

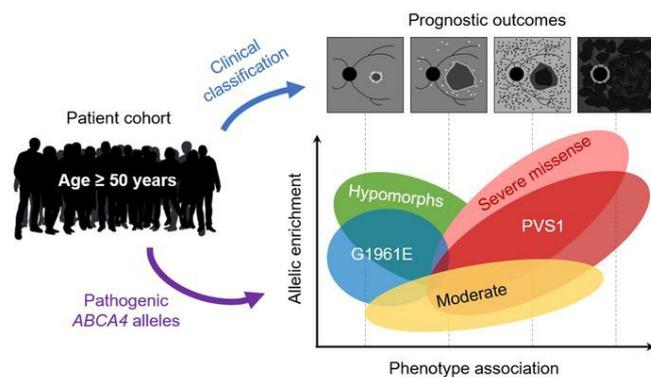
A genotype-phenotype correlation matrix for Stargardt/ABCA4 disease based on long-term prognostic outcomes

Winston Lee, ... , Stephen H. Tsang, Rando Allikmets

JCI Insight. 2021. <https://doi.org/10.1172/jci.insight.156154>.

Clinical Medicine In-Press Preview Genetics Ophthalmology

Graphical abstract



Genotype-Phenotype Correlation Matrix

	Prognosis 1	Prognosis 2	Prognosis 3	Prognosis 4
p.(Gly1961Glu)	67%	33%	0%	0%
Frequent hypomorph	44%	39%	17%	0%
Rare hypomorph	13%	69%	19%	0%
Moderate	14%	16%	30%	41%
Severe/PVS1	0%	0%	6%	94%

Precision Medicine

- Prognosis prediction
- Genetic counseling
- Clinical trial design

Find the latest version:

<https://jci.me/156154/pdf>



1 **A genotype-phenotype correlation matrix for Stargardt/ABCA4 disease based on**
2 **long-term prognostic outcomes**

3 *Winston Lee,^{1,2} Jana Zernant,² Pei-Yin Su,² Takayuki Nagasaki,² Stephen H. Tsang,^{2,3}*
4 *Rando Allikmets^{2,3*}*

5 **Affiliations:**

6 ¹Department of Genetics & Development, Columbia University Irving Medical Center, New
7 York, NY, USA

8 ²Department of Ophthalmology, Columbia University Irving Medical Center, New York,
9 NY, USA

10 ³Department of Pathology & Cell Biology, Columbia University Irving Medical Center, New
11 York, NY, USA

12
13 **Keywords:** ABCA4, Stargardt disease, retinal degeneration, genotype, phenotype,
14 correlation, prognosis, hypomorph

15
16 ***Correspondence:**

17 Rando Allikmets, Ph.D.
18 Department of Ophthalmology,
19 Columbia University Irving Medical Center
20 Eye Research Annex Rm 202
21 635 W 165th St.
22 New York, NY 10032
23 +1 (212) 305-8989
24 Email: rla22@cumc.columbia.edu

25
26 **Summary:** We constructed a genotype-phenotype correlation matrix that provides
27 quantifiable probabilities of long-term disease outcomes associated with specific ABCA4
28 genotypes from a large, age-restricted patient cohort

29
30 **Role of funding source:** The funding organizations for this study had no role in its overall
31 design; acquisition and interpretation of data; preparation and revision of the manuscript.

32 **Abstract**

33 Background: >1,500 variants in the *ABCA4* locus underlie a heterogeneous spectrum of
34 retinal disorders ranging from aggressive childhood-onset chorioretinopathy to milder,
35 late-onset macular disease. Genotype-phenotype correlation studies have been limited
36 in clinical applicability as patient cohorts are typically small and seldom capture the full
37 natural history of individual genotypes. To overcome these limitations, we constructed a
38 genotype-phenotype correlation matrix that provides quantifiable probabilities of long-
39 term disease outcomes associated with specific *ABCA4* genotypes from a large, age-
40 restricted patient cohort.

41 Methods: The study included 112 unrelated patients ≥ 50 years of age in whom 2
42 pathogenic variants were identified after sequencing of the *ABCA4* locus. Clinical
43 characterization was performed using the results of best-corrected visual acuity, retinal
44 imaging and full-field electroretinogram testing.

45 Results: Four distinct prognostic groups were defined according to the spatial severity of
46 disease features across the fundus. Recurring genotypes were observed in milder
47 prognoses including those associated with a newly defined class of rare hypomorphic
48 alleles. PVS1 (predicted null) variants were enriched in the most severe prognoses;
49 however, missense variants comprised a larger than expected fraction of these patients.
50 Analysis of allele combinations and their respective prognostic severity, showed that
51 certain variants such as p.(Gly1961Glu), and both rare and frequent hypomorphic
52 alleles, are “clinically dominant” with respect to patient phenotypes irrespective of the
allele in *trans*.

53 Conclusion: These results provide much needed structure to the complex genetic and
54 clinical landscape of ABCA4 disease and adds a tool to the clinical repertoire to
55 quantitatively assess individual genotype-specific prognoses in patients.

56 **Introduction**

57 Pathogenic variants in the *ABCA4* gene are the underlying molecular cause of a large
58 and complex group of autosomal recessive retinal degenerative disorders characterized
59 by progressive loss of central vision.(1) The most well-known phenotype is the
60 eponymous Stargardt disease (STGD1, MIM# 248200),(2) however, advances in genetic
61 screening capabilities, aided by high-resolution diagnostic imaging technology, have
62 broadened the phenotypic profile of *ABCA4* disease to an expansive clinical spectrum
63 encompassing severe, adolescent-onset to mild, late-onset retinal disorders.(3) This
64 phenotypic heterogeneity is matched by an equally extensive array of pathogenic
65 variation across the ~140 kb-spanning *ABCA4* locus (1p22.1). To date, more than 1,500
66 disease-causing variants have been identified in patients.(4) Consistent with the model
67 that clinical phenotypes are dependent on the residual activity of *ABCA4* protein,(5, 6)
68 variants resulting in null alleles such as stop-gain, frameshift, canonical splice site and
69 large copy number variants have been documented in the most severe phenotypes such
70 as cone-rod dystrophy, rapid-onset chorioretinopathy (ROC) and even generalized
71 choriocapillaris dystrophies with retinitis pigmentosa-like features.(6-10)

