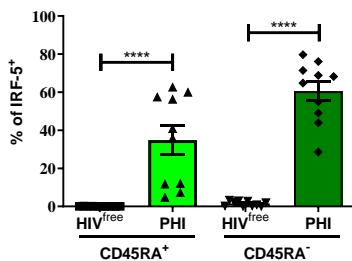
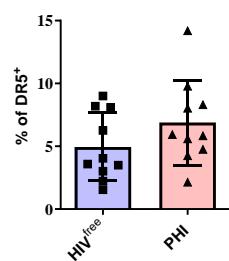


Supplemental Figure 1: Anti-IRF-5 antibody specificity and gating strategy. a) WT and *Irft^{-/-}* THP-1 cells were restimulated with either IMQ or PMA+IMQ and IRF-5 expression was assessed by flow cytometry. Graphs show a) representative FACS plots and the percentage of IRF-5 positive cells, and b) the frequency of IRF-5 nuclear localization. c) Representative FACS plots showing the gating strategy used to determine the frequency of cells positive for Annexin V, IRF-5⁺, IFN γ ⁺, and DR5⁺ in total CD4⁺ T cells or CD45RA⁺ and CD45RA⁻ CD4⁺ T cells in PBMCs isolated from PHI and HIV^{free} individuals.

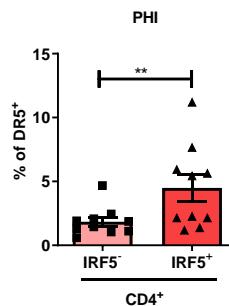
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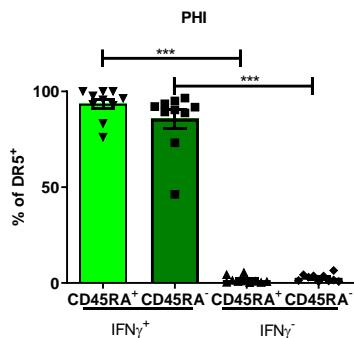
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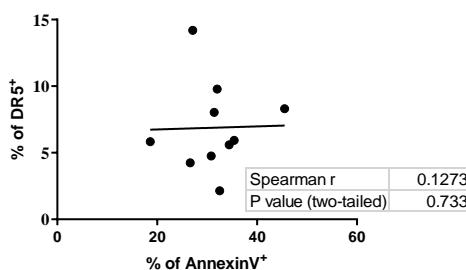
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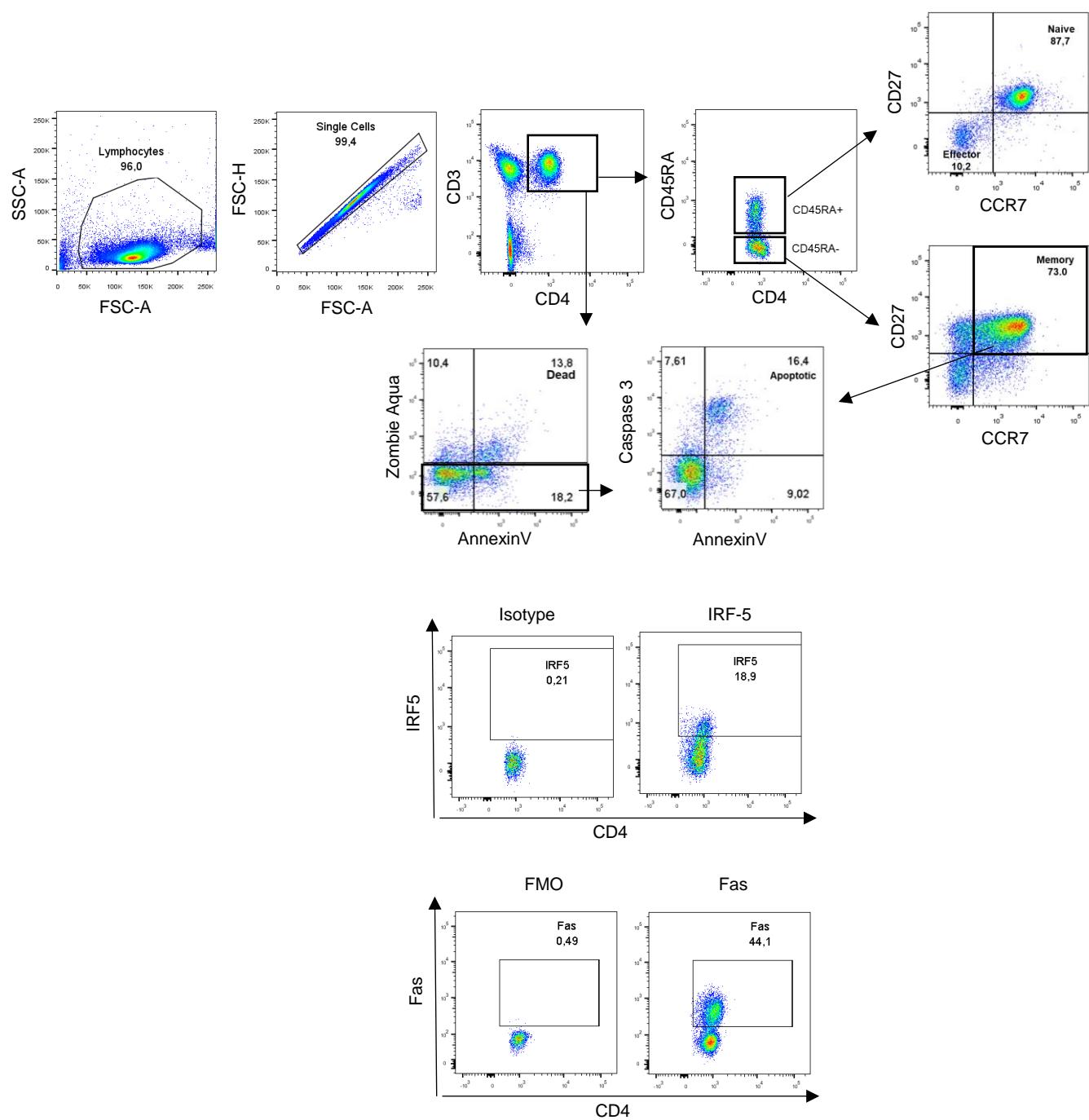
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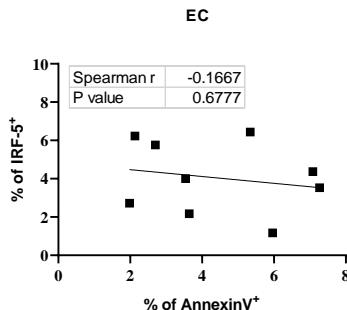
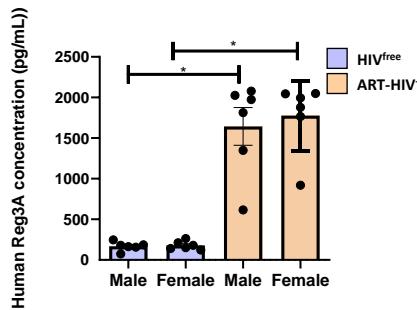
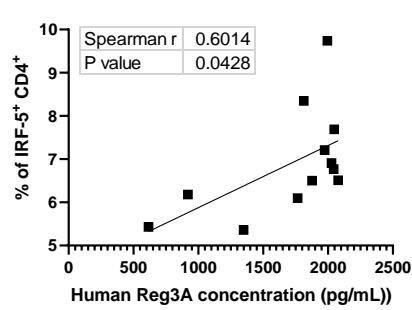
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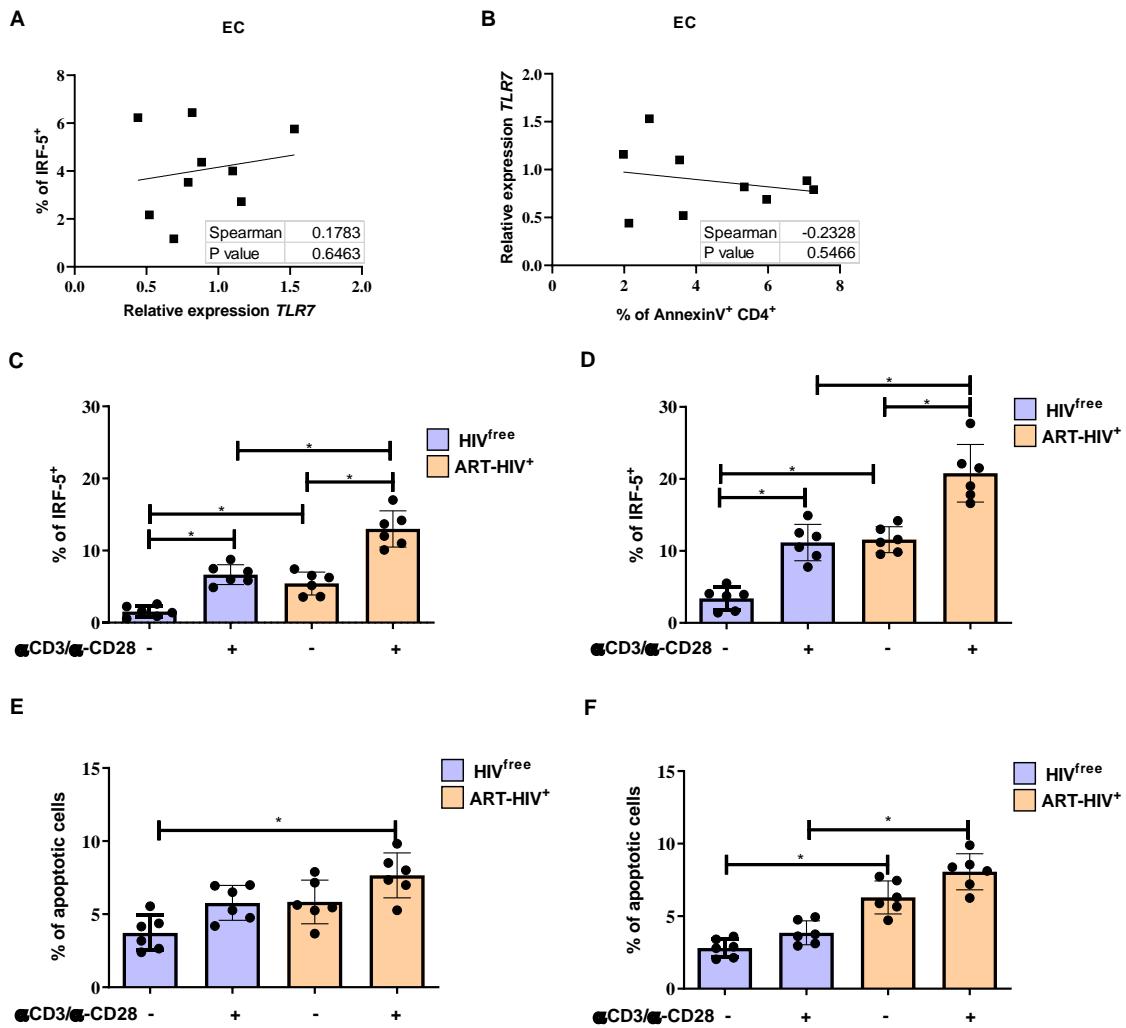
Supplemental Figure 2: DR5 expression in IRF-5⁺ cells. Graphs show a) the percentage of CD45RA⁺ and CD45RA⁻ CD4⁺ T cells expressing IRF-5 and b) the percentage of DR5⁺ CD4⁺ T cells in PHI and HIV^{free} individuals; c) the percentage of IRF5 expressing DR5⁺ CD4⁺ T cells, and d) the percentage of IRF5⁺ DR5⁺ CD4⁺ T cells expressing CD45RA⁺/ IFN γ ⁺ CD4⁺ T cells in PHI patients. Data are presented as the mean \pm SD. The Mann-Whitney U-test and the Kruskal-Wallis test followed by the Dunn's multiple comparison test were used to determine statistical significance, * p < 0.05, ** p < 0.01, *** p < 0.001, n=10. e) Graph represents correlation between Annexin V and DR5 expressions in memory CD4⁺ T cells from PHI patients. The Spearman r test was used to determine statistical significance, * p < 0.05, n=10.



