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#### A Randomized, Double-Blind, Placebo Controlled Trial of IL-7 in Critically ill COVID-19 Patients

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#### Abstract

**Background:** Lymphopenia and failure of lymphocytes to mount an early IFN-γ response correlate with increased mortality in COVID-19. Given the essential role of CD4 helper and CD8 cytotoxic cells in eliminating viral pathogens, this profound loss in lymphocytes may impair patients' ability to eliminate the virus. IL-7 is a pleiotropic cytokine that is obligatory for lymphocyte survival and optimal function.

**Methods:** We conducted a prospective, double-blind, randomized, placebo-controlled trial of CYT107, recombinant human IL-7, in 109 critically-ill lymphopenic COVID-19 patients. The primary endpoint was to assess CYT107's effect on lymphocyte recovery with secondary clinical endpoints including safety, ICU and hospital length-of-stay, incidence of secondary infections, and mortality.

**Results:** CYT107 was well-tolerated without precipitating a cytokine storm or worsening pulmonary function. Absolute lymphocyte counts increased in both groups without significant difference between CYT107 and placebo. COVID-19 patients receiving CYT107 but not concomitant antiviral medications, known inducers of lymphopenia, had a final lymphocyte count that was 43% greater than placebo (p=0.067). There were significantly fewer treatment-emergent adverse events in CYT107 versus placebo-treated patients (p<0.001), consistent with a beneficial drug effect. Importantly, CYT107 treated patients had 44% fewer hospital-acquired infections versus placebo-treated patients (p=0.014).

**Conclusions**: Given that hospital-acquired infections are responsible for a large percentage of COVID-19 deaths, this effect of CYT107 to decrease nosocomial infections could substantially reduce late morbidity and mortality in this highly lethal disease. The strong safety profile of CYT107 and its excellent tolerability provide support for trials of CYT107 in other potential pandemic respiratory viral infections.

Trial registration: NCT04379076, NCT04426201, NCT04442178, NCT04407689; NCT04927169

Key words: Immunology, immunotherapy, sepsis

#### Introduction

Currently, the predominant paradigm regarding the pathophysiology of COVID-19 is an excessive, hyperinflammatory, cytokine-mediated response leading to severe organ injury (1-3). A number of therapies have been demonstrated to improve morbidity and mortality in patients with COVID-19, e.g., corticosteroids, Janus kinase inhibitors, and IL-6 antagonists (4). These therapies act to attenuate the host immune response which is consistent with the understanding that a damaging cytokine response is the central driving force in the disorder.

Although an over-exuberant inflammatory cytokine-mediated response is likely playing a fundamental role in the COVID-19 disease process, a seemingly paradoxical alternative mechanism for the morbidity and mortality occurring in COVID-19 patients is immunosuppression with resultant failure of the host to mount an effective antiviral response (5-9). Most of the deaths due to COVID-19 occur in elderly patients with immunosenescence and/or patients with serious comorbidities, two patient groups that often fail to mount a robust immune response to invading pathogens. Additional data supportive of the contention that failure of host immunity is a critical factor in morbidity and mortality of COVID-19 is provided by studies showing a strong relationship between SARS-CoV-2 viral load and increased disease severity and mortality (10,11). Importantly, autopsy studies of patients who died of COVID-19 demonstrate viral persistence in lungs and other organs with viral inclusion bodies and extremely elevated levels of SARS-CoV-2 as detected by RT-qPCR (12-15). These postmortem studies showing persistence of high levels of the virus systemically are consistent with a failure of the host to eliminate the invading virus and that unchecked viral replication is likely contributing to morbidity and mortality. Impaired host immunity may also be a factor in the high incidence of secondary hospital acquired infections which occur in many critically ill COVID-19 patients and that are often the more proximate cause of death (16-19).

One of the earliest and most common reported defects in host immunity in patients with COVID-19 is lymphopenia (20). Autopsy studies of patients who died of COVID-19 showed that the decrease in circulating lymphocytes was accompanied by a corresponding depletion of tissue lymphocytes (12). In addition to a decrease in the number of lymphocytes, there is also a concomitant decline in lymphocyte function as indicated by impaired production of IFN- $\gamma$ , a cytokine which plays a key role in host defenses (5,9,21). Importantly, both lymphopenia and the failure of T cells to mount an IFN- $\gamma$  response correlate with an increase in fatal outcomes in COVID-19 patients (20,21). Given the essential role of CD4 helper T cells and CD8 cytotoxic T cells in eliminating viral pathogens, it is likely

that the profound loss in the number and function of lymphocytes is impairing the ability of the host to eliminate the virus (22-24).

IL-7 is a pleiotropic, anti-apoptotic cytokine that is required for lymphocyte survival and expansion (25,26). IL-7 reverses T cell exhaustion, a common immunologic abnormality occurring in viral infections such as COVID-19 (5,22). Animal studies have shown that IL-7 is required for the optimal response to acute influenza infection as it shapes the early priming of CD8 cytotoxic T cells (27). Prophylactic administration of IL-7 protects against lethal influenza in a murine model (27). IL-7 immune therapy has also shown the ability to decrease viral load and ameliorate organ injury in other viral infectious models (28).

Although not yet clinically approved, CYT107, a glycosylated, long-acting, recombinant human IL-7, has been used in over 650 adult and pediatric patients with severe lymphopenia due to various etiologies including sepsis, AIDS, radiation and chemotherapy, bone marrow reconstitution following stem cell transplantation, and refractory viral and fungal infections (29-32). In patients with septic shock and lymphopenia, CYT107 caused a 3-4-fold increase in circulating CD4 and CD8 T cells as well as inducing T cell proliferation and activation (31). In clinical trials of HIV-infected patients who had lymphopenia despite optimal anti-retroviral therapy, CYT107 caused a 2-4-fold dose-dependent increase in circulating CD4 and CD8 T cells (32). CYT107 treated patients consistently demonstrate increased circulating lymphocyte counts, and tolerate the drug well with injection site reactions being the only serious adverse effect. Importantly, we previously described the compassionate use of CYT107 in 12 critically ill, mechanically ventilated, COVID-19 patients with profound lymphopenia (33). The study showed that CYT107 was well-tolerated without exacerbating inflammation or aggravating lung injury and was associated with a more robust return of lymphocytes to normal levels compared to a matched control population.

The current study reports the results of a prospective, randomized, double blind, multi-center, placebo-controlled trial of CYT107 in 109 critically ill COVID-19 patients with lymphopenia. The primary endpoint of the study was to determine if CYT107 was more effective in restoring absolute lymphocyte counts versus placebo. An additional key study objective was to determine if CYT107 could be safely used in COVID-19 patients with severe respiratory failure without leading to further respiratory compromise or exacerbating systemic inflammation. Other aims of the study were to evaluate the effects of CYT107 on major clinical outcomes including ICU and hospital length of

stay, incidence of secondary hospital-acquired infections, and mortality as potentially indicative of putative beneficial effects of immune adjuvant therapy in this patient population with severe viral pneumonia and lymphopenia.

#### RESULTS

A total of 109 patients were enrolled in four different countries (Fig. 1 and supplemental Fig. 1) between May of 2020 and February of 2022. There were no significant differences between the CYT107 group and the placebo group in patient age, gender, BMI, race, co-morbidities, WHO or NEWS-2 clinical scores, C-reactive protein, ferritin, or D-Dimer at enrollment (Table 1). There was also no difference in baseline patient values for complete white blood cell count, absolute neutrophil count, or absolute monocyte at trial initiation; there was a small but statistically significant difference in baseline absolute lymphocyte counts (supplemental Table 1). The average number of doses of CYT107 received by patients was 3.29 doses (supplement Fig. 2). 62% of the patients enrolled were being mechanically ventilated at baseline or during the study. CYT107 or placebo were discontinued in 75 patients due to hospital discharge or death. Study enrollment was halted at 109 patients out of a planned enrollment of 220 patients because of difficulty in recruiting additional patients with the decline of the COVID-19 pandemic.

#### Lymphocyte counts increased in both CYT107 and placebo treated patients and were not different.

The effect of CYT107 on the absolute lymphocyte count was determined using a mixed model of repeated measures starting at multiple time points after initiation of CYT107 until day 30 or at hospital discharge if occurring before day 30 (34). The last absolute lymphocyte counts in the CYT107 and placebo-treated patients were 1,335 ( $\pm$ 136) and 1,125 ( $\pm$ 87) cells/µL (SEM) respectively, and not statistically different (Table 2). Importantly, 60% of COVID-19 patients receiving CYT107 were also receiving antiviral nucleoside analogs, predominantly remdesivir (supplemental Table 2). Lymphopenia is a well-known side effect of this class of drugs (35-37). Given this potential effect of antiviral therapy to inhibit CYT107's ability to induce lymphocyte proliferation and to increase lymphocyte counts, the effect of CYT107 on absolute lymphocyte counts was also analyzed separately in CYT107 and placebo-treated patients who did not receive concurrent antiviral therapy. COVID-19 patients who received CYT107 *without* concomitant nucleoside analogs had a final absolute lymphocyte count that was 43% greater than placebo-treated patients, i.e., 1,624 ( $\pm$ 28) versus 1,136 ( $\pm$ 119) cells/µL (SEM) respectively; *p*=0.067.

#### CYT107 was safe and did not exacerbate organ dysfunction

CYT107 was well tolerated and surprisingly there were fewer overall treatment emergent adverse events recorded in the CYT107 treated patients vs placebo, i.e. 195 versus 396 treatment emergent adverse events, respectively; p<0.0001 (Table 3). Noteworthy, the CYT107 group also experienced statistically fewer adverse events typical of COVID pathology such as renal (57% decrease), cardiac (40% decrease), central nervous system (81% decrease), and respiratory (45% decrease) (all p<0.05) (Table 3). One patient in the study developed a fever and had increased respiratory distress requiring intubation and mechanical ventilation after the second dose of the study drug. The treatment was halted and unblinding showed that the patient was receiving CYT107. The patient rapidly improved over the next 12-24 hours and was taken off the ventilator. The patient recovered uneventfully and was discharged from the ICU and subsequently from the hospital. Following this event, the DSMB recommended restarting the study without any protocol modifications. The study was continued with no further similar episodes noted.

