

Supplementary materials

Radiotherapy exposure directly damages the uterus and causes pregnancy loss

Griffiths et al

SUPPLEMENTARY FIGURE 1

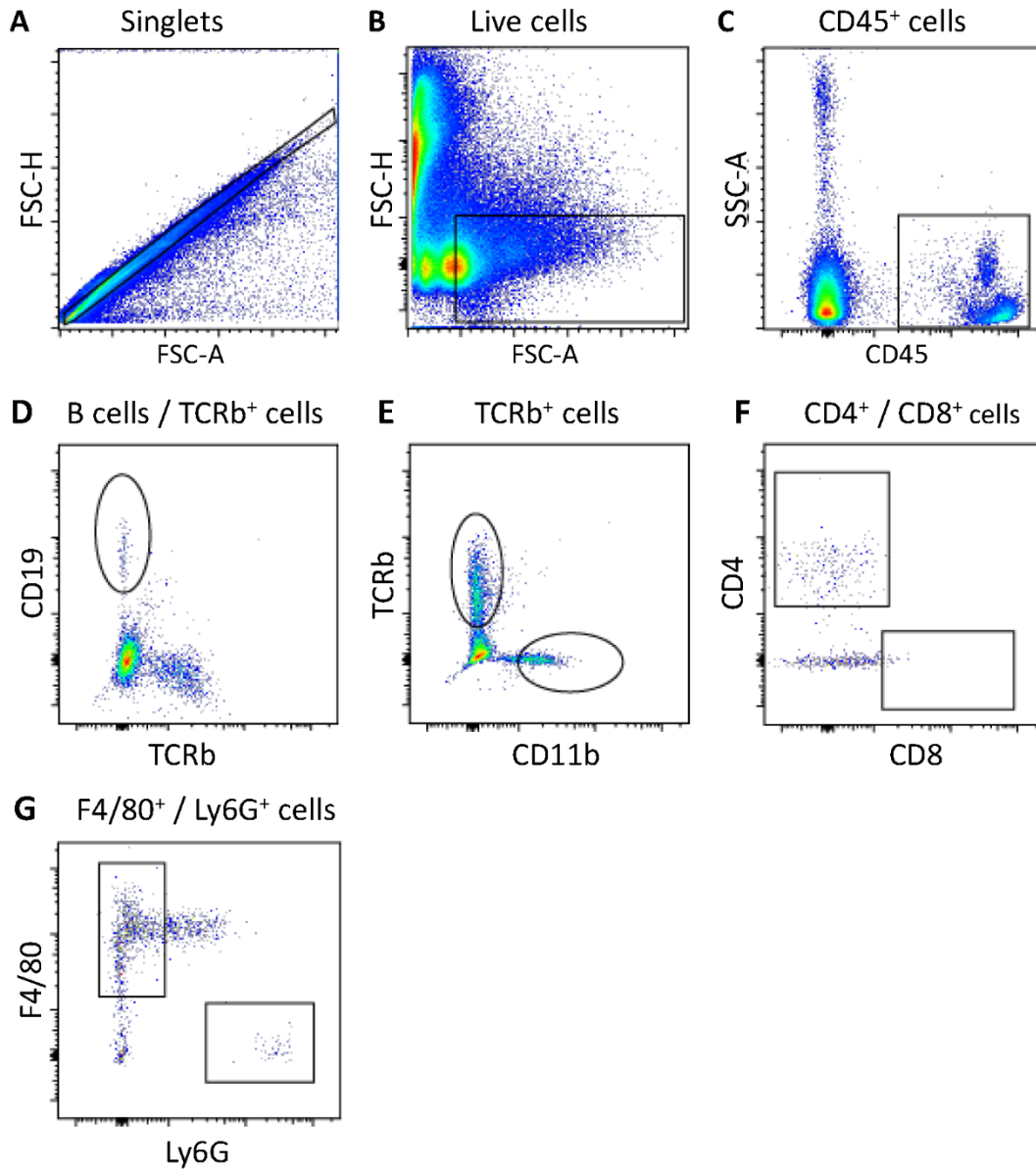
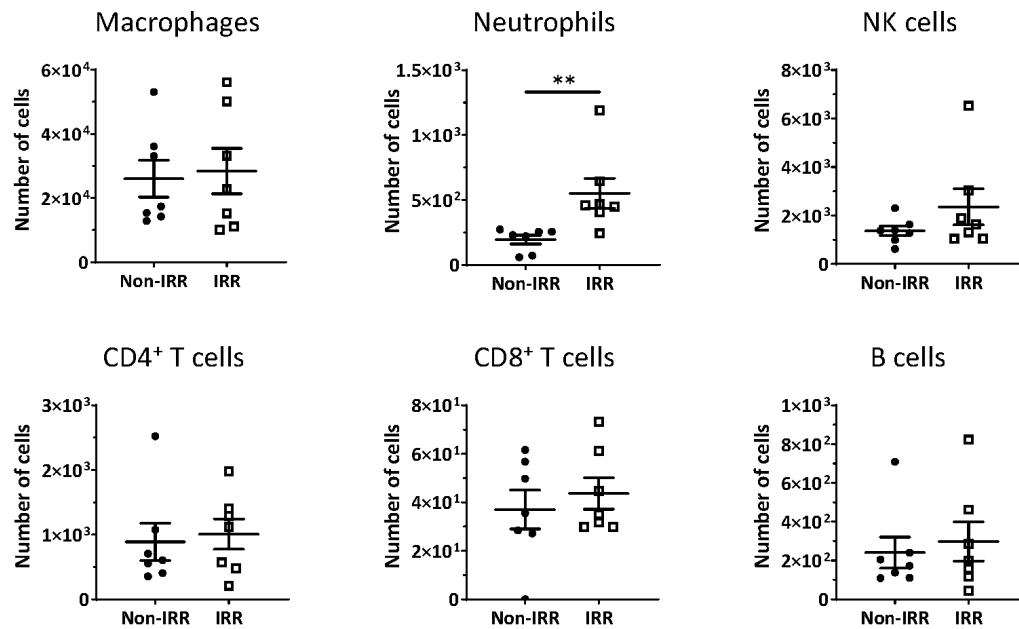


