| | Mitochondrial Targeting Sequence FAD-binding domain | | |
|------------------------|---|------------|--|
| H.sapiens | MQSWSRVYCSLAKRGHFNRISHGLQGLSAVPLRTYADQPIDADVTVIGSGPGGYVAAIKA | 60 | |
| C.elegans | MSLSRTTQLPFAKRQFFQVLARNY-SNTQDADLVVIGGGPGGYVAAIKA *. . *. . *. . *. . *. . *. . *. . *. . . | 48 | |
| H.sapiens C.elegans | AQLGFKTVCIEKNETLGGTCLNVGCIPSKALLNNSHYYHMAHGTDFASRGIEMSEVRLNL AQLGMKTVCVEKNATLGGTCLNVGCIPSKALLNNSHYLHMAQ-HDFAARGIDCT-ASLNL ****:****:*** ************************ | 120 106 | |
| H.sapiens C.elegans | DKMMEQKSTAVKALTGGIAHLFKQNKVVHVNGYGKITGKNQVTATKADGGTQVIDTKNIL PKMMEAKSNSVKQLTGGIKQLFKANKVGHVEGFATIVGPNTVQAKKNDGSVETINARNIL **** **.:** ***** :*** *** **:*:*.* * * *.*** NAD-binding domain | 180 166 | |
| H.sapiens C.elegans | IATGSEVTPFPGITIDEDTIVSSTGALSLKKVPEKMVVIGAGVIGVELGSVWQRLGADVT IASGSEVTPFPGITIDEKQIVSSTGALSLGQVPKKMVVIGAGVIGLELGSVWQRLGAEVT **:********************************** | 240 226 | |
| H.sapiens C.elegans | AVEFLGHVGGVGIDMEISKNFQRILQKQGFKFKLNTKVTGATKKSDGKIDVSIEAASGGK AVEFLGHVGGMGIDGEVSKNFQRSLTKQGFKFLLNTKVMGASQN-GSTITVEVEGAKDGK *********************************** | 300 285 | |
| H.sapiens C.elegans | AEVITCDVLLVCIGRRPFTKNLGLEELGIELDPRGRIPVNTRFQTKIPNIYAIGDVVAGP | | |
| H.sapiens C.elegans | MLAHKAEDEGIICVEGMAGGAVHIDYNCVPSVIYTHPEVAWVGKSEEQLKEEGIEYKVGK 4 MLAHKAEDEGILCVEGIAGGPVHIDYNCVPSVVYTHPEVAWVGKAEEQLKQEGVAYKIGK 4 ************************************ | | |
| H.sapiens C.elegans | FPFAANSRAKTNADTDGMVKILGQKSTDRVLGAHILGPGAGEMVNEAALALEYGASCEDI FPFVANSRAKTNNDQEGFVKVLADKQTDRMLGVHIIGPNAGEMIAEATLAMEYGASAEDV ***.******* * :*:*:*:*.:*.**:**.** | 480 465 | |
| H.sapiens C.elegans | ARVCHAHPTLSEAEAFREANLAASFGKSINF- 511 ARVCHPHPTLSEAFREANLAAYCGKAINNV 495 ***** ****** ******** **:** | | |

Supplemental Figure 1. Sequence homology of human and *C. elegans* **DLD proteins**. Different interfaces and domains are specified. The mitochondrial targeting sequence (MTS) would be truncated following transport into the mitochondrial matrix and is absent in the active ketoacid dehydrogenase complexes.



Supplemental Figure 2. DLD-1 knockdown by feeding RNAi in *C. elegans* from egg

hatching reduces adult growth. At L4 stage, worms were not significantly different in size. Their reduced size differential becomes more evident as adults and reaches statistical significance relative to wild-type (N2) worms at adult day 3 for the full dose *dld-1(RNAi)* worms. Note that adult worm growth is not as evident when comparing images because of the indicated sequential reductions in size of the images. The white scale bar is a constant 30 pixels in all panels.