#### CYT107 did not induce cytokine storm – no effect on circulating IL-6, TNF-α, or IL-10 cytokine levels

To determine the potential effect of CYT107 to impact levels of pro-inflammatory or anti-inflammatory cytokines, blood samples were collected at 6-7 hours and 20-22 hours after administration of the first two doses of CYT107 or placebo from approximately 60% of the patients. Results showed that CYT107 had no significant effect on circulating levels of IL-6, TNF- $\alpha$ , or IL-10 versus placebo (Fig. 2 and supplemental Figs.3,4).

Patients receiving CYT107 trended to shorter ICU length of stays versus placebo treated patients

Comparison of the number of ICU days in the CYT107 versus placebo group showed that there was no difference in the number of ICU days in the patients receiving CYT107 versus placebo, i.e. 31 days vs 45 days respectively, p=0.3366, (Fig. 3A). Because of potential effects of antiviral agents to inhibit the putative beneficial effects of CYT107, the number of ICU days was also calculated for patients receiving CYT107 or placebo but no additional antiviral drugs. Patients receiving CYT107 but no antiviral medications had a statistically significant decrease in their number of ICU days versus placebo treated patients not receiving antivirals, i.e., 24 days versus 45 days respectively; p=0.0482, (Fig. 3B).

#### Patients receiving CYT107 trended toward shorter hospital length of stays versus placebo-treated patients

Results for hospital length of stay showed similar trends as ICU length of stay. Hospitalization days in patients receiving CYT107 versus placebo, i.e. 29 days vs 45 days respectively, p=0.4096. (Fig. 3C). Comparison of the days of hospitalization in CYT107 versus placebo-treated patients who were not receiving antiviral agents was close to statistical significance at 28 days versus 45 days for CYT107 and placebo treated patients respectively, (Fig. 3D); p=0.0623).

#### Patients receiving CYT107 had fewer secondary infections vs placebo treated patients

The development of hospital-acquired secondary infections was evaluated by two groups of a three member panel of experienced intensive care physicians using the *Center for Disease Control* criteria for healthcare-associated infections (*https://www.cdc.gov/healthcare-associated-infections/index.html*) as previously described (31) – see supplementary section. The panel members were blinded to the patient identity and treatment group. Data showed that 28 of 55 (50.9%) versus 49 of 54 (90.7%) of CYT107 treated and placebo-treated patients respectively developed secondary infections, (*p*=0.013) (Table 4). The particular pathogens that were identified as the cause of the secondary hospital acquired infection in each individual patient include many opportunistic type organisms (Table 5).

Mortality rates - no difference in all-cause mortality in CYT107 vs placebo-treated patients

The mortality rate was calculated at day 30 or hospital discharge if patients left the hospital prior to day 30. The all-cause mortality was 32.7% in CYT107 treated patients (n=55) versus 38.9% in placebo treated patients, (n=54), and was not statistically different, (p=0.59), (Fig. 4A, Table 2). The individual patient cause of death as assigned by the treating physicians and then validated by pharmacovigilance is provided in supplemental Table 3. Because of potential effects of antiviral agents to inhibit the putative beneficial effects of CYT107 on lymphocyte proliferation, all-cause mortality was also determined separately for patients who received CYT107 or placebo alone (no antiviral drugs). All cause mortality in these patients was 22.7% (n=5/22) and 47.8% (n=11/23) for CYT107 and placebo groups respectively, (p=0.17), (Fig. 4B, Table 3). The mortality rate for patients being treated with both CYT107 and antiviral nucleoside inhibitors was 39.4% (n=13/33) while the mortality rate for patients being treated with both antiviral drug therapy) was 22.7% (n=5/22)., (Fig 4B, Table 2). The mortality rate for patients being treated with both antiviral nucleoside inhibitors and CYT107 or placebo were respectively 39.4% (n=13/33) and 31.3% (10/31), (p=0.94 NS). (Table 3).

#### DISCUSSION

Our study has particular importance as it represents one of the few clinical trials to test drug therapies that *enhance* rather than inhibit the host immune response in patients with COVID-19. Results demonstrated that recombinant human IL-7 (CYT107) which potently enhances adaptive immunity did not precipitate a cytokine storm and was safe and well-tolerated.

#### CYT107 clinical tolerability and safety

An important finding of the present study was the excellent clinical tolerability of CYT107 in critically ill COVID-19 patients. Patients receiving IL-7 did have a highly statistically significant decrease in treatment emergent adverse events (TEAEs), i.e. 195 versus 396 treatment emergent adverse events, respectively; p<0.0001. Many of these adverse events directly reflect COVID-19 pathology (Table 3). Therefore, this significant decrease in TEAEs is consistent with a putative beneficial effect of IL-7 in COVID-19 patients. With the exception of a single patient who had a transient reversible respiratory deterioration, there was no evidence that CYT107 worsened pulmonary function, precipitated hemodynamic instability, or increased levels of proinflammatory cytokines (Table 3, Fig. 2, supplemental Figs 3,4). Specifically, circulating levels of IL-6, TNF- $\alpha$ , and IL-10 were not different in CYT107 versus placebo treated patients after drug administration (Fig. 2, supplemental Figs 3,4). Interestingly, the levels of IL-6 and IL-10 were higher in the laboratory testing center in France compared to the United Kingdom and United States testing sites (Fig. 2, supplemental Figs 3,4). Potential reasons for this site variation include different reference standards used for instrument calibration, differences in aliquoting and freezing after blood draw, time of storage, and storage conditions . Importantly, there were no differences in the effect of CYT107 versus placebo to increase cytokine levels at the three independent testing sites (38,39).

The present results showing the safety of CYT107 in this patient population with severe COVID-19 are in agreement with a previous smaller study of 12 mechanically-ventilated critically-ill patients with COVID-19 treated with CYT107 (33). In the previous study, CYT107 was well tolerated without inducing changes in temperature, blood pressure, or PaO<sub>2</sub>:FiO<sub>2</sub> ratios. As in the present study, there were no detectable adverse clinical effects of CYT107 on any organ system. Increased clotting disorders including pulmonary emboli and

deep venous thrombosis are another common complication of COVID-19. In that regard, there was no increase in the incidence of venous thrombosis in the CYT107 treated group versus placebo. Furthermore, the day 30 D-Dimer levels in CYT107 versus placebo-treated patients were similar in the present trial, i.e., 3041.7 +/- 7478.9 vs 3088.2 +/- 3749.9 ng/mL respectively. It is important to note that because of requirements to verify that patients met the inclusion and exclusion criteria, e.g., decreased absolute lymphocyte counts, and the necessity of obtaining informed consent, CYT107 was typically not administered until 48-72 hours after hospital admission. This delay in administering CYT107 is important because studies in patients with sepsis show that levels of pro-inflammatory cytokines fall precipitously during the first 48-72 hours and are not much reduced compared to their peak concentrations. Thus, this delay in administration of CYT107 makes it unlikely that CYT107 was administered during the early more inflammatory phase of COVID-19. It is also important to note that CYT107 was administeried intramuscularly resulting in a slower uptake and better tolerability compared to intravenous administration (40). One likely explanation for the safety of CTY107 relates to the unique properties of the IL-7 receptor. Once activated by IL-7, the IL-7 receptor is internalized and non-responsive to additional doses of IL-7 for 48-72 hours (341). In addition, as T cells become more activated, IL-7 receptor expression decreases making IL-7 unable to over stimulate the activated lymphocytes. *Protection from hospital-acquired secondary infections:* 

Although patients with COVID-19 may die during the initial hyper-inflammatory cytokine storm mediated phase, many patients survive this early phase but subsequently develop nosocomial bacterial and fungal infections which are a major cause of morbidity and mortality (14-17). Unresolving secondary pneumonia has been linked to death in patients with COVID-19 (42). The high incidence of hospital acquired infections in COVID-19 patients is likely due in part to the effect of SARs-CoV-2 to weaken essential components of the immune system including by inducing profound lymphopenia which has been associated with increased risk of morbidity and mortality in patients with sepsis and COVID-19 (6,18,19,24, 43-45). IL-7 increases both the number as well as the functional activity of CD4 helper and CD8 cytotoxic T lymphocytes that play a critical role in anti-viral host defenses (27,28,32,46,47). The present results showing a major decrease in secondary infections in CYT107 treated COVID-19 patients are consistent with results of an earlier encouraging trial of CYT107 in 12 critically ill COVID-19 patients in which secondary infections occurred in seven (58%) COVID-19 patients treated with CYT107 versus 11 of 13 (85%) historical control COVID-19 control patients (33).

#### Potential mechanisms for the effect of CYT107 to decrease secondary infections

IL-7 has been termed the "maestro of the immune system" because of its myriad effects to orchestrate host immunity (25). Consequently, there are several potential mechanisms whereby CYT107 could have led to a decrease in secondary infections in critically ill COVID-19 patients. Despite the absence of a significant increase in circulating lymphocyte numbers, there is compelling historical evidence to suggest that CYT107 administration impact lymphocyte function and host protective immunity that would result in the more rapid containment and elimination of the invading SARS-CoV-2 virus (5,22,47,48). Early control of the initial viral infection could prevent secondary bacterial or fungal infections (17,18,20,21). IL-7 plays a critical role in shaping T cell responses to respiratory viral infections by increasing both the number and function of viral antigen-specific T cells including CD8 cytotoxic T cells (29-31,51,52). The importance of an early robust anti-viral antigen specific T cell response in COVID-19 is highlighted by Tan and colleagues who reported that the early induction of functional SAR-CoV-2 specific T cells was associated with mild disease and more rapid viral clearance (21). Similarly, Sattler et al. observed that COVID-19 patients who had a blunted T cell response to SARS-CoV-2 cell membrane, nucleocapsid, or spike protein antigens were more likely to die compared to COVID-19 patients who did mount a robust T cell response (49). Importantly, we previously reported that therapy with CYT107 in a critically ill patient with COVID-19 with lymphopenia and poor T cell response to SARS-CoV-2 antigens resulted in an approximate 3-fold increase in the number of IFN-y producing lymphocytes that were reactive to the SARS-CoV-2 nucleocapsid and/or spike protein antigens (50). The increase in the number of CYT107-induced SARS-CoV-2 reactive T cells was temporally associated with rapid clinical improvement and recovery in the patient (48). IL-7 not only increases the number of T cells directed against SARS-CoV-2 viral antigens but also increases T cell receptor diversity thereby potentially enhancing the intensity of the T cell response and lowering the risk of secondary infections. (30, 32,51). The trend toward shorter ICU days in COVID-19 patients treated with CYT107 is consistent with an effect of CYT107 to accelerate viral clearance (Fig. 3).

