological age in livers of heterozygous female IGF-1R^{R1096C} mice. In this preliminary report, we cannot assess aging through survival analysis. Accordingly, we use transcriptomic biomarkers as proxies of aging and mortality. Recently developed transcriptome-based clocks can predict age-associated mortality and identify the molecular pathways that mediate differences in biological age (40).

We focused on IGF-1R mice using RNA-Seq from gastrocnemius muscle and liver of WT and heterozygote $IGF-1R^{R1096C}$ of both sexes at 4 months of age. We did not include heterozygote $InsR^{R1109C}$ mice in the analysis because they develop mild hyperinsulinemia. PCA of muscle samples separated the samples based on sex but not by genotype (Supplemental Figure 4A). PCA of the liver tissue separated samples by sex on the first principal component and by genotype among females on the second principal axis (Figure 8A). This suggests there is a sex-specific effect of the $IGF-1R^{R1096C}$ KID within the liver. Accordingly, over 5,000 differentially expressed genes ($P_{adj} < 0.05$) were identified in livers of $IGF-1R^{R1096C}$ heterozygous females, while few DEGs were seen in female muscle or in either tissue of males (Figure 8B, Supplemental Figure 4B, and Supplemental Table 1). Interestingly, the expression of *Igf1* and *Igf1r* were significantly upregulated in the liver of female $IGF-1R^{R1096C}$ mice, while IGF-1 in plasma was significantly decreased in $IGF-1R^{R1096C}$ male heterozygotes (Supplemental Figure 4C).

We compared the transcriptional profiles from the female $IGF-1R^{R1096C}$ liver to characteristic signatures seen in established murine models of delayed aging (41, 42). This revealed a positive correlation with longevity-associated signatures and a negative correlation with aging- and mortality-associated biomarkers. We observed a

Table 1. Summary of changes in physiological and biochemical parameters in heterozygous male and female mice of *InsR*^{R1109C} and *IGF-1R*^{R1096C} genotypes compared with WT mice

Parameter	<i>InsR</i> ^{R1109C}		<i>IGF-1R</i> ^{R1096C}	
	Female	Male	Female	Male
Growth Rate and Body Composition				
Body weight	NC	NC	↓	NC
Lean mass %	NC	NC	NC	NC
Fat mass %	NC	↑	NC	NC
Metabolic rates				
RER	NC	NC	NC	NC
Feeding	NC	NC	NC	NC
EE	NC	NC	NC	NC
Glucose metabolism				
Glucose AUC	NC	NC	NC	NC
Insulin AUC	↑	↑	NC	NC
HbA1c%	NC	NC	NC	NC
Neuromuscular function				
Rotarod: latency to fall	NC	NC	NC	NC
OF: distance traveled	↓	NC	NC	NC
OF: vertical activity	NC	NC	NC	↑
OF: margin time	NC	NC	NC	NC
Wheels: total distance	NC	NC	NC	NC
Wheels: time spent	NC	↑	NC	NC
Hormone signature				
IGF-1	NC	NC	NC	↓
Insulin	↑	↑	↓	NC
Triglycerides	NC	NC	↓	NC
Total cholesterol	NC	NC	NC	NC
Leptin	NC	↑	↓	NC
Adiponectin	↑	↑	NC	NC
FGF21	↓	↑	NC	NC
GDF15	NC	NC	NC	↓
Signaling pathways				
pERK/ERK	↑	↑	NC	↑
pACC/ACC	NC	↓	↑	↑
pS6/S6	↓	NC	↓	↑
Transcriptome-based biological age				
Transcriptomic clock	Not tested	Not tested	↓ (liver)	NC

Arrows indicate an increase (↑), decrease (↓), or no change (NC) compared with WT mice. Bold arrows are changes associated with extended longevity in other murine models and humans.

positive correlation with the signature specific to calorie restriction ($P_{\text{adj}} < 0.001$) and a negative association with signatures characteristic of aging-associated degeneration (kidney aging and rodent aging, each $P_{\text{adj}} < 0.001$) (Supplemental Figure 4D). Gene set enrichment analysis (GSEA) from *IGF-1R*^{R1096C} liver samples revealed a negative normalized enrichment score (NES) for metabolic processes (fatty acid metabolism, oxidative phosphorylation), complement, mTORC1 signaling, and hypoxia (Figure 8C and Supplemental Table 2). The observed downregulation of mTORC1-associated genes is consistent with our Western blot data where pS6 is decreased in *IGF-1R*^{R1096C} livers (Supplemental Figure 3C).

We then applied multitissue transcriptomic clocks of expected mortality (40) to estimate transcriptomic age (tAge) in tissue samples of female WT and IGF-1R^{R1096C} heterozygous animals. While individual organs did not show statistically significant differences, likely due to low sample size (Supplemental Figure 4E), female IGF-1R^{R1096C} heterozygotes displayed significantly lower tAge when liver and muscle tissues were pooled (Figure 8D). We applied module-specific transcriptomic mortality clocks to gain mechanistic insight into the biological pathways contributing to this effect (40). Several transcriptomic modules showed reduced tAge in liver tissue of IGF-1R^{R1096C} females ($P_{\text{adj}} < 0.05$) (Figure 8D), but no significant differences were observed in other tissues or in males. Strongly affected modules included lipid metabolism, VEGF signaling, mRNA splicing, mitochondria, NRF2 signaling, adaptive immunity, amino acid metabolism, heat-stress response, and translation (Figure 8E). Modules related to IFN signaling and chromatin modification showed modest increases in tAge, suggesting that while antiaging effects are predominant in this model, some proaging signals may be present.