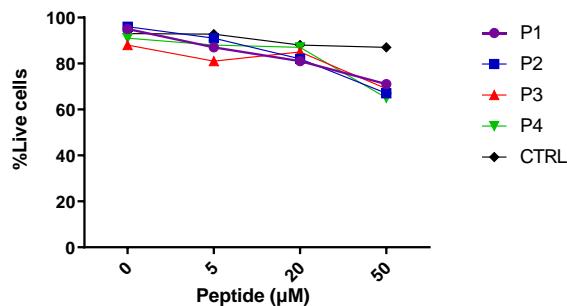
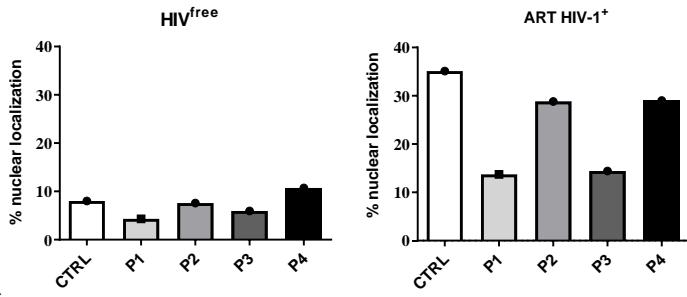
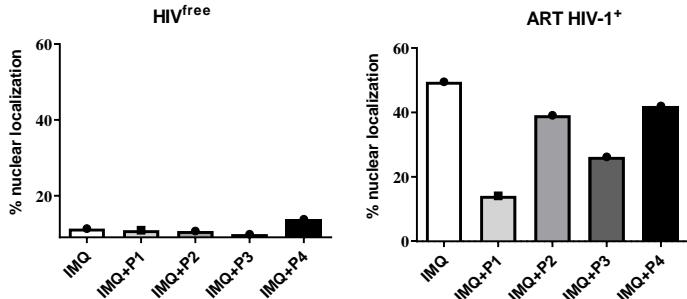
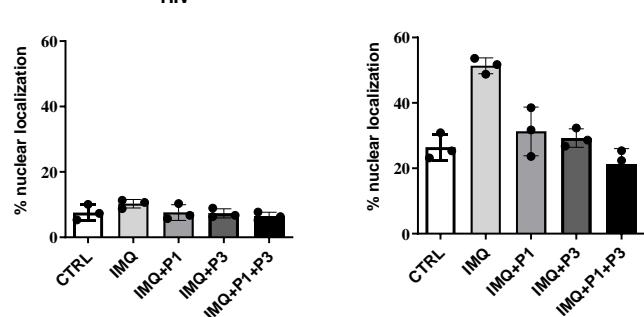
Supplemental Figure 3: Representative FACS plots showing the gating strategy used to determine the frequency of apoptotic, dead, IRF-5⁺, and Fas⁺ CD4⁺ T cells or to gate naïve, effector and memory CD4⁺ T cells from ART HIV-1⁺, EC, and HIV^{free} donors.