72 More recently, the complex genetic architecture of milder *ABCA4* disease manifestations
73 has been uncovered. The most frequent pathogenic allele, c.5882G>A p.(Gly1961Glu),
74 is associated with a slow-progressing disease trajectory in patients who often present
75 with transient phenotypes such as bull's eye maculopathy and occult macular
76 dystrophy.(11, 12) Despite being highly prevalent in patients, the disease penetrance of
77 this allele has been disputed as its frequency in the general population is also relatively
78 high (MAF \approx 0.5% in Europeans), and much higher in some ethnic groups.(13, 14) We

79 recently resolved this controversy, at least in most part, by showing that the contribution
80 of an additional deep intronic variant, c.769-784C>T,(15, 16) present in *cis*, is required
81 for clinical penetrance, particularly in homozygotes.(17) Alleles causing late-onset
82 ABCA4 disease, such as c.5603A>T (p.(Asn1868Ile)) and c.4253+43G>A, occur at even
83 higher frequencies in the general population of European descent (up to 7% MAF) and,
84 unlike p.(Gly1961Glu) and other disease alleles, are only clinically penetrant under the
85 condition that the allele in *trans* is sufficiently deleterious.(18, 19)

86 Steady progress in defining genotype-phenotype correlations has been made and the
87 addition of such knowledge to the medical repertoire has inarguably elevated the clinical
88 care of patients. Studies to date have often relied on cross-sectional cohorts of a patient
89 population that include all age-groups. As a result, the correlated “phenotype” studied is
90 often a stage-specific feature, e.g., bull’s eye maculopathy, occult macular dystrophy, etc.
91 Such information, while no doubt useful at the diagnostic stage, is not informative of an
92 individual patient’s long-term prognosis. To address this issue, we constructed a
93 genotype-phenotype correlation matrix based on the most temporally advanced
94 phenotypes of 112 patients aged 50 years or older who have 2 confirmed pathogenic
95 variants in *ABCA4* coupled with comprehensive clinical characterization. We also re-
96 classified many frequent disease-causing alleles thereby further clarifying the impact of
97 *ABCA4* variants on clinical outcome. Our findings provide structure to the complex
98 genotype-phenotype correlation landscape of *ABCA4* disease and establish a
99 quantitative approach for predicting the prognosis of individual patients by clinicians and
100 genetic counselors and for assessing the severity of pathogenic variants. The prognostic

101 matrix will also aid in selecting specific patient groups for clinical trials, depending on the
102 specific therapeutic application.

103

104 **Results**

105 ***Four clinically defined prognostic outcomes of ABCA4 disease***

106 Demographic, clinical, and genetic characteristics of all 112 patients in the study are
107 summarized in Supplemental Table 1. Clinical data from the most recent visit for each
108 patient were used in the study. Each patient was categorized into one of four “Prognosis”
109 categories based on the observable spatial progression of ABCA4-associated disease
110 features in the fundus (Figure 1A) by age 50 years or older. Patients categorized as
111 having Prognosis 1 (n=28) had the mildest disease outcome (in the cohort) manifesting
112 early RPE atrophy within the central macula without any apparent pisciform flecks.
113 Patients with Prognosis 2 (n=31) were at a more progressed stage of chorioretinal atrophy
114 across the macula and developed nascent flecks that appeared outside the vascular
115 arcades (Figure 1A, yellow arrowheads). All patients with Prognosis 3 (n=20) had multi-
116 focal regions of chorioretinal atrophy which, in some cases, extended beyond the macula
117 and exhibited a pattern of highly confluent flecks in non-atrophic regions. Patients with
118 Prognosis 4 (n=33) progressed to the stage characterized by the large atrophic,
119 coalescing lesions across the entire posterior pole.

120 There were no significant differences in the mean age of patients between Prognosis
121 groups (Supplemental Data 1). The mean age of symptomatic onset was earliest among
122 patients with Prognosis 4 (17.1 years) compared to the milder prognostic groups which
123 had peak distributions at 41.7 years (Prognosis 1) and 40.9 years (Prognosis 2) due to
124 the large number of late-onset disease cases in these latter groups (Figure 1B, see
125 Supplemental Data 2). Best-corrected visual acuity (BCVA) from the most recent visits
126 were also poorest among patients with Prognosis 4 of which ~40% were counting fingers

127 or worse ($P < 0.00001$) (Figure 1C, see Supplemental Data 3). Comparatively, BCVA
128 distributions were multi-modal among patients with Prognosis 1-3 most of whom had
129 20/200 (logMAR 1.00) or worse and 20/20 in cases with foveal sparing. Full-field ERG
130 responses were largely unremarkable in Prognosis 1 and 2 (Figure 1D). Significant
131 defects were found in Prognosis 3 (50% Lois Group II) and Prognosis 4 (93% Lois Group
132 III). There were no significant differences in mean age of patients in across Prognosis
133 groups ($P = 0.254$) (Supplemental Table 3).

134 ***Classification of p.(Gly1961Glu), p.(Asn1868Ile) and a new class of rare***
135 ***hypomorphic alleles***

136 Genotypes consisting of the major disease-causing allele, p.(Gly1961Glu) and the
137 frequent hypomorphic allele, p.(Asn1868Ile), were the most prominent variants among
138 the mild phenotypes, together accounting for 56% of patients in Prognosis 1 and
139 Prognosis 2 (Figure 2A). Despite the advanced age of this cohort, 3 patients (P12, P18
140 and P20) presented with early stage bull's eye maculopathy (Supplemental Figure 1A-
141 1C). As we have previously shown, p.(Asn1868Ile) is highly associated with foveal
142 sparing which is a major contributing factor to the delayed symptomatic onset age in most
143 patients (Figure 2B).(19) Among the remaining patients in the mild Prognosis categories,
144 we identified another group of patients with 6 recurring alleles, p.(Ala1038Val),
145 c.4253+43G>A,(18) p.(Pro1486Leu), p.(Thr1526Met), p.(Ile1562Thr), p.(Arg2030Gln),
146 that have features in common with p.(Asn1868Ile), most notably, delayed symptomatic
147 onset due to foveal sparing (Table 1, Figure 2C and 2D). Disease features in the fundus
148 of these cases were confined to a delineable area around the vascular arcades in a
149 reticular appearance studded along the peripheral boundary with elongated "tails"