Fig. S1. Flow cytometry gating strategy. Cells were identified by gating for singlets (A), and live cells (B). From these, haematopoietic cells were identified by gating for CD45⁺ cells (C). These cells were separated into CD19⁺ and TCRb⁺ cells to identify B and T cells respectively (D), as well as CD11b⁺ cells by gating against TCRb (E). TCRb⁺ cells were further defined as CD4⁺ and CD8⁺ T cells (F) whereas CD11b⁺ cells were further defined as F4/80⁺ and Ly6G⁺ cells (G).

SUPPLEMENTARY FIGURE 2

A

Uterine immune cells



B

Splenic immune cells

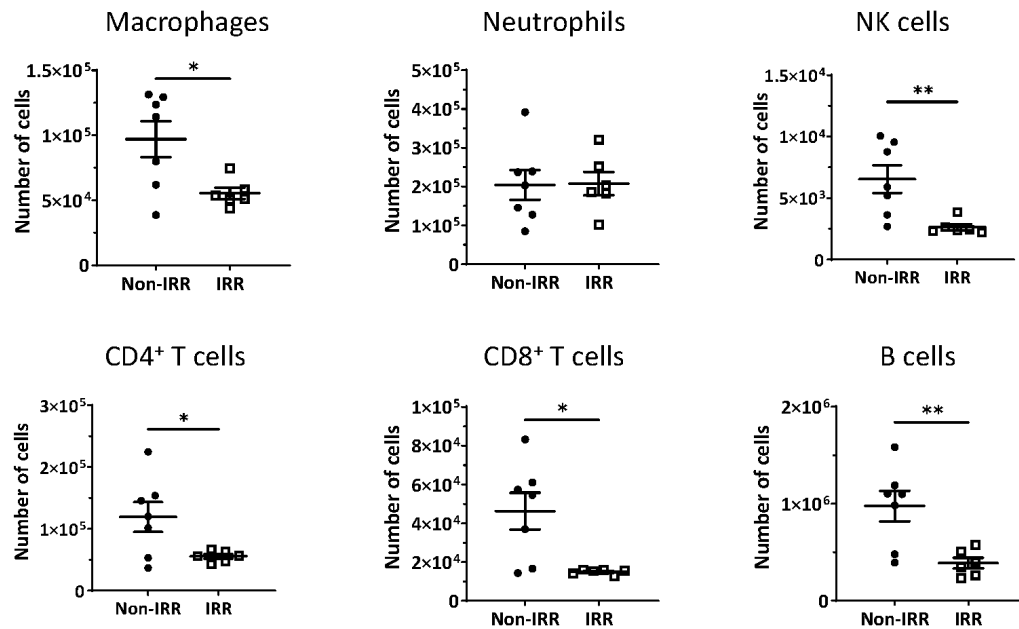


Fig. S2. Uterine cell populations are restored by 4-weeks post-irradiation. Uterine (A) and peripheral (B) immune cell populations were analysed by flow cytometry 4 weeks post-irradiation. Macrophages (i), neutrophils (ii), natural killer (NK) cells (iii), CD4⁺ T cells (iv), CD8⁺ T cells (v) and B cells (vi) were quantified. Data are mean ± SEM; unpaired t-test (2 groups; parametric distribution) or Welch's t-test (2 groups; parametric distribution; unequal variances); * = p<0.05, ** = p<0.01; n=6-7/group.