In summary, female IGF-1R^{R1096C} heterozygous mice exhibit decreased biological age, particularly in the liver, highlighting potential molecular pathways contributing to the health-promoting effect of this genetic model, and this finding warrants an actual lifespan experiment.

Discussion

Mouse InsR^{R1109C} or IGF-1R^{R1096C} substitutions in the KID. A mutation in the KID of the *Drosophila* insulin/IGF receptor extends lifespan without impairing insulin-sensitivity, growth, and reproduction (5). The allele (referred to as *dInr^{KID}* or *dInr³⁵³*) contains an amino acid substitution (Arg1466Cys) within a sequence of the KID that is conserved in the KID of the mammalian insulin receptor (IR-A; Arg1107 in human, Arg1109 in mouse) and the IGF-1R (Arg1095 in human, Arg1096 in mouse) (Supplemental Table 3). This receptor domain has previously been known only for its effect on pathology. Donohue syndrome (OMIM #246200) is a rare and severe autosomal recessive disease caused by mutations in the insulin receptor. Homozygotes have strong insulin resistance, severe growth retardation, and hypertrichosis (43). Donohue syndrome can be caused by a homozygous Arg1107Gln substitution in the insulin receptor KID (44), the residue mutated in the long-lived of *Drosophila* (Arg1466Cys). Heterozygote parents have mild type A insulin resistance syndrome (43, 44). In recent cell-based studies, Chen et al. (45) showed Arg1104Glu combined with Arg1107Glu inhibits A-loop-tyrosine phosphorylation in stimulated mutant IR homodimers. Hall et al. (46) investigated how human IR substitutions induce premature endocytosis, finding that homodimers with the Arg1107Gln substitution remain at the plasma membrane in cells at their basal, unstimulated state. No data yet describe how substituting nonpolar cysteine for arginine in the IR KID will affect cell traits or animal phenotypes, but we expect it will not recapitulate what is seen with substitution with polar Gln or negatively charged Glu.

To empirically explore the biological relevance of Arg>Cys in the mammalian receptors, we generated 2 knock-in mouse models that carry the homologous fly substitution (Arg>Cys). Like flies — when the KID mutant is heterozygous with a WT receptor allele — young heterozygote KID mice have normal growth and glucose metabolism, yet they variably modify several hormones that are associated with slow aging in rodents and humans (Table 1).

Modified hormones of murine KID mutants. The murine InsR^{R1109C} and IGF-1R^{R1096C} KID substitutions alter several hormones associated with delayed aging in established mouse longevity models and in long-lived humans.

Heterozygote IGF1R^{Arg1096Cys} males have reduced plasma IGF1. In mammals, growth hormone produced by the anterior pituitary regulates the biosynthesis and release of IGF-1 by the liver and peripheral tissue to control mammalian growth. Several models of dwarf mice (Prop1, Pit1, GHRHR, and GHR) are long lived and all produce less IGF-1 (47–50). Compensatory upregulation of IGF-1 levels has been observed in some models with a tissue-specific KO of IGF-1R (51–53), while heterozygous *Igfl* mutant mice have been reported to have reduced circulating IGF-1 (54–56). Our model is potentially more like the second case, as we have full-body knock-in mutant mice and we observe decreased plasma IGF-1 levels in heterozygous IGF-1R^{R1096C} male mice. The absence of altered plasma IGF-1 in heterozygous IGF-1R^{R1096C} female mice may arise because they transcriptionally upregulate *Igflr* and *Igfl* mRNA.

Heterozygote InsR^{R1109C} males and females have elevated plasma adiponectin. Adiponectin null mice have reduced health span and lifespan, while transgenic mice with high circulating adiponectin improve health span and lifespan (57). A pan-adiponectin receptor agonist (AdipoRon) administered

Overall, the murine KID mutations variously affect a range hormones that are associated with attenuated aging in long-lived mice and humans. Aspects of these profiles parallel observations seen with dietary MetR.

Genetic background. We have explored substitutions in the KID in 1 genetic background, C57BL/6J. The knock-in mutations were produced in C57BL/6J and backcrossed to WT C57BL/6J mice to minimize the effect of off-target mutations. Previous work with *igf1r* and insulin receptor mutants shows that outcomes can vary among strain backgrounds (C57BL/6, 129Sv, and DBA) (66). Among these backgrounds, C57BL/6J may provide a stringent test condition because loss of either receptor produces severe phenotypes with obesity and diabetes. It is a sensitive background to detect deleterious metabolic phenotypes of the *InsR*^{R1109C} or *IGF-1R*^{R1096C} knock-in mutations. Here we found these KID substitutions (as WT heterozygotes) are metabolically healthy (IGF-1R mice) or have moderate hyperinsulinemia (IR mice). Potential future work with these receptor mutants will include additional metabolic tests (insulin tolerance test and hyperinsulinemic-euglycemic clamp studies) and alternative backgrounds such as outbred HET3 (67, 68).