150 projecting eccentrically in a radial pattern (Figure 3A and 3B and Figure 4A and 4B).
151 Generalized dysfunction of the cone and rod systems were not detected on full-field
152 electroretinogram (ffERG) testing (Figure 3C). Although each of these variants is
153 exceedingly rare in the general population ($0.005 > \text{MAF} > 0.00005$), unaffected
154 homozygotes have been reported for p.(Ala1038Val), p.(Ile1562Thr) and
155 c.[4253+43G>A] resulting in some cases conflicting interpretations of pathogenicity
156 (Supplemental Table 2). Furthermore, as has been observed with p.(Asn1868Ile), the
157 allele *in trans* in these genotypes are mostly loss-of-function alleles, including an 8.4 kb
158 deletion that was identified in P58 (Figure 4C). Considering these differences, we
159 separated these mild *ABCA4* alleles into three classes: p.(Gly1961Glu), Frequent
160 hypomorph and Rare hypomorph.

161 ***Classification of PVS1 and “severe” non-PVS1 alleles***

162 The distribution of PVS1 (i.e., null or loss-of-function alleles) (Table 1) were skewed
163 towards the most severe clinical phenotypes although at a lower-than-expected
164 proportion. Genotypes with a PVS1 allele comprised ~1/3 of Prognosis 3 and Prognosis
165 4 cases while the remaining ~2/3 fraction consisted mostly of missense variants and, in
166 part, functionally validated deep intronic and synonymous variants. The majority of these
167 missense alleles have been observed to be the causal allele *in trans* from
168 p.(Asn1868Ile). (19) Using our current dataset, we further classified 5 additional alleles,
169 p.(Thr1019Met), p.(Ala1598Asp) p.([Asp1532Asn;Asn1868Ile]),
170 p.([Gly863Ala;Asn1868Ile]) and c.5714+5G>A, as severe based on their recurrence in
171 compound heterozygous and/or homozygous patients with Prognosis 3 or Prognosis 4.

172 To distinguish these severe non-PVS1 alleles from moderate/milder alleles, we grouped
173 them into separate “severe” sub-class (Table 1).

174 ***Classification of moderate variants***

175 After classifying 65% of alleles in the study cohort as either mild or severe, a remaining
176 group of 36 unique variants (35%, 56 total alleles) did not meet any of the aforementioned
177 classification criteria. These alleles were uniformly distributed across Prognosis
178 categories as compared to the other classified allele groups which skewed accordingly
179 towards mild or severe Prognoses (Figure 5A). The coding effect in 93% of these alleles
180 is missense (Figure 5B). The three non-missense variants in this group were an exonic
181 in-frame duplication, deep intronic 15 nucleotide deletion and the known c.859-9T>C
182 variant, which prior midi-gene studies in HEK293T cells have determined to have a
183 “moderate” effect as the variant results in 75% of wild-type *ABCA4* RNA.⁽¹⁶⁾ Considering
184 the nonspecific genetic attributes of these alleles and their collectively uniform distribution
185 across Prognosis categories, we classified them in a “Moderate” group.

186 ***Construction of a genotype-phenotype correlation matrix***

187 We generated probability matrices representing correlations between the four clinical
188 Prognosis categories (Prognosis 1-4) and all possible genotypic combinations for the
189 following allele classes: p.(Gly1961Glu), Frequent hypomorph, Rare hypomorph,
190 Moderate, Severe and PVS1 (Table 1, Figure 6). Genotypes consisting of either a
191 p.(Gly1961Glu), Frequent Hypomorphic or Rare Hypomorphic allele had the mildest
192 prognostic outcomes with most cases having either Prognosis 1 or Prognosis 2 (Figure
193 6A-6C). Genotypes of these three allele classes were also the least heterogenous in

194 terms of prognostic distribution ($P = 0.1164$, two-sided FET, see Supplemental Data 4)
195 compared to both Moderate and Severe/PVS1 genotypes ($P < 0.001$, two-sided FET, see
196 Supplemental Data 4). This is due at least in part to the absence of homozygotes and
197 cases with other mild allele combinations. The apparent non-penetrance, coupled with
198 the consistent clinical phenotype, suggests that these three allele classes exhibit a form
199 of “clinical dominance” whereby the allele in *trans*, while necessary for disease
200 expression, has minimal to no effect on the phenotypic variability.

201 Conversely, all Prognosis categories were represented in Moderate, Severe and PVS1
202 allele combinations and the additive severity of the allele in *trans* strongly correlated with
203 prognostic severity for these allele combinations. For instance, Moderate allele
204 genotypes with another Moderate allele in *trans* give a 43% probability of having
205 Prognosis 1 whereas having a p.(Gly1961Glu) allele in *trans* increases the Prognosis 1
206 probability to 83% and having Severe or PVS1 allele in *trans* reduces the Prognosis 1
207 probability to 0-12% and increases the probability of Prognosis 3 and 4 to 44-55% (Figure
208 6D). Similar trends were true for both Severe and PVS1 genotypes. Prognosis
209 correlations between PVS1 and Severe allele genotypes were also remarkably similar
210 suggesting very little clinical distinction between the two allele classes (Figure 6E and
211 6F). To simplify these observations for clinical applicability, we excluded allele
212 combinations that were not present in patients for any prognosis, thereby collapsing each
213 allele matrix into only those representing the genotypes of all patients across the study
214 cohort (Figure 7).

215

216 **Discussion**

217 Advances in genomic medicine in recent decades have allowed genetic testing in the
218 clinic to be a routine option for patients with monogenic diseases. While this has
219 undoubtedly improved the standard-of-care for patients, the utility of a genetic result rarely
220 extends beyond diagnostic confirmation. The underleveraging of variant level insight in
221 the clinic is attributable to the lack of concrete genotype-phenotype correlations that are
222 difficult to assess for several reasons. First, Mendelian disorders like *ABCA4* disease are
223 both rare and profoundly heterogeneous. Prior studies have noted strong trends with
224 specific alleles,(12, 20-24) however, most cohorts are typically insufficient in size and
225 scope to make conclusions that are applicable to clinical care. Moreover, cross-sectional
226 study cohorts themselves are demographically heterogeneous, particularly in terms of
227 age, adding further limitations such as unknown disease trajectory and clinical outcome
228 of younger patients.