SUPPLEMENTARY FIGURE 3

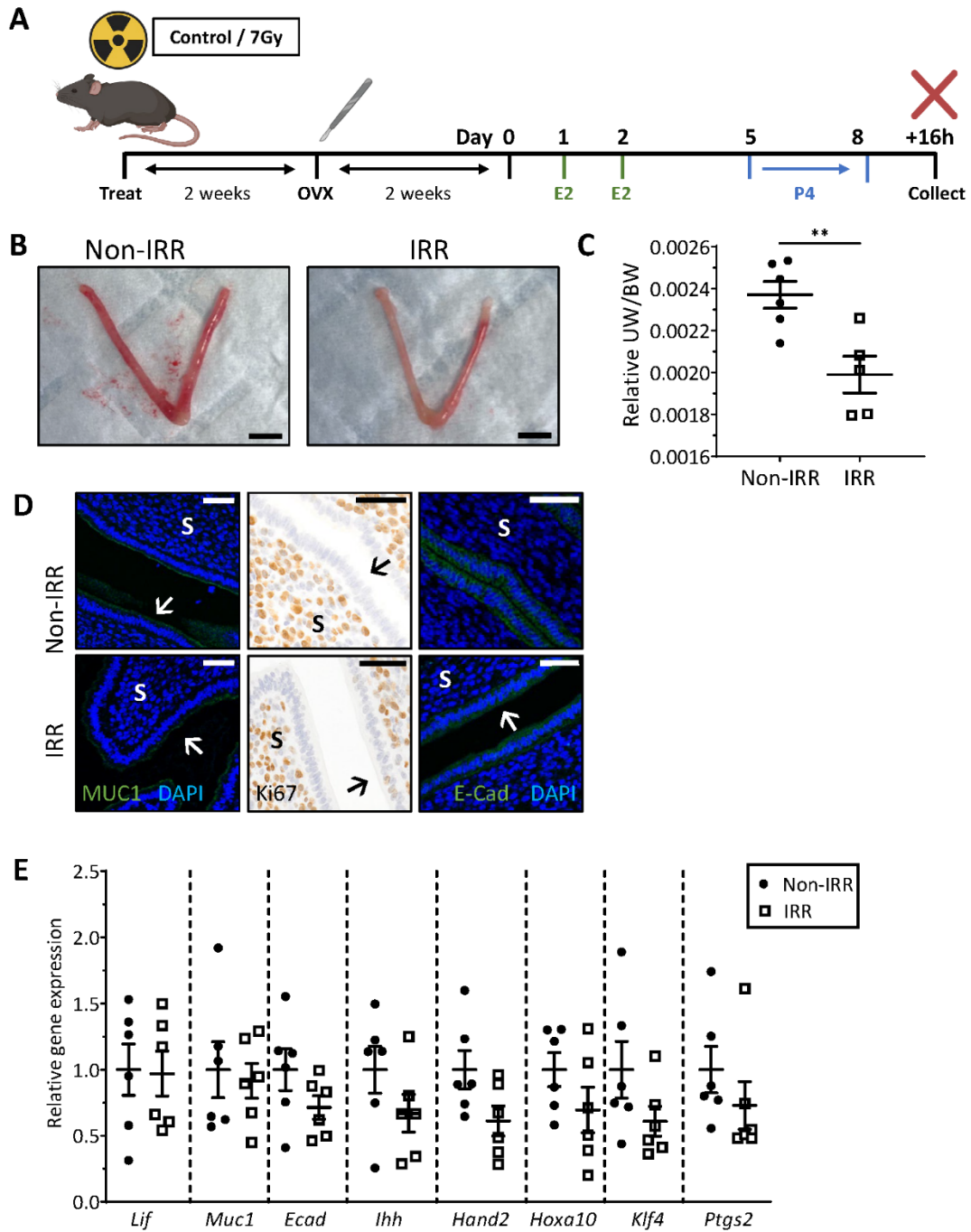


Fig. S3. Endometrial receptivity occurs in female mice following irradiation. (A) Ovariectomised (OVX) adolescent female IRR or Non-IRR wild-type mice were hormone-primed with E2 and P4 to artificially induce endometrial receptivity, then collected 16 h later. (B) Representative images of receptive uteri and (C) relative uteri weight normalised to bod weight. (D) Representative images of immunostaining for MUC1, Ki67 and ECAD (whose expression is lost to attain successful endometrial receptivity) are shown. (E) Expression of genes critical to endometrial receptivity were analysed by RT-qPCR. Data are mean $\pm$ SEM; unpaired t-test (2 groups; parametric distribution) or Mann-Whitney test (2 groups; non-parametric distribution); n=5-7/group.

SUPPLEMENTARY FIGURE 4

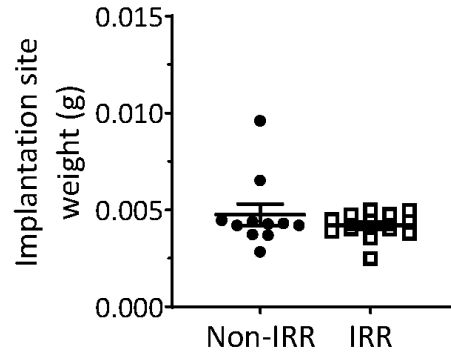


Fig. S4. Implantation site weight unchanged following irradiation. Individual implantation site weights 3-days after embryo transfer are unchanged between non-irradiated and irradiated animals. Data are mean $\pm$ SEM; unpaired t-test (2 groups; parametric distribution) or Mann-Whitney test (2 groups; non-parametric distribution); n=11-13/group.