Models of mutant IR and IGF1R to study aging. Manipulations of insulin/IGF receptors typically used to extend lifespan in *C. elegans* and *Drosophila* also reduce insulin sensitivity, impede development (dauer, maturation, growth), and impair fecundity. We subsequently described a potentially novel mutant *Drosophila* receptor that slowed aging without such negative effects, and here sought to test similar alleles in mammals, being aware of how fly and mammalian insulin/IGF systems differ (69). Our work with a functional allomorph allele in mice contrasts with previous mouse studies that tested IR and IGF-1R loss-of-function genotypes. One early report found *igf1r*^{+/-} males and females were long-lived, but this benefit was not replicated in a different genetic background (9). KO of the IR in adipose tissue with the aP2 Cre driver extended mouse longevity, but the fat in these mice was developmentally abnormal (17, 18). Later studies demonstrated the aP2 promoter was promiscuous, and further studies using other fat-specific Cre drivers found detrimental effects of insulin receptor KO on metabolic health that accompanied opposite effects on lifespan (70, 71). Two teams mutated the insulin receptor substates IRS1 or IRS2 (72–74), but the gene attributed to extend lifespan differed between the labs. Mao et al. injected IGF-1R antibody into 18-mo old mice, which improved life expectancy and health span in females but not in males (23). Ambiguity also extends to humans where individuals with Laron syndrome, a human dwarf disease associated with IGF-1 deficiency, seem protected against cancer but otherwise appear to have normal lifespan (75).

Past studies of insulin and IGF receptor modulation of mouse aging have reduced the abundance of ligand, receptor, or receptor substrate, but these trails did not consistently slow aging. We propose an alternative approach to manipulate the quality of IR or IGF-1R signaling rather than their quantity. We show that heterodimer receptors composed of a mutant and WT protomer retain the ability to be autophosphorylated, although to a lesser extent. Nonetheless, ligand is equally able to induce pAkt in both IR and IGF-1R heterodimer containing cells, while the IGF-1R mutant impairs the induction of pErk. The IGF-1R KID may have a selective effect on MAPK signaling that cannot be revealed by fully knocking out an allele. The KID mutations may recapitulate targeting of the insulin/IGF/mTORC1 axis with rapamycin and the Ras/MEK/ERK axis with trametinib, where simultaneous inhibition of both axes additively extends lifespan in HET3 mice (76).

Biological age of IGF1R KID mutant. Progressive change in DNA methylation levels are sometimes used to systematically predict age and evaluate longevity interventions across different species, including humans (77, 78). Although epigenetic (or methylation) clocks are a powerful approach to estimate aging using blood or other tissues, they provide limited mechanistic insight. In contrast, recently developed transcriptome-based clocks can predict expected mortality and identify molecular pathways associated with the observed differences in biological age (40). These transcriptome-based clocks were trained on multiple interventions with known effects on lifespan derived from the Intervention Testing Program and other large survival studies. This approach not only predicts effects on mortality using gene expression data, but it also compares these changes to transcriptome profiles induced by lifespan-extending interventions. When we compared the transcriptional profiles from the livers of female IGF-1R^{R1096C} mice to characteristic signatures observed in established murine models of delayed aging, we found a positive correlation with the signature characteristic of calorie restriction and a negative association with signatures characteristic of aging-related degeneration (kidney aging and multitissue rodent aging). The correlation with the calorie restriction signature is not surprising, given the critical role of the insulin/IGF/mTORC1 axis in mediating the beneficial effects of calorie restriction (79). Interestingly, while kidney pathology is a hallmark manifestation of diabetes and insulin resistance, we observe that the IGF-1R KID mutant opposes kidney aging, consistent with its prolongevity effect and absence of detrimental effects on metabolic health.

To connect the transcriptional changes to specific processes that drive the aging process, the clocks were also trained on coregulated transcriptomic modules enriched for specific cellular pathways associated with aging and mortality (40). Applying module-specific transcriptomic mortality clocks to the IGF1R KID samples, we found that IGF-1R^{R1096C} females exhibited a statistically significant reduction in tAge in liver tissue associated with 39% of the module clocks. The most affected modules with the strongest tAge decrease included those related to lipid metabolism, VEGF signaling, mRNA splicing, mitochondria, NRF2 signaling, adaptive immunity, amino acid metabolism, heat stress response, and translation. Although we do not yet know how the IGF-1R KID mutation drives transcriptional changes in these modules, our recent work in *Drosophila* identified methionine metabolism as a critical downstream effector of the KID mutation (80), and methionine metabolism is known to regulate most of the processes identified in these transcriptionally altered modules (81). It should also be noted that data from transcriptome-based clocks are predictive of delayed aging but are not a substitute for an actual lifespan experiment, as some of the effects (both beneficial and detrimental) of the IGF-1R KID mutation may only appear later in life.

We suggest that understanding how insulin or IGF signaling can modulate mammalian aging may be advanced through analysis of a single amino acid substitution in the KID, as we demonstrated in *Drosophila* (5). Here we describe how a homologous substitution in the murine KID produces viable animals with normal growth and robust carbohydrate metabolism. These animals have encouraging early signatures of slowed aging, which may be fully revealed through further analysis across the lifespan of aging cohorts.

Methods

Sex as a biological variable. Both male and female mice were used in this study.

Igf1r-R1096C (AGG >TGC) and Insr-R1109C (AGG>TGC) mouse model generation. C57Bl/6J mice (stock no. 000664) were purchased from Jackson Laboratory and housed with ad libitum diet and drinking water in a barrier facility (biosafety level 2) on 12-hour light-dark cycles. The protocol for generating genetically modified mice through CRISPR-Cas9 genome editing was approved by the IACUC of Brown University. All procedures involving mice were conducted in accordance with protocols approved by the IACUC of the University of Pittsburgh.