229 The large clinical and genetic repository we have built over 20+ years has allowed us to
230 overcome most of these issues. Using the well-characterized clinical data of an age-
231 restricted (≥ 50 years of age) cohort of 112 patients, we were able to precisely dissect
232 apart the complex genotype-phenotype correlation landscape of *ABCA4* disease in a
233 quantitative manner which can be immediately used to assess and predict the long-term
234 prognosis of patients following genetic testing. The correlation matrix can be improved
235 upon by the addition of more cases in follow-up studies to increase statistical power and
236 accommodate other *ABCA4* variants not described in this study. These data also provide
237 precise insight into magnitude differences in disease severity between different alleles
238 which should be considered in the selection of patients for clinical trials.

239 These data can also be used to clinical classify the pathogenicity of different *ABCA4*
240 alleles. Analyzing patients with the mildest prognoses, for instance, identified a class of
241 rare hypomorphic variants that exhibit clinical overlap with p.(Asn1868Ile) cases,
242 including slow-progressing disease and persistent sparing of the fovea. Results of prior
243 functional and clinical studies of these variants were also consistent with mild
244 characterization. For instance, transgenic expression of human p.(Ala1038Val) in both *X.*
245 *laevis* tadpole retinae and HEK293T cells revealed no observable defects in sub-cellular
246 localization.(25, 26) The latter study also showed that p.(Ala1038Val) mutant structure
247 closely resembles the WT *ABCA4* structure using single particle analysis (cryo-EM).(26)
248 The clinical phenotypes of all well-characterized patients harboring p.(Ala1038Val),(26)
249 p.(Arg2030Gln),(27, 28) p.(Pro1486Leu),(29) p.(Thre1526Met)(27, 28, 30) and
250 p.(Ile1562Thr)(30, 31) alleles in the literature are also consistent with milder disease in
251 general and with specific hypomorphic features.

252 These and other mild *ABCA4* alleles, including p.(Gly1961Glu) and the frequent
253 hypomorph p.(Asn1868Ile), also exhibit some collective characteristics that are
254 inconsistent with most autosomal recessive diseases. Under an additive pathogenicity
255 model which has been proposed for *ABCA4*,(32) patient phenotypes are expected to vary
256 according to the combined effects of both *ABCA4* alleles and indeed, the phenotypic
257 outcome of Moderate and Severe/PVS1 alleles vary widely depending on the allele in
258 *trans* (Figure 5D-5F). Mild alleles, however, appear to be “clinically dominant” in that all
259 genotypes are invariably mild in overall severity (long-term prognosis) and additionally,
260 each respective allele has unique and consistent sub-phenotypic features (e.g., foveal
261 sparing (hypomorphs) and optical gap (p.(Gly1961Glu))) irrespective of the type of allele

262 in *trans*. This phenomenon may be partially explained by the non-penetrance of mild
263 genotypes resulting in a more “homogeneous” genotype combinations in observed cases.
264 The underlying mechanisms resulting in sub-phenotypes, while of diagnostic value for
265 solving cases without genetic confirmation, remain unknown.

266 Our analysis also re-classified several non-PVS1 alleles as clinically severe such as the
267 c.5714+5G>A substitution in intron 40 which was previously reported to have a
268 “moderate” effect as it results in ~39% correctly spliced mRNA in HEK293T cells.(16)
269 Consistent with other clinical studies,(6, 28, 33, 34) we also found the variant to be
270 exclusively associated with severe phenotypes (compound heterozygous in two patients
271 with Prognosis 3 and three patients with Prognosis 4) which led us to the conclude that
272 the allele is at least clinically severe in patients. Several other missense alleles were also
273 classified as severe, including p.(Ala1598Asp), p.(Thr1019Met), and
274 p.([Asp1532Asn;Asn1868Ile]), based on their recurrence in patients (including
275 homozygotes) with Prognosis 3 and 4. These, and a large group of other missense alleles,
276 comprised an unexpectedly large proportion of genotypes leading to the most severe
277 prognostic outcomes. This observation should caution against the common interpretation
278 that most missense variants, at least in the *ABCA4* gene, are less severe than PVS1
279 variants.

280 This study has several limitations. While the patient cohort is large considering the rarity
281 of this disease, not all possible *ABCA4* genotypes are represented. Notably, biallelic
282 PVS1 genotypes which are known to underlie the most severe *ABCA4* disease
283 phenotypes such as RP-like, ROC and cone-rod dystrophy were not included.(6-10)
284 Prognostic assessment in these cases, however, usually unambiguous as visual

285 deterioration begins early in life and disease progresses rapidly. The four prognostic
286 classifications defined in the study may also not fully represent the breadth of clinical
287 outcomes in ABCA4 disease. Further studies based on our study design in larger,
288 preferably multi-ethnic, cohorts of more comprehensively characterized patients would
289 help address many of the current limitations and critically advance precision medicine for
290 ABCA4 disease. In summary, we constructed a genotype-phenotype matrix based on
291 the long-term prognostic outcomes of 112 genetically confirmed patients with ABCA4
292 disease patients. Two major disease-causing variants of ABCA4, p.Gly1961Glu) and
293 p.(Asn1868Ile) accounted for more than half of the genotypes (patients) with mildest
294 prognoses. We also identified new class of rare hypomorphic variants among mild
295 prognoses cases which, together with p.Gly1961Glu) and p.(Asn1868Ile), exhibit “clinical
296 dominance” in their consistent clinical features irrespective of the allele in *trans*. We
297 identify a large group of missense variants that are associated with the more severe
298 prognoses, and clinically re-classified others including c.5714+5G>A that were previously
299 suggested to be non-severe. The genotype-phenotype correlation matrix provides
300 prognostic probabilities based on underlying *ABCA4* genotype and can be used as a tool
301 assess disease severity in patients and as a framework for designing and selecting of
302 patients for clinical trials.