SUPPLEMENTARY FIGURE 5

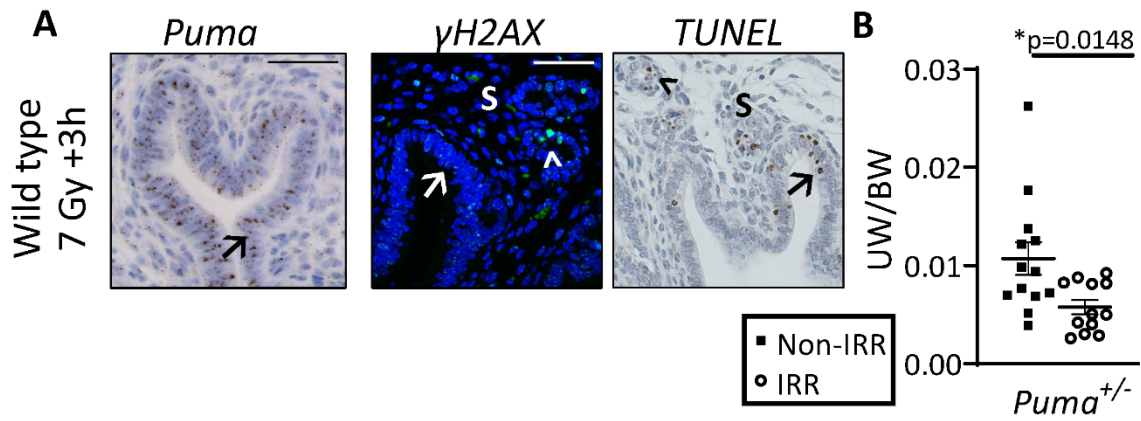


Fig. S5. (A) Activation of *Puma*, DNA damage (γ H2AX), and apoptosis (TUNEL) in wild-type mouse uterus 3 hours following irradiation. **(B)** *Puma*^{+/-} mice with one functional copy of *Puma* gene have significantly reduced uterine to body weight ratio following artificial induction of decidualisation. Scale bars are 25 μ m (*Puma*, γ H2AX), or 50 μ m (TUNEL). $\rightarrow$ luminal epithelium, S stroma, > glandular epithelium Data are mean $\pm$ SEM; unpaired t-test (2 groups; parametric distribution) or Mann-Whitney test (2 groups; non-parametric distribution); n=3-13/group, *p<0.05.

Table S1. Mesenteric artery responses in non-pregnant control and 7Gy irradiated mice. Data presented as mean $\pm$ SEM.

<i>Mesenteric artery</i>	n	Control	n	7Gy
<i>ACh</i>				
pEC50	7	7.93 $\pm$ 0.25	8	7.47 $\pm$ 0.45
AUC	8	271.4 $\pm$ 32.29	8	213.9 $\pm$ 37.63
<i>SNP</i>				
pEC50	7	7.98 $\pm$ 0.17	8	8.05 $\pm$ 0.19
AUC	8	308.60 $\pm$ 29.40	8	285.00 $\pm$ 21.22
<i>AngII</i>				
pEC50				
AUC	8	33.49 $\pm$ 6.12	7	38.26 $\pm$ 11.68
<i>PE</i>				
pEC50	10	6.11 $\pm$ 0.12	7	6.13 $\pm$ 0.18
AUC	10	192.00 $\pm$ 13.77	7	199.2 $\pm$ 37.36
<i>ET1</i>				
pEC50	9	9.27 $\pm$ 0.36	9	9.34 $\pm$ 0.26
AUC	9	289.00 $\pm$ 48.43	9	253.20 $\pm$ 36.88
<i>U46619</i>				
pEC50	9	8.77 $\pm$ 0.23	7	8.67 $\pm$ 0.33
AUC	10	396.9 $\pm$ 19.62	7	412.20 $\pm$ 67.07

Table S2. Immunofluorescence and immunohistochemistry antibodies.

	Catalogue	Species	Antigen retrieval	Conc	Secondary	Conc
αSMA	Dako M0815	Mouse	Citrate	1:200	Goat anti-mouse (488)	1:500
CD31	R&D Systems AF3628	Goat	Citrate	1:100	Horse anti-goat biotinylated	1:500
Pan-Cytokeratin	ab9377	Rabbit	Citrate	1:400	Goat anti-rabbit (568)	1:500
Cleaved caspase-3	CST#9661	Rabbit	Citrate	1:100	Goat anti-rabbit (488)	1:500
E-cadherin	AF748	Goat	Citrate	1:1000	Donkey anti-goat (488)	1:500
Desmin	CST#5332	Rabbit	Citrate	1:100	Goat anti-rabbit (488)	1:500
Mucin1	Ab15481	Rabbit	Citrate	1:100	Goat anti-rabbit (488)	1:500
Ki67	Ab16667	Rabbit	EDTA	1:500	Goat anti-rabbit biotinylated	1:500
γH2AX	CST#9718	Rabbit	Citrate	1:200	Goat anti-rabbit (488)	1:500

Table S3. Flow cytometry antibodies.