Three- to 4-week-old WT C57Bl/6J females were superovulated with pregnant mare serum gonadotropin (PMSG, Prospec Bio; I.P. 5IU/mouse) and human chorionic gonadotropin (HCG, Sigma-Aldrich; I.P. 5IU/mouse), followed by mating with WT stud males. At about 43.5 hours post-HCG administration, 2-cell embryos were harvested from successfully mated females and cultured in KSOM^{AA} medium (CytoSpring) before microinjection. The targeted mutation in mouse embryos was generated by cytoplasmic microinjection of both blastomeres with the mixed CRISPR-Cas9 reagents. The microinjection reagents (final concentration) were prepared in UltraPure DNase/RNase-Free distilled water (Invitrogen) following the steps of annealing crRNA and tracrRNA (molar ratio 1:1) according to IDT protocol, incubating Alt-R S.p. HiFi Cas9 Nuclease V3 (100 ng/ μ L, IDT) and annealed crRNA-tracrRNA (150 ng/ μ L, IDT) at room temperature for 10 minutes to form Cas9 ribonucleoprotein; and adding Ultramer DNA oligo template containing the mutation *Igf1r*-R1096C (AGG >TGC) or *Insr*-R1109C (AGG>TGC) (250 ng/ μ L, IDT). After microinjection, 25–30 embryos per recipient were transferred into CD1 pseudo-pregnant females.

The mice born from the microinjected embryos were genotyped to identify the targeted mutation. Genotypes were determined by Sanger sequencing analysis of the purified PCR products specifically amplified by a pair of primers with at least 150 nucleotides upstream and downstream of the targeted mutation site. The mice carrying the targeted mutation (founder mice, F0) were saved for germline transmission. Natural mating or in vitro fertilization (male founder mice only) of the founders with WT mice was performed to produce the F1 generation of mice. The heterozygous mice with the germline-transmitted targeted mutation were identified by genotyping. A heterozygous mouse was further backcrossed to WT C57Bl/6J mice to establish its colony. Supplemental Table 4 contains reagent sequence information.

Insr^{R1109C} and IGF-1R^{R1096C} breeding. To minimize the risk of off-target effects caused by CRISPR-Cas9, founder mice for both knock-in mutations were backcrossed with C57Bl/6J WT mice (JAX, stock no. 000664) for 2 generations and selected for the presence of *Insr^{R1109C}* and *IGF-1R^{R1096C}* allele.

Genotyping. Tail tips were collected from pups, and DNA was extracted with the Qiagen DNeasy Blood & Tissue kit (no. 69506). PCR reactions were performed using Hot Start Taq Blue master mix (Apex Bioreserch), with forward primers *GGCAAGTGAGATTTGCTTGGG* for *Insr^{R1109C}* and *GGGTAGTTTC-CCCGTTGCAT* for *IGF-1R^{R1096C}*, and reverse primers *ACAGGGTTGCAATTAGCACT* for *Insr^{R1109C}* and

CCATGACACGTGGTAGAGCA for IGF-1R^{R1096C} with the following program: first cycle of denaturation at 95°C for 15 minutes and 28 cycles of denaturation at 95°C for 30 seconds, annealing at 59°C for 30 seconds, extension at 72°C for 1 minute, and a final extension at 72°C for 5 minutes. Fragments were visualized on 1% agarose gel using SYBER Safe gel stain (Thermo Fisher Scientific, S33102): InsR^{R1096C} amplicon of 476 bp and IGF-1R^{R1096C} amplicon of 588 bp. PCR products were sent for sanger sequencing (Azenta Genewiz) and point mutations were assessed with SnapGene 7.2 software. Simultaneous peaks for A and T and for G and C indicated individuals heterozygous for a WT codon AGG and a mutant codon tGC.

Blood biochemical tests. ELISA measured FGF-21 (EZRMFGF21-26K) (Millipore), adiponectin (ADIPOQ) (MRP300) (R&D Systems), insulin-like growth factor 1 (IGF-1) (MG100), leptin (MOB00B), and GDF15 (R&D Systems, DY6385). A colorimetric assay determined plasma triglycerides (TG) (TR22421) (Thermo Fisher Scientific) and total cholesterol (TR13421) (Thermo Fisher Scientific).

IR/IGF-1R double knockout 293FT cells. IR and IGF-1R double KO 293FT cells — generated in ref. 82 — were cultured in high-glucose (4.5 g/L) DMEM supplemented with 10% (v/v) FBS, 2 mM L-glutamine, and 1% penicillin/streptomycin. Cells were maintained at 37°C with a humidified atmosphere of 5% CO₂.

Cell-based receptor activation assays. IR and IGF-1R activation assays were performed as previously described with some modifications (82, 83). For IR mutant generation, we used the short isoform of human IR (hIR-A) in pCS2-Myc vector resistant to IR gRNAs. IR R1109C mutation was generated by Q5 site-directed mutagenesis (pCS2-hIR-A-Myc, primer sequences: 5'- CCGTTCTCTGtgcCCA-GAGGCTG -3' and 5'- AGGTAGCTCTTCAGGTCTC -3'). For IGF-1R R1096C mutant generation, we used human IGF-1R in pCS2-Myc vector resistant to IGF1R gRNAs. IGF-1R R1096C mutant was generated by Q5 site-directed mutagenesis (pCS2-hIGF-1R-Myc, primer sequences: 5'- CCGGTCTCT-GtgcCCAGAAATGG -3' and 5'- AGATAACTTTTGAGATCGC -3').

Plasmid transfection in double-knockout 293FT cells was performed with Lipofectamine 2000 (Invitrogen). After 1 day, the cells were serum-starved for 16–18 hours. Serum-starved cells were treated with the indicated concentrations of human insulin (Sigma, I2643) or human IGF-1 (PeproTech, 100-11).