303 **Methods**

304 ***Study subjects and clinical characterization***

305 Patients diagnosed with Stargardt or *ABCA4*-related disease were recruited from the
306 Department of Ophthalmology at Columbia University Irving Medical Center. In total, 112
307 unrelated patients harboring two pathogenic variants in *ABCA4* and ≥ 50 years of age

308 were included in the study. The lower age limit threshold of 50 years was chosen to
309 ensure that all major genotype groups were accommodated in the analysis, particularly
310 patients with the common hypomorphic allele, p.(Asn1868Ile) whose median age of
311 symptomatic onset is ~35 years (interquartile range 28-48 years).(19) Each patient
312 underwent a complete ophthalmic examination by a retinal physician (SHT), which
313 included slit-lamp and dilated fundus examination, best corrected visual acuity (BCVA;
314 Snellen), color fundus photography, fundus autofluorescence (AF, 488-nm, 532-nm and
315 787-nm), spectral domain-optical coherence tomography (SD-OCT) scanning and full-
316 field electroretinogram (ffERG) testing. Conversion of “counting fingers” (CF) and “hand
317 motion” (HM) to logMAR units were calculated in accordance with Schulze-Bonsel et
318 al.(35) In short, CF was replaced with the calculated decimal acuity of 0.014 which
319 corresponds to approximately Snellen 20/1500 or logMAR 1.875; HM was replaced with
320 the decimal acuity of 0.005 which corresponds to approximately Snellen 20/4000 or
321 logMAR 2.300.

322 Imaging across all modalities were conducted following pupil dilation (>7mm) with
323 tropicamide (1%) and phenylephrine hydrochloride (2.5%). Fundus autofluorescence
324 (488-nm) images and 9mm horizontal foveal SD-OCT scans were acquired with the
325 Spectralis HRA+OCT (Heidelberg Engineering, Heidelberg, Germany). Ultra-widefield
326 autofluorescence images were acquired with an Optos 200 Tx (Optos PLC, Dunfermline,
327 United Kingdom). Full-field electroretinograms (ffERGs) were recorded silver
328 impregnated fiber electrodes (DTL; Diagnosys LLC, Littleton, MA) on the Espion Visual
329 Electrophysiology System (Diagnosys LLC, Littleton, MA, USA) in accordance with
330 International Society for Clinical Electrophysiology of Vision (ISCEV) standards.(36)

331 ffERG classifications were assigned according to electrophysiological attributes
332 described by Lois et al.(23, 37) Group 1 is characterized by no detectable loss in scotopic
333 or photopic function; Group 2 is characterized by photopic loss, but normal scotopic
334 function; and Group 3 exhibits deterioration of both scotopic and photopic function.

335 Prognosis classifications (I, II, III or IV) were determined by two independent graders
336 (W.L. and P.Y.S.) using 55° AF (488-nm) images of each eye for all study patients. In
337 patients with inter-ocular discordance, the Prognosis classification was assigned
338 according to the more advanced eye. Discordant evaluations between graders were
339 adjudicated by an additional grader (S.H.T.). Notes from the corresponding clinical exam,
340 which included direct and indirect ophthalmoscopy details, were reviewed to confirm the
341 final Prognosis group assignment in each patient. All three graders were blinded to the
342 *ABCA4* genotype of each patient at the time of Prognosis classification.

343 ***Molecular analyses***

344 Screening of the *ABCA4* gene was performed by next-generation sequencing (NGS) as
345 previously described.(38, 39) All detected possibly disease-associated variants were
346 confirmed by Sanger sequencing and analyzed with Alamut software® (Interactive
347 Biosoftware). Segregation of the new variants with the disease was analyzed in families
348 if family members were available. Functional annotation of variants was determined using
349 computational software including ANNOVAR(40) using pathogenicity scores of M-CAP,
350 REVEL, Eigen, and CADD (v1.6). As a general guideline, pathogenic consequences are
351 predicted for variants with scores over 0.025 for MCAP, 0.5 for REVEL, 0.5 for Eigen and
352 20 for CADD. The allele frequencies of all variants were compared to those in the
353 Genome Aggregation Database (gnomAD) (accessed October 2021).

354 **Statistics**

355 A detailed summary of all statistic calculations is provided in the Appendix. Comparison
356 of mean characteristics between prognosis categories were determined by a One-way
357 ANOVA test with post-hoc Tukey HSD and Kruskal-Wallis test. Significance was set at
358 alpha level <0.05. Density plots were generated using the ggridges package in R version
359 4.0.4. Fisher's Exact Tests for Count Data (2x3 contingency table) were used to compare
360 the distributions of mild, moderate and severe allele combinations across Prognosis
361 categories.

362 **Study approval**

363 All study procedures were defined under protocol #AAAI9906 approved by the
364 Institutional Review Board at Columbia University Medical Center. The study adhered to
365 tenets set out in the Declaration of Helsinki.

366

367 **Conflicts of interests**

368 S.H.T. has received support from Abeona Therapeutics, Inc. and is a board member of
369 Emendo Biotherapeutics, Nanoscope and Rejuvitas, Inc. The other authors declare no
370 competing interests.

371

372 **Author Contributions**

373 W.L. designed the study, recruited study subjects, acquired and analyzed clinical data
374 and wrote the manuscript; J.Z. performed sequencing, analyzed molecular data and
375 critically revised the manuscript; P.Y.S. recruited subjects and acquired clinical data, T.N.
376 assisted with molecular analyses, S.H.T. clinically examined study subjects, R.A.,
377 supervised the study, critically revised the manuscript and obtained research funding.

378

379 **Funding**

380 This work was supported, in part, by the National Eye Institute, NIH grants R01
381 EY028203, R01 EY028954, R01 EY029315, P30 19007 (Core Grant for Vision
382 Research), the Foundation Fighting Blindness USA, grant no. PPA-1218-0751-COLU,
383 and the unrestricted grant to the Department of Ophthalmology, Columbia University,
384 from Research to Prevent Blindness.

385

386 **Acknowledgements**

387 The authors acknowledge Professor Joseph Terwilliger for helpful discussions and Jimmy
388 Duong and Professor Wei-Yann Tsai of the Biostatistics, Epidemiology, and Research
389 Design (BERD), Irving Institute for Clinical and Translational Research for helpful
390 statistical consultation on the project. Lastly, all authors would like to thank the patients
391 and their families for their immeasurable contributions to our *ABCA4* studies over the
392 years.