Additional mechanisms whereby IL-7 may act to decrease secondary nosocomial infections in COVID-19 patients is its effect to reverse T cell exhaustion which commonly occurs in patients with persistent viral infections including COVID-19 (52,53). Investigators have reported that CD4 and CD8 T cells from COVID-19 patients had

expression of multiple inhibitory molecules including PD-1, PD-L1, Tim-3, and CTLA-4 indicative of T cell exhaustion (8,21,46,52,53). Importantly, investigators have demonstrated that *ex vivo* treatment of COVID-19 patient blood samples with IL-7 caused restoration in T cell proliferation and IFN-γ production (5,22,50,54). Other mechanisms by which IL-7 could decrease secondary infections are its effects to activate innate lymphoid cells including mucosally associated invariant T (MAIT) cells which play a critical role in host defense against invading respiratory and gastrointestinal pathogens (48,55). Studies by Hubrack and colleagues showed that MAIT cells from COVID-19 patients have severely impaired function which was reversed by *ex vivo* treatment with IL-7 (48). These investigators concluded that IL-7 treatment might be an effective method to protect against SARS-CoV-2 infections.

#### CYT107's effect to increase lymphocyte counts - potential inhibition by nucleoside analogs

A surprising finding from the current study is the fact that both CYT107 and placebo-treated patients showed a similar increase in their absolute lymphocyte counts. IL-7 is a lymphocyte growth factor that has unfailingly been demonstrated to cause a dose-dependent increase in CD4 and CD8 T cells in numerous clinical trials in oncology/hematology patients and in patients with severe infections (29,31,32,51,55,56). Our previous trial of CYT107 in COVID-19 patients showed that the group receiving CY107 had an absolute lymphocyte count at day 30 that was more than 2-fold greater than the control group (33) Additionally, we previously reported that the absolute lymphocyte count in a COVID-19 patient who received CYT107 on a compassionate basis increased approximately 5-10 fold to a high of 5,100/µL which is considerably greater than 3,500/µL, the upper limit of normal for absolute lymphocyte counts (50). One potential reason for the lack of effect of CYT107 to increase patient lymphocyte counts compared to the placebo-treated group could be due to concomitant therapy with drugs that induce lymphopenia. Unlike previous CYT107 clinical studies in sepsis or COVID-19, 60% of patients treated with CYT107 in the present study received concurrent antiviral therapy with remdesivir or other drugs in its class that act as nucleoside analogs to inhibit nucleic acid polymerase (supplemental Table 3). Inhibition of RNA or DNA polymerase leads to accumulation of toxic purine degradation by-products that can induce cell death (35-37). Lymphocytes are the cells that are most sensitive to these toxic purine

degradation products and lymphopenia is a well-recognized complication of the use of this class of drugs (35-37).

#### Implications of the present study regarding the underlying pathogenesis of COVID-19

The findings from the present study have implications regarding the mechanisms responsible for morbidity and mortality in patients with COVID-19. Many investigators believe that most deaths in patients with COVID-19 are driven by an unbridled host immuno-inflammatory response to the invading virus (1-3). These investigators argue that there is strong evidence of end-organ damage without viral invasion. Unfortunately, there has been a relative paucity of autopsy studies that investigated the link between organ injury and the presence of viral load, likely because of initial concerns regarding the transmissibility of SARS-CoV-2. However, several groups have reported the detection of high viral loads in lung in COVID-19 postmortem specimens (11-14). Menter et al reported that postmortem examination of COVID-19 patients showed high levels of SARS-CoV-2 as detected by RT-gPCR in the lungs of 20 of 21 patients (12). Secondary superimposed bronchopneumonia was also present in 10 of the 21 patients underscoring the role of nosocomial infections. Similarly, Schaller et al conducted autopsies on 10 COVID-19 patients with severe acute respiratory distress syndrome and reported that SARS-CoV-2 was detectable by PCR in the respiratory tracts of all 10 patients (13). The most comprehensive postmortem study examining presence of SAR-CoV-2 in organ tissue samples was conducted by Stein and colleagues who did complete autopsies on 44 patients who died of COVID-19 (14). Results showed that SARS-CoV-2 was widely distributed, predominantly in patients who died with severe COVID-19, and that virus replication was present in multiple respiratory and non-respiratory tissues including the brain (14). Collectively, these studies are consistent with the concept that inability of the patients to successfully eradicate the invading SARS-CoV-2 results in ongoing viral-induced organ injury.

There is also increasing evidence that long COVID is associated with persistence of SARS-CoV-2 (57-59). Zuo and associates showed that tissue samples from a large cohort of patients who had seemingly recovered from COVID-19 had SARS-CoV-2 viral RNA present in multiple organs and blood for up to 4 months post infection (57). In addition, patients who had higher virus copy numbers were more likely to have symptoms consistent with long COVID. Findings from the present study do not directly address the critical question of whether the primary

abnormality in COVID-19 is due to a damaging host immuno-inflammatory response or a result of the failure of host immunity to contain SARS-CoV-2. However, the results showing that therapy with CYT107, a potent immune adjuvant, led to decreased secondary nosocomial infections supports the theory that failure to eliminate the SARS-CoV-2 is a key pathogenic mechanism underlying the ongoing morbidity and mortality.

#### Limitations

There are a number of limitations to the present investigation. One potential complicating factor is the evolution of the SARS-CoV-2 virus during the study with the emergence of different strains of the virus with varying degrees of virulence. Secondly, the study included COVID-19 patients from four different countries. Critical care management of patients such as respiratory ventilator support techniques were likely not identical at all sites. It is also important to note that the patients being treated with CYT107 in the present trial received fewer doses of the drug (average 3.29 doses) versus the typical 5-6 doses of CYT107 for the previous trials in sepsis and COVID-19. The lack of a more robust increase in the absolute lymphocyte count is likely partly related to this shorten course of treatment. Different clinical management protocols could impact ICU and hospital outcomes. Finally, the relatively small number of patients included in the present trial prevents any firm conclusions regarding the potential clinical efficacy of CYT107 in COVID-19 patients.

#### Conclusions

In conclusion, the findings from this study have particular significance because they represent one of the few studies to test drugs that *enhance* patient immunity in critically ill COVID-19 patients. Absolute lymphocyte counts increased in both groups without a statistically significant difference between CYT107 and placebo-treated patients. This lack of an effect of CYT107 to increase the absolute lymphocyte counts compared to placebo-treated patients, the primary endpoint of the study, may have been due in part to effects of concomitant use of antiviral drugs which can cause lymphopenia. Overall, CYT107 was well tolerated without any clinical or laboratory evidence of precipitating a cytokine storm or aggravating pulmonary inflammation in these COVID-19 patients. Specifically, there were significantly fewer treatment emergent adverse effects for most organ systems recognized as common targets of COVID-19 pathology, including the lung in the CYT107 treated group vs the

placebo group. Moreover, CYT107 decreased secondary hospital acquired infections by over 40% versus the placebo treated group. Given that hospital acquired infections are responsible for a large percentage of deaths in COVID-19 patients, this putative beneficial effect of CYT107 to decrease nosocomial infections could substantially reduce late morbidity and mortality in this highly lethal disease. The strong safety profile of CYT107 and its capacity to reduce secondary infections provide support for trials of CYT107 in other potential pandemic respiratory viral infections.

#### METHODS

#### (see supplemental materials for additional details)

#### Sex as a biological variable

The current study examined both male and female patients, similar results were found for both sexes.

#### Study design and participants

This prospective double blind, randomized, placebo-controlled phase 2b trial of CYT107 was designed as a single study but due to local regulatory constraints, it was conducted as individual cohorts in the United Kingdom, France, Brazil, and the United States using nearly identical protocols with minor exceptions – see supplemental materials for differences in protocols used. At study end, the data from all cohorts were combined for a statistical analysis with a single Statistical Analysis Plan (SAP).

The primary endpoint was an increase in the absolute lymphocyte count with CYT107 evaluated in the context of an acceptable safety profile. Secondary endpoints were clinical outcomes including: clinical improvement defined as an improvement of WHO 11-point ordinal scale from randomization through hospital discharge or Day 30, number of organ support free days during index hospitalization, incidence of secondary infections, number of days in the ICU and in the Hospital during index hospitalization, all-cause mortality through day 45.

Randomization and blinding:

Randomization assignment was assigned by the Electronic Data Capture system following data entry by the research team and informed consent with the investigator or PI approval for initiation of Study drug treatment. A permuted-block randomization was used (1-CYT107 + 1 Placebo) for an allocation ration of 1:1. Study drug or placebo were prepared at each site by the hospital pharmacy and delivered to the research team in a blinded fashion, who subsequently delivered them to ICU staff for administration to patients.

All study participants, clinical, and research teams were blinded to assignment of all but one study participant until after the final subject had completed treatment and all queries were resolved. There was one patient, as discussed above, where an emergency unblinding was required, this patient had been randomized to CYT107 and is included in all analyses.

#### Study deviations

IL-7 or placebo were discontinued in 75 patients due to hospital discharge or death (see Results section on patient safety). Study enrollment was halted at 109 patients out of a planned enrollment of 220 patients because of difficulty in recruiting additional patients as a result of the waning of the pandemic and rising costs of the trial. One patient who was randomized to placebo was treated with study drug.