A**B****C**

Supplemental Figure 4: IRF-5 expression does not correlate with Annexin V expression in EC but correlates with REG3a concentration in the serum of ART HIV-1+ donors. a) Graph represents correlation between Annexin V and IRF5 expressions in memory CD4⁺ T cells from EC. The Spearman r test was used to determine statistical significance, * p <0.05, n=10. b-c) The concentration of REG3a in the donors' serum was assessed by ELISA. Graphs show b) REG3a concentrations in the serum of male and female ART HIV-1+ participants, and c) the correlation between IRF-5 expression and REG3a serum concentrations. Data are presented as the mean \pm SD. The Friedman's test and the Dunn's multiple comparison test were used to determine statistical significance, * p <0.05, n=6. For c), the Spearman r test was used to determine statistical significance, * p <0.05, n=12 (6 female and 6 male donors).

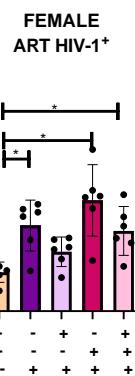
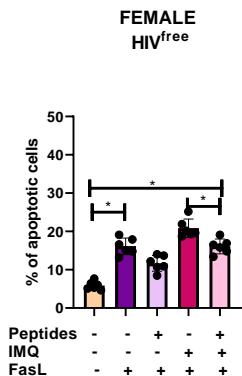


Supplemental Figure 5: Graphs represent correlations between a) *TLR7* and *IRF-5* expressions and b) *TLR7* and Annexin V expressions in memory CD4⁺ T cells from EC. Spearman r test was used to determine statistical differences * p <0.05, n=9. c-f) Purified CD4⁺ T cells from HIV-1⁺ and HIV^{free} donors were stimulated for 24h with αCD3/αCD28. Graphs show c) the percentage of *IRF-5⁺* total CD4⁺ T cells and the percentage of d) *IRF-5⁺*, e) apoptotic cells, and f) dead cells in memory CD4⁺ T cells. Data are presented as the mean ± SD. The Friedman's test and the Dunn's multiple comparison test were used to determine statistical significance, * p <0.05, n=6.

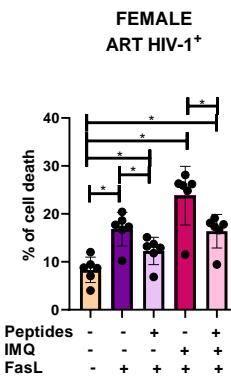
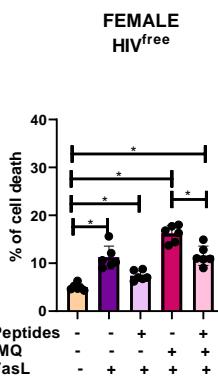
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Supplemental Figure 6: IRF-5 inhibitory peptides limit Fas-mediated apoptosis in memory CD4⁺ T cells. Four different IRF-5 blocking peptides and the control peptide were tested for their cytotoxicity and their capacity to block IRF-5 in CD4⁺ T cells. Graphs show a) the percentage of live PBMCs from HIV^{free} donors upon incubation with IRF-5 inhibitory peptides at various concentrations, and b) IRF-5 nuclear localization in unstimulated and c,d) IRF-5 nuclear localization in IMQ-stimulated CD4⁺ T cells from ART HIV-1⁺ and HIV^{free} donors in the presence or absence of 10 μ M of the indicated peptides or peptide mix. Data are presented as the mean \pm SD, n=3.