393

394 **References**

- 395 1. Allikmets R, Singh N, Sun H, Shroyer NF, Hutchinson A, Chidambaram A, et al.
396 A photoreceptor cell-specific ATP-binding transporter gene (ABCR) is mutated in
397 recessive Stargardt macular dystrophy. *Nat Genet.* 1997;15(3):236-46.
- 398 2. Stargardt K. Über familiäre, progressive degeneration in der maculagegend des
399 auges. *Albrecht von Graefes Arch Klin Ophthalmology.* 1909;71:534–50.
- 400 3. Cremers FPM, Lee W, Collin RWJ, and Allikmets R. Clinical spectrum, genetic
401 complexity and therapeutic approaches for retinal disease caused by ABCA4
402 mutations. *Prog Retin Eye Res.* 2020;79:100861.
- 403 4. Cornelis SS, Bax NM, Zernant J, Allikmets R, Fritsche LG, den Dunnen JT, et al.
404 In Silico Functional Meta-Analysis of 5,962 ABCA4 Variants in 3,928 Retinal
405 Dystrophy Cases. *Hum Mutat.* 2017;38(4):400-8.
- 406 5. Maugeri A, van Driel MA, van de Pol DJ, Klevering BJ, van Haren FJ, Tijmes N,
407 et al. The 2588G-->C mutation in the ABCR gene is a mild frequent founder
408 mutation in the Western European population and allows the classification of
409 ABCR mutations in patients with Stargardt disease. *Am J Hum Genet.*
410 1999;64(4):1024-35.
- 411 6. Cremers FP, van de Pol DJ, van Driel M, den Hollander AI, van Haren FJ,
412 Knoers NV, et al. Autosomal recessive retinitis pigmentosa and cone-rod
413 dystrophy caused by splice site mutations in the Stargardt's disease gene ABCR.
414 *Hum Mol Genet.* 1998;7(3):355-62.
- 415 7. Tanaka K, Lee W, Zernant J, Schuerch K, Ciccone L, Tsang SH, et al. The
416 Rapid-Onset Chorioretinopathy Phenotype of ABCA4 Disease. *Ophthalmology.*
417 2018;125(1):89-99.

- 418 8. Martinez-Mir A, Paloma E, Allikmets R, Ayuso C, del Rio T, Dean M, et al.
419 Retinitis pigmentosa caused by a homozygous mutation in the Stargardt disease
420 gene ABCR. *Nat Genet.* 1998;18(1):11-2.
- 421 9. Lee W, Zernant J, Nagasaki T, Tsang SH, and Allikmets R. Deep Scleral
422 Exposure: A Degenerative Outcome of End-Stage Stargardt Disease. *Am J*
423 *Ophthalmol.* 2018;195:16-25.
- 424 10. Bertelsen M, Zernant J, Larsen M, Duno M, Allikmets R, and Rosenberg T.
425 Generalized choriocapillaris dystrophy, a distinct phenotype in the spectrum of
426 ABCA4-associated retinopathies. *Invest Ophthalmol Vis Sci.* 2014;55(4):2766-76.
- 427 11. Noupuu K, Lee W, Zernant J, Tsang SH, and Allikmets R. Structural and genetic
428 assessment of the ABCA4-associated optical gap phenotype. *Invest Ophthalmol*
429 *Vis Sci.* 2014;55(11):7217-26.
- 430 12. Cella W, Greenstein VC, Zernant-Rajang J, Smith TR, Barile G, Allikmets R, et
431 al. G1961E mutant allele in the Stargardt disease gene ABCA4 causes bull's eye
432 maculopathy. *Exp Eye Res.* 2009;89(1):16-24.
- 433 13. Guymer RH, Heon E, Lotery AJ, Munier FL, Schorderet DF, Baird PN, et al.
434 Variation of codons 1961 and 2177 of the Stargardt disease gene is not
435 associated with age-related macular degeneration. *Arch Ophthalmol.*
436 2001;119(5):745-51.
- 437 14. Lee W, Schuerch K, Zernant J, Collison FT, Bearely S, Fishman GA, et al.
438 Genotypic spectrum and phenotype correlations of ABCA4-associated disease in
439 patients of south Asian descent. *Eur J Hum Genet.* 2017;25(6):735-43.