Supplemental Figure 3. Chemotaxis assay performance details. Chemoattraction was assayed by placing worms on nematode growth media agar plates at the origin, highlighted with a green circle, and isoamyl alcohol, a diffusible chemoattractant, 5 cm away in a well, identified by the red circle. Worms, identified as blue-black spots, were allowed to freely migrate for 1 h, after which time each worm's distance from the chemoattractant well was measured on different days of adulthood. At day 7, while N2 worms still migrated with some efficacy towards the chemoattractant the *dld-1(RNAi)* knockdown worms moved in largely random directions.





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Supplemental Figure 4. Determination of mitochondrial membrane potential with relative TMRE uptake. (A) Wild-type (N2) worms expressing a COX4::GFP after exposure to 1 μ M TMRE and subsequent washout (B) Full-dose *dld-1(RNAi)* worms expressing COX4::GFP following exposure to TMRE and subsequent washout.







Supplemental Figure 6. UPR^{mt} **dose response curves for DCA and thiamine.** Full dose *dld-1(RNAi)* worms were treated with 0.1 to 25 mM concentrations of either of the two drugs from hatching, that demonstrated efficacy at reducing the UPR ^{mt}, a prominent phenotype of the worms, in four biological replicates. Each data point represents the mean of ~300 worms normalized to the value for the untreated worms ± the SEM. For both drugs, the trend became statistically significant with the 25 mM dose (****P* < 0.001).

Supplemental Table 1

| Water Soluble Drugs | Size | hsp6p::GFP |
|--------------------------|-----------|------------|
| Nicotinic Acid (1 mM) | No Effect | No Effect |
| Nicotinamide (200 µM) | No Effect | No Effect |
| Riboflavin (10 μM) | Increase | Increase |
| Thiamine (25 mM) | Increase | Decrease |
| L-Carnitine (100 μM) | No Effect | No Effect |
| Folinic Acid (10 µM) | No Effect | Increase |
| Glucose (10 mM) | No Effect | Increase |
| DCA (25 mM) | No Effect | Decrease |
| Cysteamine (100 µm) | No Effect | No Effect |
| Bitartrate (100 μM) | No effect | Decrease |
| NAC (2.5 mM) | No Effect | No Effect |
| AICAR (500 μM) | No Effect | Decrease |
| Hydralazine (200 μM) | Increase | No Effect |
| Lithium Chloride (10 mM) | No Effect | Decrease |
| Cycloheximide (2 μM) | No Effect | Increase |
| Arginine (10 mM) | No Effect | Increase |
| Taurine (800 μM) | Increase | Decrease |
| Taurine (8 mM) | No Effect | No Effect |
| DMSO Soluble Drugs | Size | hsp6p::GFP |
| Epicatechin (10 nM) | No Effect | No Effect |
| Resveratrol (50 µM) | Increase | Increase |
| Ethanol Soluble Drugs | Size | hsp6p::GFP |
| Rapamycin (2.5 nM) | No Effect | No Effect |
| Probucol (5 μM) | No Effect | No Effect |
| Lipoic Acid (10 µM) | No Effect | Increase |
| Vitamin E (250 µM) | No Effect | Increase |

Supplemental Table 1. Drugs tested at the noted concentration for their effects on the growth of *dld-1(RNAi) C. elegans* for their effects on growth at adult day 3 and on the expression of GFP under the control of the HSP6 promoter at adult day 2.

Full Unedited Complete Gel for Figure 2A. Four separate images of the same gel. (A) The gel was probed with anti-DLD (red fluorescence) with fluorescent green molecular weight markers in lane 4. Lanes 1-3 were loaded with the digest of worms fed: (lane 1) full dld-1(RNAi), (lane 2) 1:20 dld-1(RNAi), (lane 3) control plasmid L4440. (B) Because the electrophoretic mobility of the tubulin and DLD-1 are nearly identical, the gel in panel A was stripped and probed with antitubulin (red fluorescence) to serve as a C. elegans protein loading standard. (C) The image in (A) was flipped and the gel converted to gray scale for integration using Image J (D) The image in (B) was flipped and the gel converted to gray scale for integration using Image J.





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