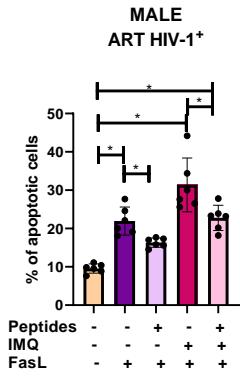
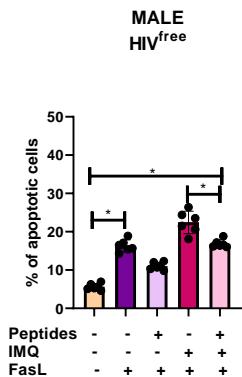
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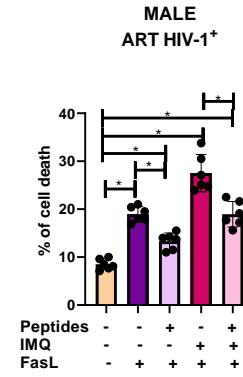
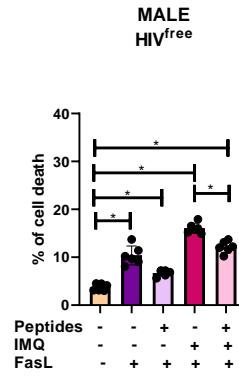
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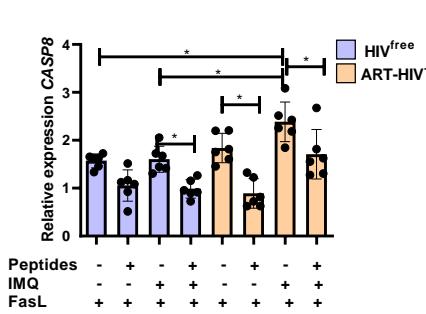
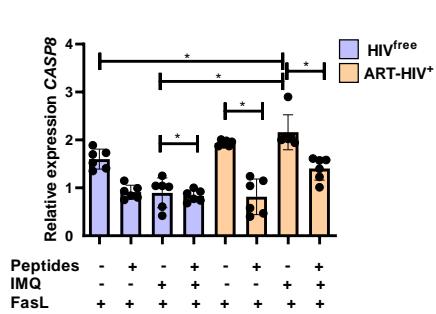
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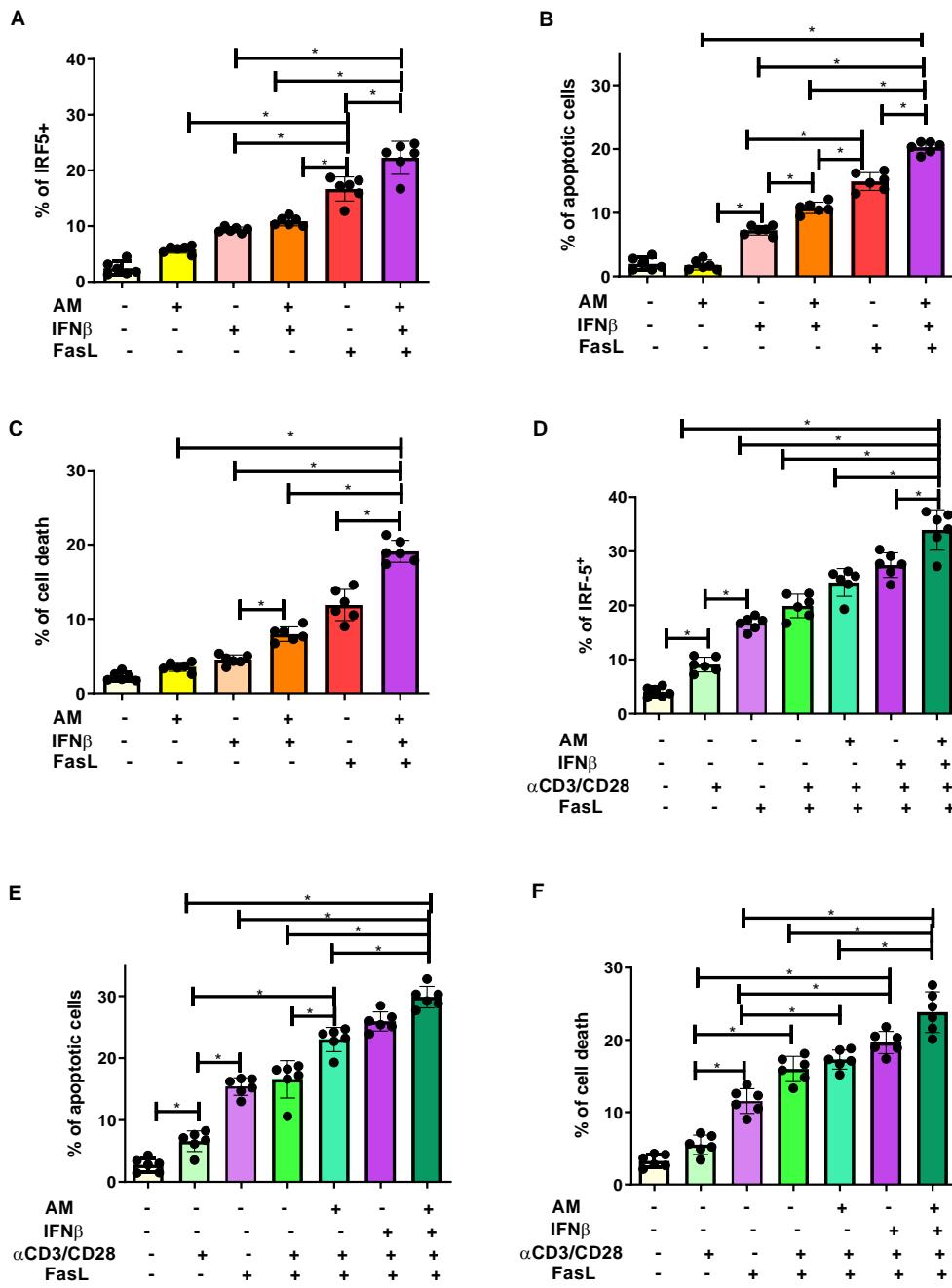
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Supplemental Figure 7: CD4 T cells from male and female participants display similar responses. a) Purified CD4⁺ T cells from female and male ART HIV-1⁺ and HIV^{free} donors were treated with IMQ or medium alone; 12h later rFasL or medium were added to the culture and the cells were incubated for further 18h for a total of 30h incubation at 37°C. Graphs show the percentage of apoptotic memory CD4⁺ T cells from a) female and c) male donors , the percentage of dead memory CD4⁺ T cells from b) female and d) male participants, and e) CASP8 mRNA levels in memory CD4⁺ T cells from male (left graph) and female (right graph) donors after 30h stimulation. Data are presented as the mean \pm SD. The Friedman's test and the Dunn's multiple comparison test were used to determine statistical significance, * p <0.05, n=6.



Supplemental Figure 8: Stimulation of memory CD4⁺ T cells in the presence of DAMPs and IFN β promotes the upregulation of TLR7 and predisposes cells to Fas-mediated apoptosis. Purified CD4⁺ T cells from HIV^{free} individuals were treated *in vitro* for 24 h with 10 ng/ml IFN β in the presence or absence of 10% v/v supernatant containing apoptotic material (AM, supernatant of staurosporine-treated cells), before stimulation with α CD3/ α CD28 for 24h. Graphs show a) the percentage of IRF-5⁺, b) the percentage of apoptotic and c) the percentage of death memory CD4⁺ T cells. CD4⁺ T cells from HIV^{free} individuals were treated as described above and additionally incubated with rFasL for further 18h after α CD3/ α CD28 stimulation. Graphs show d) the percentage of IRF-5⁺, e) the percentage of apoptotic and f) the percentage of death memory CD4⁺ T cells. Data are presented as the mean \pm SD. The Friedman's test and the Dunn's multiple comparison test were used to determine statistical significance, * p <0.05, n=6.