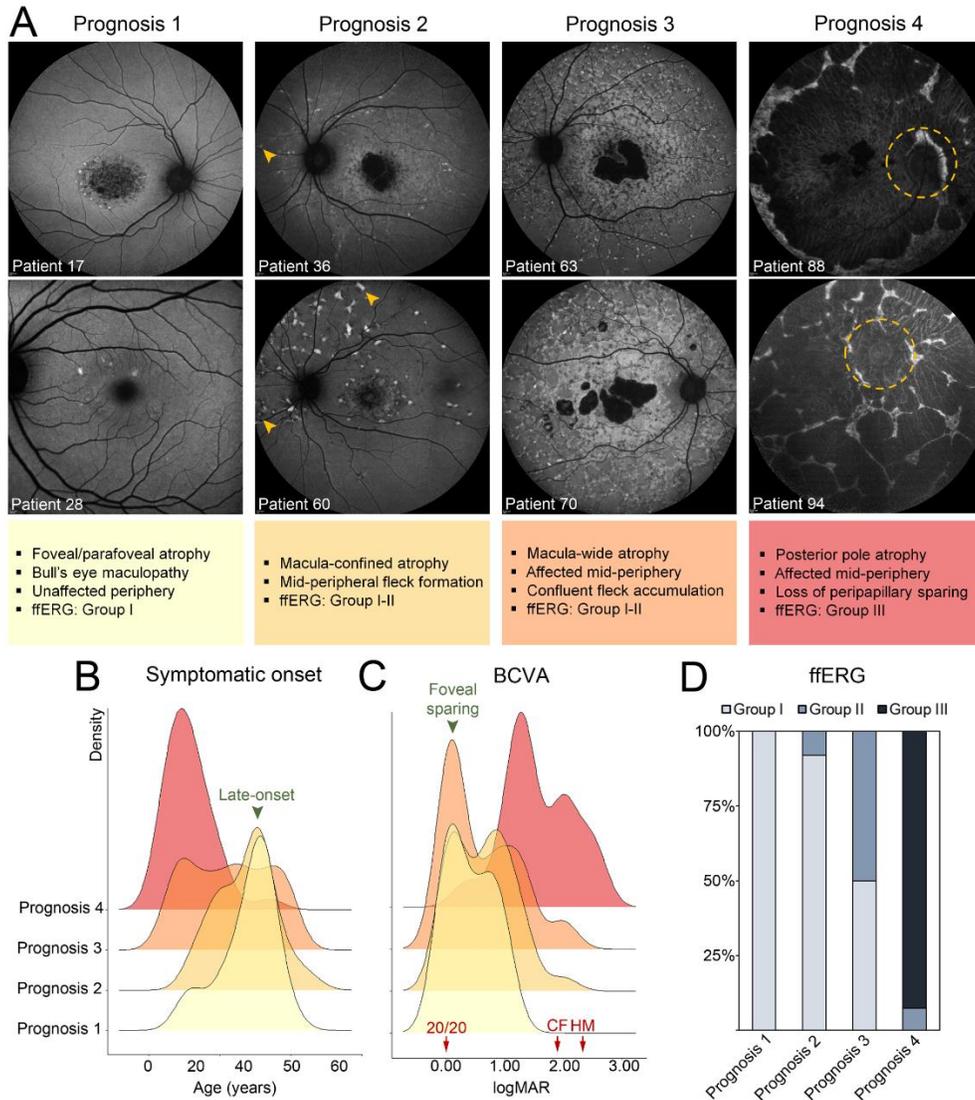
- 440 15. Sangermano R, Garanto A, Khan M, Runhart EH, Bauwens M, Bax NM, et al.
441 Deep-intronic ABCA4 variants explain missing heritability in Stargardt disease
442 and allow correction of splice defects by antisense oligonucleotides. *Genet Med.*
443 2019;21(8):1751-60.
- 444 16. Sangermano R, Khan M, Cornelis SS, Richelle V, Albert S, Garanto A, et al.
445 ABCA4 midigenes reveal the full splice spectrum of all reported noncanonical
446 splice site variants in Stargardt disease. *Genome Res.* 2018;28(1):100-10.
- 447 17. Lee W, Zernant J, Nagasaki T, Molday LL, Su PY, Fishman GA, et al. Cis-acting
448 modifiers in the ABCA4 locus contribute to the penetrance of the major disease-
449 causing variant in Stargardt disease. *Hum Mol Genet.* 2021.
- 450 18. Zernant J, Lee W, Nagasaki T, Collison FT, Fishman GA, Bertelsen M, et al.
451 Extremely hypomorphic and severe deep intronic variants in the ABCA4 locus
452 result in varying Stargardt disease phenotypes. *Cold Spring Harb Mol Case Stud.*
453 2018;4(4).
- 454 19. Zernant J, Lee W, Collison FT, Fishman GA, Sergeev YV, Schuerch K, et al.
455 Frequent hypomorphic alleles account for a significant fraction of ABCA4 disease
456 and distinguish it from age-related macular degeneration. *J Med Genet.*
457 2017;54(6):404-12.
- 458 20. Di Iorio V, Orrico A, Esposito G, Melillo P, Rossi S, Sbordone S, et al.
459 ASSOCIATION BETWEEN GENOTYPE AND DISEASE PROGRESSION IN
460 ITALIAN STARGARDT PATIENTS: A Retrospective Natural History Study.
461 *Retina.* 2019;39(7):1399-409.

- 462 21. Del Pozo-Valero M, Riveiro-Alvarez R, Blanco-Kelly F, Aguirre-Lamban J, Martin-
463 Merida I, Iancu IF, et al. Genotype-Phenotype Correlations in a Spanish Cohort
464 of 506 Families With Biallelic ABCA4 Pathogenic Variants. *Am J Ophthalmol.*
465 2020;219:195-204.
- 466 22. Lewis RA, Shroyer NF, Singh N, Allikmets R, Hutchinson A, Li Y, et al.
467 Genotype/Phenotype analysis of a photoreceptor-specific ATP-binding cassette
468 transporter gene, ABCR, in Stargardt disease. *Am J Hum Genet.*
469 1999;64(2):422-34.
- 470 23. Fujinami K, Lois N, Davidson AE, Mackay DS, Hogg CR, Stone EM, et al. A
471 longitudinal study of stargardt disease: clinical and electrophysiologic
472 assessment, progression, and genotype correlations. *Am J Ophthalmol.*
473 2013;155(6):1075-88 e13.
- 474 24. Fujinami K, Lois N, Mukherjee R, McBain VA, Tsunoda K, Tsubota K, et al. A
475 longitudinal study of Stargardt disease: quantitative assessment of fundus
476 autofluorescence, progression, and genotype correlations. *Invest Ophthalmol Vis*
477 *Sci.* 2013;54(13):8181-90.
- 478 25. Wiszniewski W, Zaremba CM, Yatsenko AN, Jamrich M, Wensel TG, Lewis RA,
479 et al. ABCA4 mutations causing mislocalization are found frequently in patients
480 with severe retinal dystrophies. *Hum Mol Genet.* 2005;14(19):2769-78.
- 481 26. Zhang N, Tsybovsky Y, Kolesnikov AV, Rozanowska M, Swider M, Schwartz SB,
482 et al. Protein misfolding and the pathogenesis of ABCA4-associated retinal
483 degenerations. *Hum Mol Genet.* 2015;24(11):3220-37.