	Fluorophore	Laser	Concentration	Company
CD19	BV650	405	1:400	BD Biosciences, 563235
CD4	BUV395	355	1:400	BD Biosciences, 563790
CD25	BV786	405	1:400	BD Biosciences, 564023
CD11b	FITC	488	1:400	BD Biosciences, 557396
TCRb	BV510	405	1:400	BD Biosciences, 563221
F4/80	PE	561	1:200	BD Biosciences, 565410
NK1.1	PE-Cy7	561	1:800	BD Biosciences, 552878
CD8a	PerCP-Cy5.5	488	1:800	BD Biosciences, 551162
CD45	APC Cy7	628	1:400	BD Biosciences, 557659
Ly6G	BV421	405	1:400	BD Biosciences, 562737
Live-dead	FVS700	628	1:2000	BD Biosciences, 564997

Table S4. Mouse primer sequences.

Gene	Forward	Reverse
<i>18s</i>	GTAACCCGTTGAACCCCAT	CCATCCAATCGGTAGTAGCG
<i>Bmp2</i>	GGGACCCGCTGTCTTCTAGT	TCAACTCAAATTCGCTGAGGAC
<i>Ecad</i>	CGTCCATGTGTGTGACTGTG	GCTCTTTGACCACCGTTCTC
<i>Esr1</i>	GCTCCTAACTTGCTCCTGGAC	CAGCAACATGTCAAAGATCTCC
<i>Hand2</i>	TCGGTTATCTAGTGCTGTC	ATACTTACAATGTTTACACCTT
<i>Hoxa10</i>	GCCCCTTCAGAAAACAGTAAA	AGGTGGACGCTACGGCTGAT
<i>Ifng</i>	CGGCTGACCTAGAGAAGACAC	CCAAGATGCAGTGTGTAGCG
<i>lhh</i>	CTCTTGCCTACAAGCAGTTCA	CCGTGTTCTCCTCGTCCTT
<i>Il10</i>	GCTCTTACTGACTGGCATGAG	CGCAGCTCTAGGAGCATGTG
<i>Il15</i>	CATCCATCTCGTGCTACTTGTGTT	CATCTATCCAGTTGGCCTCTGTTT
<i>Il17</i>	AAGGCAGCAGCGATCATCC	GGAACGGTTGAGGTAGTCTGAG
<i>Il1a</i>	GTATGCCTACTCGTCGGGAG	GGCAACTCCTTCAGCAACAC
<i>Il1b</i>	GAAGAAGAGCCCATCCTCTG	GGAGCCTGTAGTGCAGTTGT
<i>Il23</i>	CCAGCAGCTCTCTCGGAATC	CAGACCTTGGCGGATCCTTT
<i>Klf4</i>	GTGCCCCGACTAACCGTTG	GTCGTTGAACTCCTCGGTCT
<i>Lif</i>	AAAAGCTATGTGCGCCTAACA	GTATGCGACCATCCGATACAG
<i>Muc1</i>	GGCATTTCGGGCTCCTTTCTT	TGGAGTGGTAGTCGATGCTAAG
<i>Pgr</i>	CTCCGGGACCGAACAGAGT	ACAACAACCCTTTGGTAGCAG
<i>Ptgs2</i>	AGCCAGGCAGCAAATCCTT	CAGTCCGGGTACAGTCACAC
<i>Tnfa</i>	CCTGGCCTCTCTACCTTGTTG	AGCCTGGTCACCAAATCAGC

Table S5. Human primer sequences.

Gene	Forward	Reverse
<i>PRL</i>	AAAGGATCGCCATGGAAAG	GCACAGGAGCAGGTTTGAC
<i>IGFBP1</i>	AATGGATTTTATCACAGCAGACAG	AATGGATTTTATCACAGCAGACAG
<i>b-ACTIN</i>	TCACCCACACTGTGCCCATCTACGA	CAGCGGAACCGCTCATTGCCAATGG
<i>GAPDH</i>	GAAGGTGAAGGTCGGAGTCAAC	CAGAGTTAAAAGCAGCCCTGGT