#### Inclusion criteria (see supplement for additional details)

The main inclusion criteria were the following i) hospitalized men and women aged  $\geq$ 25-80 years of age, ii) COVID-19 diagnosed by any acceptable test available/utilized at each site; iii.) one absolute lymphocyte count (ALC)  $\leq$  1000 cells/mm<sup>3</sup> collected at hospitalization or no more than 72 hours after admission, iv.) hypoxemia requiring oxygen therapy at  $\geq$ 2L per minute nasal cannula or greater to keep saturations >90%, non-invasive positive pressure ventilation (e.g., BIPAP) or intubation with mechanical ventilatory support for respiratory failure; v.) signed informed consent by the patient or the patient's legally authorized representative. (see additional inclusion criteria for the oncology patients enrolled in the trial in supplemental material section).

#### Exclusion criteria:

The main exclusion criteria were the following: i) elevated liver function tests greater than five times upper limit of normal; ii) known or active autoimmune disease; iii) SOFA score  $\geq$  9 at baseline; iv.) history of organ transplantation; v.) patients with baseline Rockwood Clinical Frailty Scale  $\geq$  6. (**supplement Table 1**). Patients who were receiving antiviral therapies such as remdesivir, the IL-6 antagonist tocilizumab, and/or Janus kinase inhibitors were not excluded from enrollment.

#### Demographic reporting:

Demographic identities were classified on hospital admission. This was typically performed by either the patient or a patient representative (next of kin, family member, etc.) during intake to the hospital on a form which contained differing classifications and the option to write in a custom option. In the case of the patient being unresponsive and no viable representative being present, hospital staff would make initial determinations to be validated by patient/patient representatives at a later opportunity. No racial/ethnic classification was made for patients in the French cohort as French law typically forbids collection of racial/ethnic identity in most cases.

#### CYT107 dosing regimens:

Consistent with IL-7 receptor kinetic of reappearance (39), CYT107 was administered intramuscular twice a week. Due to concerns that CYT107 might exacerbate the early hyper-inflammatory phase of COVID-19, the first 12 patients who were randomized to receive CYT107 had an initial dose of 3 µg/kg of CYT107 followed by10 µg/kg of CYT107 for all subsequent doses. After Data Safety Monitoring Board (DSMB) confirmation that there was no evidence of any adverse effects in the these first 12 patients receiving the initial lower dose of CYT107, the initial dose of CYT107 was increased to 10 µg/kg of CYT107 for all subsequent patients who were randomized to the treatment arm. CYT107 was continued for a total of 3 to 4 weeks or until the patient was discharged from the hospital. Patients randomized to placebo received the same volume and dosing frequency of drug vehicle.

#### Quantitation of cytokines

Cytokine kits for TNF-α, IL6, and IL-10 were obtained from Biolegend and R&D Systems — see supplemental materials and methods. Assays were performed exactly as per the manufacturer's instructions and as previously described (31). Cytokines analyzed by 3 different laboratories in the UK, France, and US.

#### Development of secondary hospital-acquired infections

The development of hospital-acquired secondary infections was evaluated by a 3-member panel of experienced intensive care physicians in the US and a 3-member panel of experienced intensive care physicians in France using the Center for Disease Control criteria for healthcare-associated infections (*Center for Disease Control criteria for healthcare-associated infections*) as previously described (31). The panel members were blinded to patient identity and treatment group.

#### Statistical analysis

All study endpoints were analyzed on the ITT (intention to treat) population (patients randomized to CYT107 or placebo). All safety data were analyzed on the safety Population (Patients who received at least one injection of CYT107). Categorical outcomes were evaluated using Fisher's exact test. Continuous outcomes and especially absolute lymphocyte count were analyzed with linear mixed models for repeated measures incorporating treatment group, measurement day. Last ALC values were analyzed by *t* test for unequal variances. Survival, ICU and Hospital length of stay were analyzed by Kaplan Meier curve and a log rank (Mantel-Cox) test to compare the curves. Note that patients who died during their ICU or Hospital stay were censored at day 45 even if they died prior to that day. The ratio of the death rates with associated P-value is based on Poisson distribution. Statistical Analysis was performed in the MedCalc software suite (MedCalc Software Ltd, Belgium) except Log-Rand (Mantel-Cox) were performed in Graphpad Prism 10.3.1 (Graphpad Software LLC, Boston, MA, USA)

#### Study Registrations and Ethical approval

The study was conducted at major academic sites in the United Kingdom, France, Brazil, and the United States *-Trial registration*: NCT04379076, NCT04426201, NCT04442178, NCT04407689; NCT04927169. The regulatory agencies at each country (FDA / USA, MHRA / UK, ANSM /FR, CONEP/ Brazil) as well as centralized IRBs/EC or local ones when applicable approved the study. *United Kingdom regulatory approval #'s*: Eudract *#* 2020-001786-36; ISRCTN *#* ISRCTN15913068. French regulatory agency (ANSM) 2020-001573-78. French Ethical Committee number : 20-36/SI 20.04.14.45420. *Brazil* CONEP (37281020.91001.5505). United States, Missouri Baptist Hospital, St. Louis, Missouri, USA (protocol MOBAP1138). University of Florida, Gainesville, Florida, IRB2020000924. Cleveland Clinic, Cleveland, Oh, USA CCF IRB 3 20-938.

All patients or their legal representatives gave written informed consent prior to study inclusion.

#### Data availability statement

Data used for all analyses in the present study are included in a separate file "supporting data file.xlsx" which is included as supplemental materials. Patient information is de-identified.

#### Author contributions

MSH, BF, KR, CG, SP, TD, RS, SB, LM, CY, VV, MM, MVDB, and RH conceived and designed the study. MM, AHW, JB, FB, RJ, and RH analyzed data and maintained databases. MSH, BF, KR, CG, SP, TD, RS, GA, DS, RM, NA, JB, TB, SB, VV, and FDP provided clinical management and site coordination. All authors reviewed the manuscript and discussed results.

*Members of the Data Safety Monitoring Board:* Dr. Steve Opal, Dr. Elliott Crouser, Dr. Philippe Eggimann, Dr. Peter Pickkers.

Members of the Secondary Infection Adjudication Committee -

-United States and Brazil: Dr. John Mazuski, Dr. Brian Fuller, Dr. Isaiah Turnbull

-France and United Kingdom: Dr. Bruno Abraham, Dr. Marc Clavel, Dr. Valerie Gissot.

#### Acknowledgments

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#### Study funding

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**Conflict of Interest**: M. Morre and F Berbille are RevImmune employees.

## Table 1: Patient Clinical Characteristics – ITT

Clinical Characteristics	CYT107 (n=55)	Placebo (n=54)	P value
Age	N=55	N=54	
Mean	63.42±11.90	63.46±9.92	P=0.9830
Range	33-81	32-80	
Gender	N=55	N=54	
Male	42	42	P=0.8606
Female	12	12	
Weight (kg)	N=52	N=54	
Mean ± SD	92.79±20.43	90.54±20.30	P=0.5714
Range	54.7 – 144	38.3 – 155	
ВМІ	N=52	N=54	
Mean ± SD	31.33±5.47	30.41±5.88	P=0.4042
Range	22.76 - 48.36	17.72 - 47.05	
Race n(%)*			
White	40 (72.7	38(70.4)	p>0.9999
African American or Black	4 (7.3)	3 (5.6)	
Asian	3 (5.5)	3 (3.7)	
Ethnicity n(%)			
Hispanic or Latino	2 (3.6)	5 (9.3)	
Not Hispanic or Latino	48 (87.3)	42 (77.8)	
WHO Clinical Score	N=55	N=50	
Baseline	6.29±1.05	6.52±1.09	P=0.2756
D30 Hospital discharge	6.07±2.85	6.56±2.8.3	

Other Baseline			
Clinical Scores			
NEWS2 Score	6.33 ± 2.45	7.18 ± 2.32	P= 0.202
D-Dimer ng/mL	4688 ± 6882	5412 ± 15658	P= 0779
CRP	91.7 ± 81.2	104 ± 92	P= 0.500
Ferritin	1954 ± 1813	2021 ± 1914	P=0.870

\* Race/Ethnicity is not available for all patients; It is largely prohibited by French law to collect data relating to the race of patients.

		N	CYT107 Cell/uL and %	N	Placebo	%ALC increase CYT107 vs Placebo	p-value final ALC CYT107 vs Placebo	ODD ratio Death Rate CYT107 vs Placebo	P death rate
All patients	Final ALC	55	1,334	54	1124	18.7%			
	Death Rate		32.7%		38.9%			0.76	0.59
Antivirals	Final ALC	33	1,142	31	1,115	2.4%			
	Death Rate		39.4%		32.3%			1.37	0.94
No Antivirals	Final ALC	22	1,624	23	1136	43.0%	0.067		
	Death Rate		22.7%		47.8%			0.32	0.17

 Table 2. CYT107 effects on lymphocyte counts and mortality

60% of CYT107 treated patients received additional antiviral treatment (mostly Remdesivir) whereas 40% of patients received CYT107 treatment only. 57% of placebo treated patients received antiviral treatment whereas 43% of placebo treated patients received no antiviral therapies.