After treatment, cells were incubated with cell lysis buffer [50 mM Hepes pH 7.4, 150 mM NaCl, 10% (v/v) Glycerol, 1% (v/v) Triton X-100, 1 mM EDTA, 10 mM sodium fluoride, 2 mM sodium orthovanadate, 10 mM sodium pyrophosphate, 0.5 mM dithiothreitol (DTT), 2 mM phenylmethylsulfonyl fluoride (PMSF)] supplemented with cOmplete Protease Inhibitor Cocktail (Roche) and PhosSTOP (Roche) on ice for 1 hour. Cell were centrifuged at 18,213g at 4°C for 20 minutes, and cell lysates analyzed by SDS-PAGE and Western blotting. Primary antibodies included anti-IR-pY1150/1151 (1:2000; 19H7, Cell Signaling, 3024), anti-IR (1:500; CT-3, Santa Cruz, sc-57342), anti-IGF-1R (1:1000; ZI001, Invitrogen, 39-6700), anti-AKT (1:2000; 40D4, Cell Signaling, 2920), anti-pS473 AKT (1:2000; D9E, Cell Signaling, 4060), anti-ERK1/2 (1:2000; L34F12, Cell Signaling, 4696), and anti-pERK1/2 (1:2000; 197G2, Cell Signaling, 4377). Secondary antibodies for quantitative Western blots included anti-rabbit immunoglobulin G (IgG) (H + L) (Dylight 800 conjugates, Cell Signaling, 5151) and anti-mouse IgG (H + L) (Dylight 680 conjugates, Cell Signaling, 5470). The membranes were scanned with the Odyssey Infrared Imaging System (LI-COR, Lincoln, NE). Levels of receptor autophosphorylation, pERK, and pAKT were normalized to total receptor, ERK, and AKT levels, respectively. For IR activation assay, intensities were shown relative to that of IR-WT treated with 50 nM insulin. For IGF-1R activation assay, intensities were shown relative to that of IGF-1R-WT treated with 50 nM IGF-1.

RNA-Seq. Reads were mapped to mouse genome (GRCm39) with STAR (version 2.7.11b) and counted via featureCounts (version 2.0.6). To filter out nonexpressed genes, we required at least 10 reads in at least 20% of samples separately for liver and skeletal muscle data. Differentially expressed genes were identified separately for each tissue and sex using 1-way ANOVA model through edgeR package (84). *P* values were adjusted for multiple testing with the Benjamini-Hochberg method.

Transcriptomic signature analysis. We conducted functional enrichment analysis to describe how the transcriptome changes in IGF-1R^{R1096C} heterozygous mice in various tissues and across sex, and we related these to established molecular signatures of aging, mortality, and lifespan regulation. We applied reference signatures from tissue-specific aging biomarkers of liver, kidney, and brain, and from multitissue biomarkers of chronological age and expected mortality; adjusted for chronological age (40). Additionally, we included hepatic signatures of expected maximum lifespan in rodents and signatures of individual

longevity interventions, such as caloric restriction, genetic models of growth hormone deficiency, and rapamycin (41). For each tissue (liver and skeletal muscle) and sex, we ranked genes using a signed log-transformed P value metric estimated through differential expression analysis: $-\log(\text{pv}) \times \text{sgn}(\text{lfc})$, where pv and lfc are P value and $\log\text{FC}$ of a certain gene, respectively, and sgn is the signum function (equal to 1, -1, and 0 if value is positive, negative, or equal to 0, respectively). Ranked gene lists were subjected to GSEA using the *fgsea* package in R, with 10,000 permutations and multilevel Monte Carlo sampling. Gene sets were drawn from the HALLMARK, KEGG, and REACTOME collections of the Molecular Signatures Database (MSigDB). The same enrichment pipeline was applied to reference signatures of aging, mortality, and longevity interventions. Individual P values were adjusted for multiple testing with the Benjamini-Hochberg method. We computed Spearman correlations between NES to quantify similarities between signatures of IGF-1R^{R1096C} heterozygous mice and reference gene expression biomarkers.

tAge analysis. The filtered RNA-Seq data were processed with Relative Log Expression (RLE) normalization, log-transformation, and YuGene normalization (85). Missing expression values for clock genes not detected in the dataset were imputed using their corresponding precomputed average values. Normalized gene expression profiles were then centered to the median profile of control samples within each tissue. tAge for each sample was estimated using Elastic Net-based multitissue transcriptomic clocks of expected mortality (40). Module-specific transcriptomic clocks of expected mortality were applied to the scaled, relative gene expression profiles using the same framework. The resulting tAge values from module-specific clocks were standardized within each tissue and sex. One-way ANOVA was used to compare tAge estimates from composite and module-specific clocks. One-way ANOVA model including tissue as a covariate was used to compare composite clock tAges across tissues in females. Resulting P values were adjusted for multiple comparisons using the Benjamini-Hochberg method.

Statistics. For all comparisons between WT and heterozygous mice, data were assessed for normality (Shapiro-Wilk test) and evaluated by an unpaired 2-tailed Student's t test. Welch's correction was used if variances were not equal among groups. A Mann-Whitney U test was used when data were not normally distributed. Two-way ANOVA was used to evaluate the main and interaction effects upon phosphorylation events when ligands stimulated mutant receptors in cell culture. Metabolic cage data were normalized to weight and analyzed using 2-way ANOVA to compare across time cycles and genotypes. Alternatively (depending on whether all assumptions were satisfied: normality, homoscedasticity, and linearity), the data were also analyzed with ANCOVA, GAM, or GLM using lean mass as a covariate. Results from both analyses were nonsignificant; therefore, graphs based on the 2-way ANOVA were shown as a reference. All data are represented either by mean \pm SD for normally distributed data or as median \pm 95% CI for nonnormally distributed data (GraphPad Prism, version 10.4). Outliers were identified by the ROUT method. Growth curves were generated using a logistic 3-parameter model, and statistical differences between curves were determined using an equivalence test with JMP Pro software, version 17 (SAS Institute). ANCOVA was performed in R (version 4.5.0). The packages *rstatix*, *mgcv*, *car*, and *emmeans* were used for modeling, assumption testing, and post hoc comparisons.

