Supplemental Materials

Leishmania major drives host phagocyte death and cell-to-cell transfer depending on intracellular pathogen proliferation rate

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Figure S1: Gating strategies for determining original host cell material uptake in vivo and in vitro during *Leishmania major* infection. (A) Gating strategy for CD11b⁺Ly6G⁻ cells in cells isolated from the infected ear dermis of the mouse. (B) Gating strategy for single live infected (red) and non-infected (grey) human monocyte-derived macrophages (MDM).



Figure S2: Analysis of caspase-3 activity in vivo and in vitro. (A) Gating strategy for live CD45.2⁺ CFP-DEVD-YFP and CFP-DEVG-YFP cells in whole blood isolated from biosensor-transfected bone marrow chimeric mice. BM, bone marrow. **(B)** Gating strategy (upper left panel), histogram plots (lower left panel) and percent of cells (right panel) showing increased CFP/FRET ratio, indicating FRET loss, in cells expressing the CFP-DEVD-YFP (blue) and CFP-DEVG-YFP control (grey) biosensor as measured by flow cytometry analysis according to gating shown in (A-B). Each symbol represents one mouse. FRET, Foerster Resonance Energy Transfer. **(C)** Gating strategy for single live human MDM (left panel) and for DsRed expression (right panels) in infected (left) and non-infected (right) human MDM. MDM, monocyte-derived macrophages.



Figure S3: In vitro infection rates versus pathogen proliferation and impact of KBMA generation on fluorescence protein expression. (A) Number of parasites per infected macrophage determined in >100 cells infected with Lm^{SWITCH} 48h after photoconversion by widefield microscopy in vitro. Data pooled from 10 independently imaged fields of view. (B) Pathogen proliferation determined by widefield microscopy for Lm^{SWITCH} 48h after photoconversion in macrophages infected with 1, 2 or more than 2 parasites. Each symbol represents one cell, data pooled from 10 independently imaged fields of view. Horizontal bars denote the median. No significant differences were found according to Kruskal-Wallis test. (C) Gating strategy for single live cells (left panel) and histogram plots showing DsRed fluorescence (right panel) in BMDM infected with non-fluorescent Lm^{WT} (black), proliferationcompetent Lm^{DsRed} (green) and proliferation-incompetent KBMA Lm^{DsRed} (red) parasites. (D) DsRed mean fluorescence intensity (left) and percent of DsRed⁺ cells (right) in BMDM infected with nonfluorescent Lm^{WT} (black), proliferation-competent Lm^{DsRed} (green) and proliferation-incompetent KBMA Lm^{DsRed} (red) parasites. Each symbol represents one sample. Data represent three independent samples for each condition. Horizontal bars denote the mean. ****, p < 0.0001; ***, p < 0.001; ***, p < 0.01 according to one-way analysis of variance (ANOVA). KBMA, killed but metabolically active; BMDM, bone marrow-derived macrophages; WT, wild-type.



Figure S4: AnnexinV staining in BMDM infected with proliferation-competent and proliferation-incompetent KBMA *Lm*^{*DsRed*} **parasites. (A)** Gating strategy for live CD45⁺F4/80⁺ macrophages (left panels) and DsRed expression (right panels) in uninfected, proliferation-competent-infected and KBMA-infected BMDM. (B) Gating strategy for AnnexinV in uninfected, proliferation-competent-infected and KBMA-infected CD45⁺F4/80⁺ macrophages. (C) Percent of AnnexinV⁺ cells (normalized to uninfected) in uninfected (black), proliferation-competent-infected (green) and KBMA-infected (red) BMDM. Each symbol shows one individual sample. Horizontal bars denote the mean. Data were pooled from three independent experiments. KBMA, killed but metabolically active; BMDM, bone marrow-derived macrophages. No significant differences according to one-way ANOVA.



Figure S5: Host cell metabolism and *L. major* intracellular proliferation rate. (A) Gating strategy for live CD45⁺ murine BMDM. (B) Gating strategy for DsRed expression (upper panels) and histogram plots showing Glut1 expression (lower panel) in uninfected, Lm^{DsRed} proliferation-competent-infected and Lm^{DsRed} KBMA-infected BMDM. (C) Mean fluorescence intensity (normalized to uninfected) for Glut1 in uninfected (black), Lm^{DsRed} proliferation-competent-infected (green) and Lm^{DsRed} KBMA-

infected (red) BMDM. Each symbol shows one individual biological replicate. Horizontal bars denote the mean. Data pooled from three independent experiments. **(D)** Gating strategy for mKikume expression in $Lm^{mKikume}$ -infected (left panel) and Lm^{WT} -infected (right panel) CD45⁺CD11c⁺ cells isolated from the murine ear dermis. **(E)** Gating strategy for DsRed expression in Lm^{DsRed} -infected (left panel) and Lm^{WT} -infected (right panel) CD45⁺CD11c⁺ cells isolated from the murine ear dermis. **(F)** CD36 expression (black curve) and isotype control staining (grey histogram) in CD45⁺CD11c⁺ cells isolated from the murine ear dermis. **(G)** Gating strategy for DsRed expression (upper panels) and histogram plots showing C1/12 fatty acid uptake (lower panel) in uninfected (grey), Lm^{DsRed} proliferation competent-infected (green) and Lm^{DsRed} KBMA-infected (red) BMDM. **(H)** Mean fluorescence intensity (normalized to uninfected) for C1/12 fatty acid uptake in uninfected (black), Lm^{DsRed} proliferationcompetent-infected (green) and Lm^{DsRed} KBMA-infected (red) BMDM. Each symbol shows one individual sample. Horizontal bars denote the mean. Data are pooled from at least three independent experiments. KBMA, killed but metabolically active; BMDM, bone marrow-derived macrophages. No significant differences according to one-way ANOVA.