		CYT107		Plac		
		(n=	54)	(n=	55)	
		, ,	,	, , , , , , , , , , , , , , , , , , ,	,	
		Subject	Events	Subjects	Events	
System Organ	Severity	N (%)	E	N (%)	E	p-value
Class	-					Event
						rate
All TEAE's		44 (81.5)	195	50 (90.9)	396	<0.0001
	Mild	27 (50)	50	31 (56.4)	106	<0.0001
	Moderate	21 (38.9)	48	34 (61.8)	172	<0.0001
	Severe	31 (57.4)	78	35 (63.6)	90	0.42
	Unknown	11 (20.4)	19	16 (29)	28	0.2156
Blood and		11(20.4)	15	17(30.9)	33	0.0113
Lymphatic	Mild	3 (5.6)	5	7 (12.7)	19	0.0047
System	Moderate	4 (7.4)	4	6 (10.9)	9	0.1902
Disorders	Severe	5 (9.3)	6	5 (9.1)	5	0.7517
Cardiac		10 (18.5)	12	17 (30.9)	22	0.0999
Disorders	Mild	3 (5.6)	3	7 (12.7)	7	0.2375
	Moderate	3 (5.6)	4	7 (12.7)	8	0.2804
	Severe	5 (9.3)	5	6 (10.9)	7	0.6025
			0			0.0020
Renal and		5 (9.3)	5	11(20)	14	0.0454
Urinarv	Mild	0		4 (7.3)	5	0.0267
Disorders	Moderate	3 (5.6)	3	7 (12.7)	7	0.2375
	Severe	2 (3.7)	2	2 (3.6)	2	0.9862
Gastro-intestinal		9 (16.7)	9	12 (21.8)	16	0.1830
Disorders	Mild	4 (7.4)	4	4 (7.3)	5	0.7743
	Moderate	2 (3.7)	2	5 (9.1)	8	0.0696
	Severe	3 (5.5)	3	3 (5.5)	3	0.9828
General		19 (35.2)	19	20 (36.4)	49	0.0003
Disorders and	Mild	3 (5.6)	3	5 (9.1)	5	0.5240
Administration	Moderate	2(37)	2	7 (12 7)	34	<0.0001
Site Conditions	Severe	14 (25.9)	14	10 (18 2)	10	0 4505
		11 (20:0)			10	0.1000
Musculoskeletal		0	0	3 (5 5)	4	0 2260
and Connective	Mild	0	0	0	0	0.2200
Tissue	Moderate	0	0	3 (5 5)	4	0.0475
Disorders	Severe	0	0	0	0	0.0170
				Ť		
Nervous System		3 (5 6)	3	13 (23.6)	16	0.0029
Disorders	Mild	2 (3 7	2	1 (1 8)	1	0.61/7
	Moderate	2 (3.7	0	1 (1.0)	6	0.0147
	wouerate	U	U	<u> +(1.3)</u>	U	0.0102

## TABLE 3. Treatment Emergent Adverse Events by Organ Class - Safety population

	Severe	1 (1.8)	1	8 (14.5)	9	0.0126
Respiratory						
Thoracic and		22 (40.7)	28	28 (50.9)	49	0.0209
Mediastinal	Mild	4 (7.4)	5	3 (5.5)	4	0.7337
Disorders	Moderate	5 (9.3)	6	13 (23.6)	19	0.0106
	Severe	16 (29.6)	17	19 (34.5)	26	0.1938

## Table 4. Adjudicated Opportunistic Infections

	CYT107		Placebo	p-value	
	Infections	Total Patients	Infections	Total Patients	
All	28	55	49	54	0.0135
No antivirals	13	22	23	23	0.1288
Antivirals	15	33	26	31	0.0572

Significantly fewer patients with secondary infections in CYT107 group:

CYT107: 28/55 (51%)

Placebo: 49/54 (91%)

p-value: 0.0135

Study arm	Site of Infection	Infectious Agent
CYT107		Strenotrophomonas maltophilia and
	Colitis + Lower Respiratory Tract Infection	Klebsiella pneumoniae
CYT107	Bacteremia	Staph. epidermis
CYT107	bacterial Chest infection	Pseudomonas aeroguinosa
CYT107	bacterial Chest infection	Enterococcus faecalis (BAL)
CYT107		Mycoplasma pneumoniae
		agglutination test - positive
	bacterial Chest infection	(Titre>1280)
CYT107		Klebsiella pneumoniae, Citrobacter
	bacterial Chest infection+ Bacteremia	koseri, Gram negative bacilli seen
CYT107		Enterococcus faecalis,
	Dialysis cath Bacteremia + Pneumonia	Staphylococcus epidermidis
CYT107	Pulmonary + Urinary source Bacteremia	Enterobacter aerogenes
CYT107	Pulmonary Bacterial + septic shock	NA
CYT107	Pulmonary Bacterial + peritonitis + sepsis	Pseudomonas aeruginosa
CYT107	Pulmonary VAP	Sputum Culture - Kleb. pneumoniae
CYT107	Pulmonary VAP	E-Coli
CYT107	Pulmonary VAP	Streptococcus constellatus
CYT107		Staphylococcus aureus,
		Haemophilus influenzae, Raoultella
	Pulmonary VAP + Bacteremia	planticola
CYT107	Pulmonary VAP + urinary	Pseudomonas aeruginosa
01/74 07	Liringry Li guyar Daspiratory Trast Infaction	Staphylococcus lugdupopsis
CY1107	Unitary + Lower Respiratory Tract Intection	Staphylococcus luguuriensis
CY1107	Officially + Lower Respiratory fract infection	
Placebo	Officially + Lower Respiratory fract infection	Klebsiella aerogenes (BAL)
Placebo	bacteremia + viremia + Chest infection	Klebsiella aerogenes (BAL) Pseudomonas aeruginosa
Placebo Placebo	bacteremia + viremia + Chest infection Bacteremia + HSV	Klebsiella aerogenes (BAL) Pseudomonas aeruginosa Enterococcus faecalis
Placebo Placebo Placebo	bacteremia + viremia + Chest infection Bacteremia + HSV	Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip
Placebo Placebo Placebo	bacteremia + viremia + Chest infection Bacteremia + HSV Bacteremia + Mouth Fungal	Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip         infection
Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection Bacteremia + HSV Bacteremia + Mouth Fungal	Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip infection         Streptococcus anginosus
Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection Bacteremia + HSV Bacteremia + Mouth Fungal	Klebsiella aerogenes (BAL)       Pseudomonas aeruginosa       Enterococcus faecalis       Klebsiella pneumoniae, catheter tip infection       Streptococcus anginosus ;Staphylococcus haemolyticus;
Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection Bacteremia + HSV Bacteremia + Mouth Fungal	Staphylococcus luguriensis         Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip infection         Streptococcus anginosus ;Staphylococcus haemolyticus; Enterococcus faecalis; Enterobacter
Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection Bacteremia + HSV Bacteremia + Mouth Fungal	Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip infection         Streptococcus anginosus ;Staphylococcus haemolyticus; Enterococcus faecalis; Enterobacter cloacae, Prevottela bucae;
Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection         Bacteremia + HSV         Bacteremia + Mouth Fungal         Bacteremia + Pulmonary VAP + Sinus	Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip infection         Streptococcus anginosus ;Staphylococcus haemolyticus; Enterococcus faecalis; Enterobacter cloacae, Prevottela bucae; Fusobacterium nucleatum
Placebo Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection         Bacteremia + HSV         Bacteremia + Mouth Fungal         Bacteremia + Pulmonary VAP + Sinus         bacterial Chest infection	Staphylococcus luguriensis         Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip infection         Streptococcus anginosus ;Staphylococcus haemolyticus; Enterococcus faecalis; Enterobacter cloacae, Prevottela bucae; Fusobacterium nucleatum         Klebsiella pneumoniae in non-directed bronchial lavage
Placebo Placebo Placebo Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection Bacteremia + HSV Bacteremia + Mouth Fungal Bacteremia + Pulmonary VAP + Sinus bacterial Chest infection	Staphylococcus luguriensis         Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip infection         Streptococcus anginosus ;Staphylococcus haemolyticus; Enterococcus faecalis; Enterobacter cloacae, Prevottela bucae; Fusobacterium nucleatum         Klebsiella pneumoniae in non-directed bronchial lavage         Stenotrophomonas maltophilia
Placebo Placebo Placebo Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection         Bacteremia + HSV         Bacteremia + Mouth Fungal         Bacteremia + Pulmonary VAP + Sinus         bacterial Chest infection         bacterial Chest infection	Staphylococcus luguriensis         Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip         infection         Streptococcus anginosus         ;Staphylococcus haemolyticus;         Enterococcus faecalis; Enterobacter         cloacae, Prevottela bucae;         Fusobacterium nucleatum         Klebsiella pneumoniae in non-directed         bronchial lavage         Stenotrophomonas maltophilia         (BAL)
Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection         Bacteremia + HSV         Bacteremia + Mouth Fungal         Bacteremia + Pulmonary VAP + Sinus         bacterial Chest infection         bacterial Chest infection + gastroenteritis	Staphylococccus luguriensis         Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip infection         Streptococcus anginosus ;Staphylococcus haemolyticus;         Enterococcus faecalis; Enterobacter cloacae, Prevottela bucae;         Fusobacterium nucleatum         Klebsiella pneumoniae in non-directed bronchial lavage         Stenotrophomonas maltophilia (BAL)         Klebsiella Oxytoca in sputum
Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection         Bacteremia + HSV         Bacteremia + Mouth Fungal         Bacteremia + Pulmonary VAP + Sinus         bacterial Chest infection         bacterial Chest infection         bacterial Chest infection + gastroenteritis         Pneumonia	Staphylococcus luguriensisKlebsiella aerogenes (BAL)Pseudomonas aeruginosaEnterococcus faecalisKlebsiella pneumoniae, catheter tipinfectionStreptococcus anginosus;Staphylococcus haemolyticus;Enterococcus faecalis; Enterobactercloacae, Prevottela bucae;Fusobacterium nucleatumKlebsiella pneumoniae in non-directedbronchial lavageStenotrophomonas maltophilia(BAL)Klebsiella Oxytoca in sputumMRSA, Pseudomonas, yeast
Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection         Bacteremia + HSV         Bacteremia + Mouth Fungal         Bacteremia + Pulmonary VAP + Sinus         bacterial Chest infection         bacterial Chest infection         bacterial Chest infection + gastroenteritis         Pneumonia	Staphylococccus luguriensis         Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip         infection         Streptococcus anginosus         ;Staphylococcus haemolyticus;         Enterococcus faecalis; Enterobacter         cloacae, Prevottela bucae;         Fusobacterium nucleatum         Klebsiella pneumoniae in non-directed         bronchial lavage         Stenotrophomonas maltophilia         (BAL)         Klebsiella Oxytoca in sputum         MRSA, Pseudomonas, yeast         H. Influenza A, CMV
Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection         Bacteremia + HSV         Bacteremia + Mouth Fungal         Bacteremia + Pulmonary VAP + Sinus         bacterial Chest infection         bacterial Chest infection         bacterial Chest infection + gastroenteritis         Pneumonia         Pneumonia         Pulmonary + Urinary + bacteremai+	Staphylococcus luguriensisKlebsiella aerogenes (BAL)Pseudomonas aeruginosaEnterococcus faecalisKlebsiella pneumoniae, catheter tipinfectionStreptococcus anginosus;Staphylococcus haemolyticus;Enterococcus faecalis; Enterobactercloacae, Prevottela bucae;Fusobacterium nucleatumKlebsiella pneumoniae in non-directedbronchial lavageStenotrophomonas maltophilia(BAL)Klebsiella Oxytoca in sputumMRSA, Pseudomonas, yeastH. Influenza A, CMVStaphylococcus aureus
Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection         Bacteremia + HSV         Bacteremia + Mouth Fungal         Bacteremia + Mouth Fungal         Bacteremia + Pulmonary VAP + Sinus         bacterial Chest infection         bacterial Chest infection         bacterial Chest infection + gastroenteritis         Pneumonia         Pneumonia         Pulmonary + Urinary + bacteremai+         Pulmonary + Urinary bacterial infection	Staphylococcus luguriensis         Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip infection         Streptococcus anginosus ;Staphylococcus haemolyticus;         Enterococcus faecalis; Enterobacter cloacae, Prevottela bucae;         Fusobacterium nucleatum         Klebsiella pneumoniae in non-directed bronchial lavage         Stenotrophomonas maltophilia (BAL)         Klebsiella Oxytoca in sputum         MRSA, Pseudomonas, yeast         H. Influenza A, CMV         Staphylococcus aureus         Klebsiella pneumoniae
Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection         Bacteremia + HSV         Bacteremia + Mouth Fungal         Bacteremia + Mouth Fungal         Bacteremia + Pulmonary VAP + Sinus         bacterial Chest infection         bacterial Chest infection         bacterial Chest infection + gastroenteritis         Pneumonia         Pulmonary + Urinary + bacteremai+         Pulmonary + Urinary bacterial infection	Staphylococcus luguriensisKlebsiella aerogenes (BAL)Pseudomonas aeruginosaEnterococcus faecalisKlebsiella pneumoniae, catheter tipinfectionStreptococcus anginosus;Staphylococcus haemolyticus;Enterococcus faecalis; Enterobactercloacae, Prevottela bucae;Fusobacterium nucleatumKlebsiella pneumoniae in non-directedbronchial lavageStenotrophomonas maltophilia(BAL)Klebsiella Oxytoca in sputumMRSA, Pseudomonas, yeastH. Influenza A, CMVStaphylococcus aureusKlebsiella pneumoniae
Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo Placebo	bacteremia + viremia + Chest infection         Bacteremia + HSV         Bacteremia + Mouth Fungal         Bacteremia + Mouth Fungal         Bacteremia + Pulmonary VAP + Sinus         bacterial Chest infection         bacterial Chest infection         bacterial Chest infection + gastroenteritis         Pneumonia         Pneumonia         Pulmonary + Urinary + bacteremai+         Pulmonary bacterial infection	Staphylococcus lagumensis         Klebsiella aerogenes (BAL)         Pseudomonas aeruginosa         Enterococcus faecalis         Klebsiella pneumoniae, catheter tip infection         Streptococcus anginosus ;Staphylococcus haemolyticus;         Enterococcus faecalis; Enterobacter cloacae, Prevottela bucae;         Fusobacterium nucleatum         Klebsiella pneumoniae in non-directed bronchial lavage         Stenotrophomonas maltophilia (BAL)         Klebsiella Oxytoca in sputum         MRSA, Pseudomonas, yeast         H. Influenza A, CMV         Staphylococcus aureus         Klebsiella pneumoniae