All the key resources used in the study are listed in the Supplemental Table 5.

Extended materials and methods can be found in the Supplemental Table 6.

Study approval. The protocol for generating genetically modified mice through CRISPR-Cas9 genome editing was approved by the IACUC of Brown University. Animal experiments were approved by and in compliance with the University of Pittsburgh IACUC.

Data availability. All data are available within the article and Supporting Data Values file, or available from the authors upon a request. RNA-Seq data reported in this work are available at NCBI GEO data repository under accession no. GSE303415.

Author contributions

Conceptualization was contributed by MT and AAP. Methodology was contributed by AT, AM, JL, JKA, SJSR, GPA, EC, VNG, MJJ, MT, and AAP. Investigation was contributed by UHA, JKK, JLF, AT, AM, CH, SG, YG, IJS, JK, and DC. Visualization was contributed by UHA, AT, and EC. Supervision was contributed by MT and AAP. Writing of the original draft was contributed by UHA, MT, and AAP. Review and editing were contributed by UHA, AT, AM, SJSR, EC, VNG, MT, and AAP.

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Address correspondence to: Marc Tatar, Department of Ecology, Evolutionary Biology and Organismal Biology, Box G-W, Brown University, Providence, Rhode Island, 02912, USA. Phone: 401.863.3455; Email: marc_tatar@brown.edu. Or to: Andrey A Parkhitko, Aging Institute, Room 569, Bridgeside Point I, 100 Technology Dr., Pittsburgh, Pennsylvania, 15219, USA. Phone: 339.368.4594; Email: aparkhitko@pitt.edu.

1. Tatar M, et al. The endocrine regulation of aging by insulin-like signals. *Science*. 2003;299(5611):1346–1351.
2. Mathew R, et al. Insulin/insulin-like growth factor-1 signalling (IIS) based regulation of lifespan across species. *Biogerontology*. 2017;18(1):35–53.
3. Pan H, Finkel T. Key proteins and pathways that regulate lifespan. *J Biol Chem*. 2017;292(16):6452–6460.
4. Anisimov VN, Bartke A. The key role of growth hormone-insulin-IGF-1 signaling in aging and cancer. *Crit Rev Oncol Hematol*. 2013;87(3):201–223.
5. Yamamoto R, et al. Aging modulated by the Drosophila insulin receptor through distinct structure-defined mechanisms. *Genetics*. 2021;217(2):iyaa037.
6. Kenyon C, et al. A C. elegans mutant that lives twice as long as wild type. *Nature*. 1993;366(6454):461–464.
7. Kimura KD, et al. daf-2, an insulin receptor-like gene that regulates longevity and diapause in Caenorhabditis elegans. *Science*. 1997;277(5328):942–946.
8. Tatar M, et al. A mutant Drosophila insulin receptor homolog that extends life-span and impairs neuroendocrine function. *Science*. 2001;292(5514):107–110.
9. Xu J, et al. Longevity effect of IGF-1R(+/-) mutation depends on genetic background-specific receptor activation. *Aging Cell*. 2014;13(1):19–28.
10. Kolb H, et al. Insulin and aging - a disappointing relationship. *Front Endocrinol (Lausanne)*. 2023;14:1261298.
11. Kitamura T, et al. Insulin receptor knockout mice. *Annu Rev Physiol*. 2003;65:313–332.
12. Choi E, et al. Regulation and function of insulin and insulin-like growth factor receptor signalling. *Nat Rev Mol Cell Biol*. 2025;26(7):558–580.
13. Khan MZ, et al. The signaling landscape of insulin-like growth factor 1. *J Biol Chem*. 2025;301(1):108047.
14. Baba T, et al. Estrogen, insulin, and dietary signals cooperatively regulate longevity signals to enhance resistance to oxidative stress in mice. *J Biol Chem*. 2005;280(16):16417–16426.
15. Shimizu T, et al. Lifespan and glucose metabolism in insulin receptor mutant mice. *J Aging Res*. 2011;2011:315640.
16. Nelson JF, et al. Probing the relationship between insulin sensitivity and longevity using genetically modified mice. *J Gerontol A Biol Sci Med Sci*. 2012;67(12):1332–1338.
17. Blüher M, et al. Extended longevity in mice lacking the insulin receptor in adipose tissue. *Science*. 2003;299(5606):572–574.
18. Blüher M, et al. Adipose tissue selective insulin receptor knockout protects against obesity and obesity-related glucose intolerance. *Dev Cell*. 2002;3(1):25–38.
19. Holzenberger M, et al. IGF-1 receptor regulates lifespan and resistance to oxidative stress in mice. *Nature*. 2003;421(6919):182–187.
20. Bokov AF, et al. Does reduced IGF-1R signaling in Igf1r+/- mice alter aging? *PLoS One*. 2011;6(11):e26891.