Placebo	Pulmonary bacterial infection	Streptococcus pneumoniae
Placebo		Haemophilus influenzae Escherichia
	Pulmonary VAP	coli
Placebo		Staphylococcus aureus, Klebsiella
	Pulmonary VAP	aerogenes
Placebo		bacilles gram négatif + levures
	Pulmonary VAP	Candida albicans
Placebo	Pulmonary VAP	Hafnia alvei
Placebo		Pneumonia associated with
	Pulmonary VAP	mechanical ventilation
Placebo	Pulmonary VAP + Cutaneous viral	Stenotrophomonas maltophilia
Placebo		Citrobacter SP, Pseudomonas
	Pulmonary VAP + HSV	aeruginosa
Placebo	Tracheal infection + Bacteremia	MSSA
Placebo		Citrobacter koseri, Prevotella
	Tracheal site wound infection	denticola, and Prevotella nigrescens
Placebo	Tracheitis	MRSA
Placebo	Urinary	Escherichia coli
Placebo		Cytomegalovirus (viral load in
	Viraemia	bloods)

Table 5. Secondary Infections in CYT107 and Placebo treated patients.Abbreviations: BAL-Broncho-AlveolarLavage, VAP-Ventilator Associated Pneumonia, HSV, Herpes simplex, CMV-Cytomegalovirus, MRSA-Methicillinresistant Staphylococcus aureus, MSSA-Methicillin sensitive Staphylococcus aureus

#### REFERENCES

- 1. Cron RQ. COVID-19 cytokine storm: targeting the appropriate cytokine. *Lancet Rheumatol.* 2021;3(4):e236-e7.
- Chalmers JD, and Chotirmall SH. Rewiring the Immune Response in COVID-19. *Am J Respir Crit Care Med.* 2020;202(6):784-6.
- 3. Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS, Manson JJ, et al. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet.* 2020;395(10229):1033-4.
- 4. Looi MK. What is the future for covid drugs and treatments? *BMJ*. 2023;381:1001.
- Remy KE, Mazer M, Striker DA, Ellebedy AH, Walton AH, Unsinger J, et al. Severe immunosuppression and not a cytokine storm characterizes COVID-19 infections. *JCI Insight*. 2020;5(17).
- Vardhana SA, and Wolchok JD. The many faces of the anti-COVID immune response. *J Exp Med.* 2020;217(6).
- Tian W, Zhang N, Jin R, Feng Y, Wang S, Gao S, et al. Immune suppression in the early stage of COVID-19 disease. *Nat Commun.* 2020;11(1):5859.
- Mathew D, Giles JR, Baxter AE, Oldridge DA, Greenplate AR, Wu JE, et al. Deep immune profiling of COVID-19 patients reveals distinct immunotypes with therapeutic implications. *Science*. 2020;369(6508).
- Kusnadi A, Ramirez-Suastegui C, Fajardo V, Chee SJ, Meckiff BJ, Simon H, et al. Severely ill COVID-19 patients display impaired exhaustion features in SARS-CoV-2-reactive CD8(+) T cells. *Sci Immunol.* 2021;6(55).
- 10. Fajnzylber J, Regan J, Coxen K, Corry H, Wong C, Rosenthal A, et al. SARS-CoV-2 viral load is associated with increased disease severity and mortality. *Nat Commun.* 2020;11(1):5493.
- 11. Gutmann C, Takov K, Burnap SA, Singh B, Ali H, Theofilatos K, et al. SARS-CoV-2 RNAemia and proteomic trajectories inform prognostication in COVID-19 patients admitted to intensive care. *Nat Commun.* 2021;12(1):3406.
- Menter T, Haslbauer JD, Nienhold R, Savic S, Hopfer H, Deigendesch N, et al. Postmortem examination of COVID-19 patients reveals diffuse alveolar damage with severe capillary congestion and variegated findings in lungs and other organs suggesting vascular dysfunction. *Histopathology*. 2020;77(2):198-209.
- 13. Schaller T, Hirschbuhl K, Burkhardt K, Braun G, Trepel M, Markl B, et al. Postmortem Examination of Patients With COVID-19. *JAMA*. 2020;323(24):2518-20.
- 14. Stein SR, Ramelli SC, Grazioli A, Chung JY, Singh M, Yinda CK, et al. SARS-CoV-2 infection and

persistence in the human body and brain at autopsy. *Nature*. 2022;612(7941):758-63.

- 15. Trevelin SC, Pickering S, Todd K, Bishop C, Pitcher M, Garrido Mesa J, et al. Disrupted Peyer's Patch Microanatomy in COVID-19 Including Germinal Centre Atrophy Independent of Local Virus. *Front Immunol.* 2022;13:838328.
- De Bruyn A, Verellen S, Bruckers L, Geebelen L, Callebaut I, De Pauw I, et al. Secondary infection in COVID-19 critically ill patients: a retrospective single-center evaluation. *BMC Infect Dis.* 2022;22(1):207.
- 17. Grasselli G, Scaravilli V, Mangioni D, Scudeller L, Alagna L, Bartoletti M, et al. Hospital-Acquired Infections in Critically III Patients With COVID-19. *Chest.* 2021;160(2):454-65.
- 18. Ripa M, Galli L, Poli A, Oltolini C, Spagnuolo V, Mastrangelo A, et al. Secondary infections in patients hospitalized with COVID-19: incidence and predictive factors. *Clin Microbiol Infect.* 2021;27(3):451-7.
- Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet.* 2020;395(10229):1054-62.
- Angioni R, Sanchez-Rodriguez R, Munari F, Bertoldi N, Arcidiacono D, Cavinato S, et al. Age-severity matched cytokine profiling reveals specific signatures in Covid-19 patients. *Cell Death Dis.* 2020;11(11):957.
- 21. Tan L, Wang Q, Zhang D, Ding J, Huang Q, Tang YQ, et al. Lymphopenia predicts disease severity of COVID-19: a descriptive and predictive study. *Signal Transduct Target Ther.* 2020;5(1):33.
- 22. Tan AT, Linster M, Tan CW, Le Bert N, Chia WN, Kunasegaran K, et al. Early induction of functional SARS-CoV-2-specific T cells associates with rapid viral clearance and mild disease in COVID-19 patients. *Cell Rep.* 2021;34(6):108728.
- Bidar F, Hamada S, Gossez M, Coudereau R, Lopez J, Cazalis MA, et al. Recombinant human interleukin-7 reverses T cell exhaustion ex vivo in critically ill COVID-19 patients. *Ann Intensive Care.* 2022;12(1):21.
- 24. Vardhana S, Baldo L, Morice WG, 2nd, and Wherry EJ. Understanding T cell responses to COVID-19 is essential for informing public health strategies. *Sci Immunol.* 2022;7(71):eabo1303.
- 25. Alipoor SD, Mortaz E, Jamaati H, Tabarsi P, Bayram H, Varahram M, et al. COVID-19: Molecular and Cellular Response. *Front Cell Infect Microbiol.* 2021;11:563085.
- 26. Sprent J, and Surh CD. Interleukin 7, maestro of the immune system. *Semin Immunol.* 2012;24(3):149-50.
- 27. Barata JT, Durum SK, and Seddon B. Flip the coin: IL-7 and IL-7R in health and disease. *Nat Immunol.* 2019;20(12):1584-93.
- 28. Sheikh A, Jackson J, Shim HB, Yau C, Seo JH, and Abraham N. Selective dependence on IL-7 for antigen-specific CD8 T cell responses during airway influenza infection. *Sci Rep.* 2022;12(1):135.
- 29. Pellegrini M, Calzascia T, Toe JG, Preston SP, Lin AE, Elford AR, et al. IL-7 engages multiple

mechanisms to overcome chronic viral infection and limit organ pathology. Cell. 2011;144(4):601-13.

- 30. Sereti I, Dunham RM, Spritzler J, Aga E, Proschan MA, Medvik K, et al. IL-7 administration drives T cell-cycle entry and expansion in HIV-1 infection. *Blood.* 2009;113(25):6304-14.
- Perales MA, Goldberg JD, Yuan J, Koehne G, Lechner L, Papadopoulos EB, et al. Recombinant human interleukin-7 (CYT107) promotes T-cell recovery after allogeneic stem cell transplantation. *Blood.* 2012;120(24):4882-91.
- 32. Mazzucchelli R, and Durum SK. Interleukin-7 receptor expression: intelligent design. *Nat Rev Immunol.* 2007;7(2):144-54.
- Levy Y, Sereti I, Tambussi G, Routy JP, Lelievre JD, Delfraissy JF, et al. Effects of recombinant human interleukin 7 on T-cell recovery and thymic output in HIV-infected patients receiving antiretroviral therapy: results of a phase I/IIa randomized, placebo-controlled, multicenter study. *Clin Infect Dis.* 2012;55(2):291-300.
- Laterre PF, Francois B, Collienne C, Hantson P, Jeannet R, Remy KE, et al. Association of Interleukin 7 Immunotherapy With Lymphocyte Counts Among Patients With Severe Coronavirus Disease 2019 (COVID-19). JAMA Netw Open. 2020;3(7):e2016485.
- Detry MA, and Ma Y. Analyzing Repeated Measurements Using Mixed Models. *JAMA*. 2016;315(4):407-8.
- 36. Chien M, Anderson TK, Jockusch S, Tao C, Li X, Kumar S, et al. Nucleotide Analogues as Inhibitors of SARS-CoV-2 Polymerase, a Key Drug Target for COVID-19. *J Proteome Res.* 2020;19(11):4690-7.
- 37. Ali RA, Awadalla EA, Amin YA, Fouad SS, Ahmed MAB, Hassan MH, et al. The deleterious effects of sofosbuvir and ribavirin (antiviral drugs against hepatitis C virus) on different body systems in male albino rats regarding reproductive, hematological, biochemical, hepatic, and renal profiles and histopathological changes. *Sci Rep.* 2024;14(1):5682.
- 38. Cianci R, Massaro MG, De Santis E, Totti B, Gasbarrini A, Gambassi G, et al. Changes in Lymphocyte Subpopulations after Remdesivir Therapy for COVID-19: A Brief Report. *Int J Mol Sci.* 2023;24(19).
- Gao CA, Markov NS, Stoeger T, Pawlowski A, Kang M, Nannapaneni P, et al. Machine learning links unresolving secondary pneumonia to mortality in patients with severe pneumonia, including COVID-19. *J Clin Invest.* 2023;133(12).
- Zhou X, Fragala MS, McElhaney JE, and Kuchel GA. Conceptual and methodological issues relevant to cytokine and inflammatory marker measurements in clinical research. *Curr Opin Clin Nutr Metab Care*. 2010;13(5):541-7.
- 41. Gottfried-Blackmore A, Rubin SJS, Bai L, Aluko S, Yang Y, Park W, et al. Effects of processing conditions on stability of immune analytes in human blood. *Sci Rep.* 2020;10(1):17328.
- 42. Chen Z, and John Wherry E. T cell responses in patients with COVID-19. *Nat Rev Immunol.* 2020;20(9):529-36.
- 43. Adrie C, Lugosi M, Sonneville R, Souweine B, Ruckly S, Cartier JC, et al. Persistent lymphopenia is a

risk factor for ICU-acquired infections and for death in ICU patients with sustained hypotension at admission. *Ann Intensive Care.* 2017;7(1):30.

- 44. Wesche DE, Lomas-Neira JL, Perl M, Chung CS, and Ayala A. Leukocyte apoptosis and its significance in sepsis and shock. *J Leukoc Biol.* 2005;78(2):325-37.
- 45. Drewry AM, Samra N, Skrupky LP, Fuller BM, Compton SM, and Hotchkiss RS. Persistent lymphopenia after diagnosis of sepsis predicts mortality. *Shock.* 2014;42(5):383-91.
- 46. Bekele Y, Sui Y, and Berzofsky JA. IL-7 in SARS-CoV-2 Infection and as a Potential Vaccine Adjuvant. *Front Immunol.* 2021;12:737406.
- Hubrack S, Al-Nesf MA, Agrebi N, Raynaud C, Khattab MA, Thomas M, et al. In vitro Interleukin-7 treatment partially rescues MAIT cell dysfunction caused by SARS-CoV-2 infection. *Sci Rep.* 2021;11(1):14090.
- Sattler A, Angermair S, Stockmann H, Heim KM, Khadzhynov D, Treskatsch S, et al. SARS-CoV-2-specific T cell responses and correlations with COVID-19 patient predisposition. *J Clin Invest.* 2020;130(12):6477-89.
- 49. Mazer MB, Turnbull IR, Miles S, Blood TM, Sadler B, Hess A, et al. Interleukin-7 Reverses Lymphopenia and Improves T-Cell Function in Coronavirus Disease 2019 Patient With Inborn Error of Toll-Like Receptor 3: A Case Report. *Crit Care Explor.* 2021;3(7):e0500.
- 50. Sportes C, Hakim FT, Memon SA, Zhang H, Chua KS, Brown MR, et al. Administration of rhIL-7 in humans increases in vivo TCR repertoire diversity by preferential expansion of naive T cell subsets. *J Exp Med.* 2008;205(7):1701-14.
- 51. Aghbash PS, Eslami N, Shamekh A, Entezari-Maleki T, and Baghi HB. SARS-CoV-2 infection: The role of PD-1/PD-L1 and CTLA-4 axis. *Life Sci.* 2021;270:119124.
- 52. Barnova M, Bobcakova A, Urdova V, Kosturiak R, Kapustova L, Dobrota D, et al. Inhibitory immune checkpoint molecules and exhaustion of T cells in COVID-19. *Physiol Res.* 2021;70(S2):S227-S47.
- 53. Mazer MB, Davitt E, Turnbull IR, Caldwell CC, Brakenridge SC, Remy KE, et al. In Vitro-Administered Dexamethasone Suppresses T Cell Function With Reversal by Interleukin-7 in Coronavirus Disease 2019. *Crit Care Explor.* 2021;3(4):e0378.
- Sortino O, Dias J, Anderson M, Laidlaw E, Leeansyah E, Lisco A, et al. Preserved Mucosal-Associated Invariant T-Cell Numbers and Function in Idiopathic CD4 Lymphocytopenia. *J Infect Dis.* 2021;224(4):715-25.
- Sereti I, Estes JD, Thompson WL, Morcock DR, Fischl MA, Croughs T, et al. Decreases in colonic and systemic inflammation in chronic HIV infection after IL-7 administration. *PLoS Pathog.* 2014;10(1):e1003890.
- 56. Francois B, Jeannet R, Daix T, Walton AH, Shotwell MS, Unsinger J, et al. Interleukin-7 restores lymphocytes in septic shock: the IRIS-7 randomized clinical trial. *JCI Insight.* 2018;3(5).
- 57. Zuo W, He D, Liang C, Du S, Hua Z, Nie Q, et al. The persistence of SARS-CoV-2 in tissues and its

association with long COVID symptoms: a cross-sectional cohort study in China. *Lancet Infect Dis.* 2024;24(8):845-55.

- 58. Proal AD, VanElzakker MB, Aleman S, Bach K, Boribong BP, Buggert M, et al. SARS-CoV-2 reservoir in post-acute sequelae of COVID-19 (PASC). *Nat Immunol.* 2023;24(10):1616-27.
- 59. Buonsenso D, and Tantisira KG. Long COVID and SARS-CoV-2 persistence: new answers, more questions. *Lancet Infect Dis.* 2024;24(8):796-8.

Figure 1.



**Figure 1**. *Trial enrollment details.* Details on patient enrollment, allocation, stratification and number analyzed are presented. Hospitalized patients with respiratory distress who were positive for SARS-CoV-2 were identified. Patients were further screened for eligibility as per Inclusion and Exclusion criteria.







**Figure 2**. *CYT107 did not increase circulating levels of IL-6*. Blood samples were collected from patients at 6-7 hours and 20-22 hours after the first and second doses of CYT107 or placebo. Results showed that circulating levels of IL-6 were not different in patients receiving CYT107 versus placebo and represent averaged data (**A**.) and individually plotted data (**B**.). Please see supplemental figures 3 and 4 for results for IL-10 and TNF- $\alpha$ . Error bars represent SEMs. *Abbreviations*. FR, France; US, United States, UK, United Kingdom; D0, day zero, D3, day 3.