21. Lorenzini A, et al. Mice producing reduced levels of insulin-like growth factor type 1 display an increase in maximum, but not mean, life span. *J Gerontol A Biol Sci Med Sci*. 2014;69(4):410–419.
22. Venz R, et al. End-of-life targeted degradation of DAF-2 insulin/IGF-1 receptor promotes longevity free from growth-related pathologies. *Elife*. 2021;10:e71335.
23. Mao K, et al. Late-life targeting of the IGF-1 receptor improves healthspan and lifespan in female mice. *Nat Commun*. 2018;9(1):2394.
24. Abdellatif M, et al. Fine-tuning cardiac insulin-like growth factor 1 receptor signaling to promote health and longevity. *Circulation*. 2022;145(25):1853–1866.
25. Locascio LE, Donoghue DJ. KIDS rule: regulatory phosphorylation of RTKs. *Trends Biochem Sci*. 2013;38(2):75–84.
26. Talal S, et al. High carbohydrate diet ingestion increases post-meal lipid synthesis and drives respiratory exchange ratios above 1. *J Exp Biol*. 2021;224(pt 4):jeb240010.
27. Livesey G, Elia M. Estimation of energy expenditure, net carbohydrate utilization, and net fat oxidation and synthesis by indirect calorimetry: evaluation of errors with special reference to the detailed composition of fuels. *Am J Clin Nutr*. 1988;47(4):608–628.
28. Elia M, Livesey G. Theory and validity of indirect calorimetry during net lipid synthesis. *Am J Clin Nutr*. 1988;47(4):591–607.
29. Choi HM, et al. An age-dependent alteration of the respiratory exchange ratio in the db/db mouse. *Lab Anim Res*. 2015;31(1):1–6.
30. Jeukendrup AE, Wallis GA. Measurement of substrate oxidation during exercise by means of gas exchange measurements. *Int J Sports Med*. 2005;26 Suppl 1:S28–S37.
31. Farinatti P, et al. Oxygen consumption and substrate utilization during and after resistance exercises performed with different muscle mass. *Int J Exerc Sci*. 2016;9(1):77–88.
32. Burtscher J, et al. Mitochondrial stress and mitokines in aging. *Aging Cell*. 2023;22(2):e13770.
33. Pence BD. Growth differentiation factor-15 in immunity and aging. *Front Aging*. 2022;3:837575.
34. Conte M, et al. GDF15, an emerging key player in human aging. *Ageing Res Rev*. 2022;75:101569.
35. Ge Y, et al. Role of AMPK mediated pathways in autophagy and aging. *Biochimie*. 2022;195:100–113.
36. Kulkarni AS, et al. Benefits of metformin in attenuating the hallmarks of aging. *Cell Metab*. 2020;32(1):15–30.
37. Laskovs M, et al. Molecular inhibition of RAS signalling to target ageing and age-related health. *Dis Model Mech*. 2022;15(10):dmm049627.
38. Mannick JB, Lamming DW. Targeting the biology of aging with mTOR inhibitors. *Nat Aging*. 2023;3(6):642–660.
39. Parkhitko AA, et al. Kinase mTOR: regulation and role in maintenance of cellular homeostasis, tumor development, and aging. *Biochemistry (Mosc)*. 2014;79(2):88–101.
40. Tyshkovskiy A, et al. Transcriptomic hallmarks of mortality reveal universal and specific mechanisms of aging, chronic disease, and rejuvenation [preprint]. <https://doi.org/10.1101/2024.07.04.601982>. Posted on bioRxiv July 7, 2024.
41. Tyshkovskiy A, et al. Identification and application of gene expression signatures associated with lifespan extension. *Cell Metab*. 2019;30(3):573–593.
42. Tyshkovskiy A, et al. Distinct longevity mechanisms across and within species and their association with aging. *Cell*. 2023;186(13):2929–2949.
43. Mesika A, et al. INSR-Related Severe Insulin Resistance Syndrome. In: Adam MP, et al. eds. *GeneReviews(R)*. University of Washington; 1993.
44. Takahashi Y, et al. A homozygous kinase-defective mutation in the insulin receptor gene in a patient with leprechaunism. *Diabetologia*. 1997;40(4):412–420.
45. Chen L, et al. Molecular basis for receptor tyrosine kinase A-loop tyrosine transphosphorylation. *Nat Chem Biol*. 2020;16(3):267–277.
46. Hall C, et al. Insulin receptor endocytosis in the pathophysiology of insulin resistance. *Exp Mol Med*. 2020;52(6):911–920.
47. Gesing A, et al. A long-lived mouse lacking both growth hormone and growth hormone receptor: a new animal model for aging studies. *J Gerontol A Biol Sci Med Sci*. 2017;72(8):1054–1061.
48. Flurkey K, et al. Lifespan extension and delayed immune and collagen aging in mutant mice with defects in growth hormone production. *Proc Natl Acad Sci U S A*. 2001;98(12):6736–6741.
49. Brown-Borg HM, et al. Dwarf mice and the ageing process. *Nature*. 1996;384(6604):33.
50. Chandrashekar V, et al. Pituitary and testicular function in growth hormone receptor gene knockout mice. *Endocrinology*. 1999;140(3):1082–1088.
51. Perez-Matute P, et al. IGF1R is a mediator of sex-specific metabolism in mice: effects of age and high-fat diet. *Front Endocrinol (Lausanne)*. 2022;13:1033208.
52. Al-Samerria S, Radovick S. The role of insulin-like growth factor-1 (IGF-1) in the control of neuroendocrine regulation of growth. *Cells*. 2021;10(10):2664.
53. Romero CJ, et al. Targeted deletion of somatotroph insulin-like growth factor-I signaling in a cell-specific knockout mouse model. *Mol Endocrinol*. 2010;24(5):1077–1089.
54. Vitale G, et al. ROLE of IGF-1 system in the modulation of longevity: controversies and new insights from a centenarians' perspective. *Front Endocrinol (Lausanne)*. 2019;10:27.
55. De Ita JR, et al. Altered liver expression of genes involved in lipid and glucose metabolism in mice with partial IGF-1 deficiency: an experimental approach to metabolic syndrome. *J Transl Med*. 2015;13:326.
56. Guerra-Menendez L, et al. IGF-I increases markers of osteoblastic activity and reduces bone resorption via osteoprotegerin and RANK-ligand. *J Transl Med*. 2013;11:271.
57. Li N, et al. Adiponectin preserves metabolic fitness during aging. *Elife*. 2021;10:e65108.
58. Balasubramanian P, et al. Adiponectin receptor agonist AdipoRon improves skeletal muscle function in aged mice. *Elife*. 2022;11:e71282.
59. Arai Y, et al. High adiponectin concentration and its role for longevity in female centenarians. *Geriatr Gerontol Int*. 2006;6(1):32–39.
60. Hill CM, et al. FGF21 is required for protein restriction to extend lifespan and improve metabolic health in male mice. *Nat Commun*. 2022;13(1):1897.

61. Fang H, et al. Nutritional regulation of hepatic FGF21 by dietary restriction of methionine. *Front Endocrinol (Lausanne)*. 2021;12:773975.
62. Olsen T, et al. Effects of dietary methionine and cysteine restriction on plasma biomarkers, serum fibroblast growth factor 21, and adipose tissue gene expression in women with overweight or obesity: a double-blind randomized controlled pilot study. *J Transl Med*. 2020;18(1):122.
63. Holland WL, et al. An FGF21-adiponectin-ceramide axis controls energy expenditure and insulin action in mice. *Cell Metab*. 2013;17(5):790–797.
64. Zhang Y, et al. The starvation hormone, fibroblast growth factor-21, extends lifespan in mice. *Elife*. 2012;1:e00065.
65. Cooke D, et al. Weight loss and concomitant adipose autophagy in methionine-restricted obese mice is not dependent on adiponectin or FGF21. *Obesity (Silver Spring)*. 2020;28(6):1075–1085.
66. Kulkarni RN, et al. Impact of genetic background on development of hyperinsulinemia and diabetes in insulin receptor/insulin receptor substrate-1 double heterozygous mice. *Diabetes*. 2003;52(6):1528–1534.
67. Nadon NL, et al. NIA interventions testing program: investigating putative aging intervention agents in a genetically heterogeneous mouse model. *EBioMedicine*. 2017;21:3–4.
68. Austad SN. Issues in the choice of genetic configuration for animal aging models. *Exp Gerontol*. 1997;32(1-2):55–63.
69. Sell C. Minireview: The complexities of IGF/insulin signaling in aging: why flies and worms are not humans. *Mol Endocrinol*. 2015;29(8):1107–1113.
70. Friesen M, et al. Adipocyte insulin receptor activity maintains adipose tissue mass and lifespan. *Biochem Biophys Res Commun*. 2016;476(4):487–492.
71. Guerra C, et al. Brown adipose tissue-specific insulin receptor knockout shows diabetic phenotype without insulin resistance. *J Clin Invest*. 2001;108(8):1205–1213.
72. Selman C, et al. Evidence for lifespan extension and delayed age-related biomarkers in insulin receptor substrate 1 null mice. *FASEB J*. 2008;22(3):807–818.
73. Selman C, et al. Replication of extended lifespan phenotype in mice with deletion of insulin receptor substrate 1. *PLoS One*. 2011;6(1):e16144.
74. Taguchi A, et al. Brain IRS2 signaling coordinates life span and nutrient homeostasis. *Science*. 2007;317(5836):369–372.
75. Shevah O, Laron Z. Patients with congenital deficiency of IGF-I seem protected from the development of malignancies: a preliminary report. *Growth Horm IGF Res*. 2007;17(1):54–57.
76. Gkioni L, et al. The geroprotectors trametinib and rapamycin combine additively to extend mouse healthspan and lifespan. *Nat Aging*. 2025;5(7):1249–1265.
77. Chen C, et al. DNA methylation: from cancer biology to clinical perspectives. *Front Biosci (Landmark Ed)*. 2022;27(12):326.
78. Wei A, Wu H. Mammalian DNA methylome dynamics: mechanisms, functions and new frontiers. *Development*. 2022;149(24):dev182683.
79. Wilson KA, et al. Evaluating the beneficial effects of dietary restrictions: a framework for precision nutrigenetics. *Cell Metab*. 2021;33(11):2142–2173.
80. Tatar M, et al. An insulin-sensitive Drosophila insulin-like receptor mutant remodels methionine metabolism to extend lifespan. *PLoS Genet*. 2025;21(6):e1011640.
81. Parkhitko AA, et al. Methionine metabolism and methyltransferases in the regulation of aging and lifespan extension across species. *Aging Cell*. 2019;18(6):e13034.
82. An W, et al. Activation of the insulin receptor by insulin-like growth factor 2. *Nat Commun*. 2024;15(1):2609.
83. Park J, et al. Activation of the insulin receptor by an insulin mimetic peptide. *Nat Commun*. 2022;13(1):5594.
84. Robinson MD, et al. edgeR: a bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics*. 2010;26(1):139–140.
85. Le Cao KA, et al. YuGene: a simple approach to scale gene expression data derived from different platforms for integrated analyses. *Genomics*. 2014;103(4):239–251.