**Figure 3**. *Effects of CYT107 on ICU and hospital length of stay.* Comparison of the number of ICU days in the CYT107 versus placebo group showed that there was no difference in the number of ICU days in the patients receiving CYT107 versus placebo, i.e. 31 days vs 45 days respectively, *p*=0.3366, (**A**). Because of potential effects of antiviral agents to inhibit the putative beneficial effects of CYT107, the number of ICU days were also calculated for patients receiving CYT107 or but no antiviral drugs versus placebo treated patients who were not receiving antiviral drugs. Patients receiving CYT107 but no antiviral medications did have a statistically significant decrease in their number of ICU days, i.e., 24

days versus 45 days respectively; p=0.0482, (**B**). Results for hospital length of stay showed similar trends as ICU length of stay. Hospitalization days in patients receiving CYT107 versus placebo, i.e. 29 days vs 45 days respectively, p=0.4096. (**C**). Comparison of the days of hospitalization in CYT107 treated patients who were not receiving antiviral agents was close to statistical significance at 28 days versus 45 days for CYT107 and placebo treated patients respectively, (**D**); p=0.0623). Survival, ICU and hospital length of stay were analyzed with a log rank (Mantel-Cox) test to compare the curves. Note that patients who died during their ICU or Hospital stay were censored at day 45 even if they died prior to that day. Comparison of ICU and hospital length of stay was performed using the Log-Rank (Mantel-Cox) test. \*p<0.05.



**Figure 4**. *No difference in mortality in CYT107 vs placebo-treated patients*. The mortality rate was calculated at day 30 or hospital discharge if patients left the hospital prior to day 30. The overall mortality was 32.7% in CYT107 treated patients (n=55) versus 38.9% in placebo treated patients, (n=54), and was not

statistically different, p=0.59 (**A**, **Table 3**). Because of potential effects of antiviral agents to inhibit the putative beneficial effects of CYT107, mortality rates were also determined separately for patients receiving CYT107 but no antiviral drugs and similarly for placebo treated patients who were not receiving antiviral drugs. Mortality for CYT107 treated patients (n=22) and placebo treated patients (n=23) was 22.7% and 47.8% respectively, p=0.17) (**B**, **Table 3**). Comparison of mortality was performed using the Log-Rank (Mantel-Cox) test.

## **ENROLLMENT PER COUNTRY**



Supplemental Figure 1. Patient enrollment in each individual county. A total of 109 patients were enrolled in the United Kingdom, France, the United States, and Brazil. There were slightly different inclusion and exclusion criteria for the COVID patients enrolled in the oncology cohort versus patients who did not have active cancer. Despite the slightly different inclusion and exclusion criteria for the oncology patient cohort, the treatment regimens were identical for both groups. Abbreviations. UK, United Kingdom, US Oncol, United States oncology cohort.



## Distribution of patients per #doses received

**Supplemental Figure 2**. Effective treatment – number of doses of CYT107 administered. The distribution of patients based upon the total number of doses of CYT107 or placebo that they received. Green color represents CYT107 and grey color represents placebo.



**Supplemental Figure 3.** Effect of CYT107 on circulating levels of IL-10. Blood samples were collected from patients at 6-7 hours and 20-22 hours after the first and second doses of CYT107 or placebo. Results showed that circulating levels of IL-10 were not different in patients

receiving CYT107 versus placebo. Error bars represent SEMs. *Abbreviations*. FR, France; US, United States, UK, United Kingdom; D0, day zero, D3, day 3.



TNF-a US



TNF-a US D0-D3

TNF-a UK D0-D3



Blue=CYT107 Red=Placebo

**Supplemental Figure 4.** CYT107 did not increase circulating levels of TNF-α. Blood samples were collected from patients at 6-7 hours and 20-22 hours after the first and second doses of CYT107 or placebo. Results showed that circulating levels of TNF-α were not different in patients receiving CYT107 versus placebo. Error bars represent SEMs. *Abbreviations*. FR, France; US, United States, UK, United Kingdom; D0, day zero, D3, day 3.

## Supplemental Table 1:

	All Patier	nts		Anti-Vira	ls		No Anti-Virals		
	CYT10	Placeb	p-value	CYT10	Placeb	p-value	CYT10	Placeb	p-value
	7	ο		7	0		7	0	
	9.39	9.98	0.9719	10.10	9.50	0.2892	8.29	10.61	0.0955
	(	(		(	(		(	(	
Baseline	0	0		0	1		0	0	
WBC		-		-	•		-		
	5	7		8	0		6	9	
	6	5		2	9		0	7	
	)	)		)	)		)	)	
	8.42	8.65	0.8364	8.87	8.30	0.2892	7.71	9.13	0.2598
Deceline	(	(		(	(		(	(	
baseline	0	0		0	0		0	1	
Neut									
rophi	5	7		7	9		7	0	
ls	6	2		8	9		6	5	
	)	)		)	)		)	)	
	0.45	0.44	0 6600	0.48	0.49	0 7556	0.40	0.37	0 8660
Baseline	(	(	0.0000	(	(	0.1000	(	(	0.0000
Mon	(	(		(	(		(	(	
ocyte	0	0		0	0		0	0	
s								-	
	0	0		0	0		0	0	

		8	ļ	5			6		7		6	4	
		)		)			)		)		)	)	
	0.61		0.51		0.0160	0.61		0.50		0.1105	0.610	0.49	0.0663
		(		(			(	0.53			(	(	
Baseline		`		`			`		(		(	(	
		0	(	0			0				0	0	
Lym									0				
phoc		•		•			•						
price		0	(	0			0		•		0	0	
ytes				_			_		3		_	_	
		3		3			3		`		5	4	
		)		)			)		)		)	)	

Reported values are k/cumm<sup>3</sup> and represent the mean (Standard Error of the Mean) of the relevant study

populations. P-values are derived from the Mann-Whitney test. Abbreviations: WBC-White Blood Cells

Pts number

#### Average Duration of

antiviral

)

Treatment(days

Confid Interval

SD	±	MEDIAN	p-value

REMDESIVIR + CYT107	=					
31		5,32	3,44	± 6,88	5	
REMDESIVIR + PBO	=					
27		9,37	12,76	± 25,52	5	P=0,095

ACICLOVIR analogs + CYT107 =					
6	31,3	21,2	± 42,4	45	
ACICLOVIR analogs + PBO =					
11	23,64	18,2	± 36,4	26	P=0,44

Supplemental Table 3:

Treatment	Primary Cause Of Death	Related	Related
Subject Number		PI assessment	PhV final assessment
CYT107	Pulmonary fibrosis	Not related	No reasonable
			possibility
CYT107	Pulmonary air leakage	Unlikely to be related	No reasonable
			possibility
CYT107	Coronavirus pneumonia	Unlikely to be related	No reasonable
			possibility
CYT107	Multiple organ dysfunction syndrome	Not related	No reasonable
			possibility
CYT107	COVID-19 pneumonia	Unlikely to be related	No reasonable
			possibility
CYT107	Acute respiratory failure	Not related	No reasonable
			possibility
CYT107	Septic shock	Not related	No reasonable
			possibility
CYT107	Multiple organ dysfunction syndrome	Not related	No reasonable
			possibility
CYT107	Shock	Not related	No reasonable
			possibility
CYT107	Respiratory failure	Not related	No reasonable
			possibility
CYT107	Multiple organ dysfunction syndrome	Not related	No reasonable
			possibility

Treatment	Primary Cause Of Death	Related	Related
Subject Number		PI assessment	PhV final assessment
CYT107	Multiple organ dysfunction syndrome	Unlikely to be related	No reasonable
			possibility
CYT107	Multiple organ dysfunction syndrome	Not related	No reasonable
			possibility
CYT107	Multiple organ dysfunction syndrome	Not related	No reasonable
			possibility
CYT107	Acute Respiratory failure	Unlikely to be related	No reasonable
	Pneumonia	Unlikely to be related	possibility
			No reasonable
			possibility
CYT107	Cardiac arrest	Not related	No reasonable
			possibility
CYT107	Multiple organ dysfunction syndrome	Not related	No reasonable
			possibility
CYT107	Acute Respiratory failure	Not related	No reasonable
			possibility
Placebo	Pulmonary fibrosis	Not related	No reasonable
			possibility
Placebo	Respiratory arrest	Not related	No reasonable
			possibility
Placebo	Hemorrhagic stroke	Not related	No reasonable
			possibility

Treatment	Primary Cause Of Death	Related	Related
Subject Number		PI assessment	PhV final assessment
Placebo	Acute respiratory distress syndrome	Not related	No reasonable
	Pulmonary fibrosis		possibility
		Unlikely to be related	No reasonable
			possibility
Placebo	Respiratory Failure	Unlikely to be related	Not related
	Cardiac arrest	Potentially-Possibly	No reasonable
		related	possibility
Placebo	Multiple organ dysfunction syndrome	Unlikely to be related	No reasonable
			possibility
Placebo	Shock hemorrhagic	Not related	No reasonable
			possibility
Placebo	Нурохіа	Not related	No reasonable
			possibility
Placebo	Pneumonia	Not related	No reasonable
			possibility
Placebo	COVID-19 pneumonia	Not related	No reasonable
			possibility
Placebo	Multiple organ dysfunction syndrome	Not related	No reasonable
			possibility
Placebo	Multiple organ dysfunction syndrome	Not related	No reasonable
			possibility
Placebo	Multiple organ dysfunction syndrome	Not related	No reasonable
			possibility

Treatment	Primary Cause Of Death	Related	Related
Subject Number		PI assessment	PhV final assessment
Placebo	Cardiac arrest	Not related	No reasonable
			possibility
Placebo	Acute respiratory failure	Unlikely to be related	Not related
	Shock	Potentially-possibly	No reasonable
		related	possibility
	Acute kidney injury		Reasonable possibility
		Potentially-possibly	
		related	
Placebo	Respiratory failure	Potentially-possibly	No reasonable
		related	possibility