- 484 27. Cideciyan AV, Swider M, Aleman TS, Tsybovsky Y, Schwartz SB, Windsor EA, et
485 al. ABCA4 disease progression and a proposed strategy for gene therapy. *Hum*
486 *Mol Genet.* 2009;18(5):931-41.
- 487 28. Fakin A, Robson AG, Chiang JP, Fujinami K, Moore AT, Michaelides M, et al.
488 The Effect on Retinal Structure and Function of 15 Specific ABCA4 Mutations: A
489 Detailed Examination of 82 Hemizygous Patients. *Invest Ophthalmol Vis Sci.*
490 2016;57(14):5963-73.
- 491 29. Riveiro-Alvarez R, Lopez-Martinez MA, Zernant J, Aguirre-Lamban J,
492 Cantalapiedra D, Avila-Fernandez A, et al. Outcome of ABCA4 disease-
493 associated alleles in autosomal recessive retinal dystrophies: retrospective
494 analysis in 420 Spanish families. *Ophthalmology.* 2013;120(11):2332-7.
- 495 30. Shroyer NF, Lewis RA, Yatsenko AN, Wensel TG, and Lupski JR. Cosegregation
496 and functional analysis of mutant ABCR (ABCA4) alleles in families that manifest
497 both Stargardt disease and age-related macular degeneration. *Hum Mol Genet.*
498 2001;10(23):2671-8.
- 499 31. Yatsenko AN, Wiszniewski W, Zaremba CM, Jamrich M, and Lupski JR.
500 Evolution of ABCA4 proteins in vertebrates. *J Mol Evol.* 2005;60(1):72-80.
- 501 32. Schindler EI, Nylén EL, Ko AC, Affatigato LM, Heggen AC, Wang K, et al.
502 Deducing the pathogenic contribution of recessive ABCA4 alleles in an outbred
503 population. *Hum Mol Genet.* 2010;19(19):3693-701.
- 504 33. Green JS, O'Rielly DD, Pater JA, Houston J, Rajabi H, Galutira D, et al. The
505 genetic architecture of Stargardt macular dystrophy (STGD1): a longitudinal 40-
506 year study in a genetic isolate. *Eur J Hum Genet.* 2020;28(7):925-37.

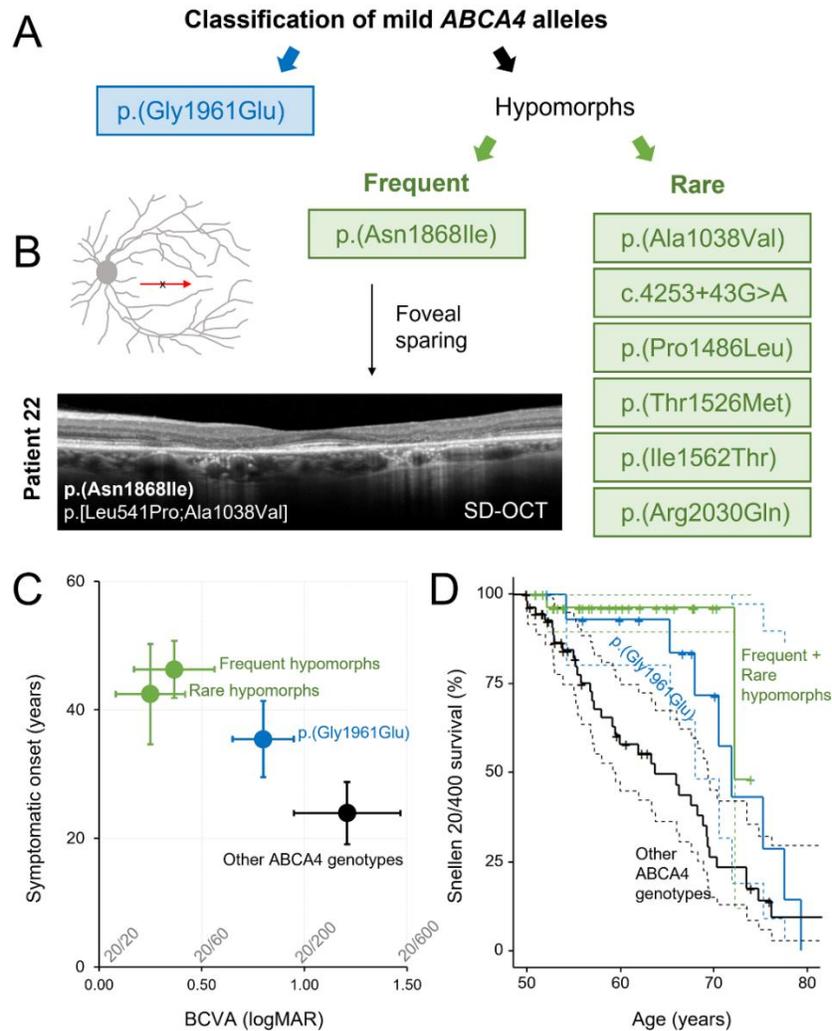
- 507 34. Klevering BJ, van Driel M, van de Pol DJ, Pinckers AJ, Cremers FP, and Hoyng
508 CB. Phenotypic variations in a family with retinal dystrophy as result of different
509 mutations in the ABCR gene. *Br J Ophthalmol*. 1999;83(8):914-8.
- 510 35. Schulze-Bonsel K, Feltgen N, Burau H, Hansen L, and Bach M. Visual acuities
511 "hand motion" and "counting fingers" can be quantified with the freiburg visual
512 acuity test. *Invest Ophthalmol Vis Sci*. 2006;47(3):1236-40.
- 513 36. McCulloch DL, Marmor MF, Brigell MG, Hamilton R, Holder GE, Tzekov R, et al.
514 ISCEV Standard for full-field clinical electroretinography (2015 update). *Doc*
515 *Ophthalmol*. 2015;130(1):1-12.
- 516 37. Lois N, Holder GE, Bunce C, Fitzke FW, and Bird AC. Phenotypic subtypes of
517 Stargardt macular dystrophy-fundus flavimaculatus. *Arch Ophthalmol*.
518 2001;119(3):359-69.
- 519 38. Zernant J, Schubert C, Im KM, Burke T, Brown CM, Fishman GA, et al. Analysis
520 of the ABCA4 gene by next-generation sequencing. *Invest Ophthalmol Vis Sci*.
521 2011;52(11):8479-87.
- 522 39. Zernant J, Xie YA, Ayuso C, Riveiro-Alvarez R, Lopez-Martinez MA, Simonelli F,
523 et al. Analysis of the ABCA4 genomic locus in Stargardt disease. *Hum Mol*
524 *Genet*. 2014;23(25):6797-806.
- 525 40. Wang K, Li M, and Hakonarson H. ANNOVAR: functional annotation of genetic
526 variants from high-throughput sequencing data. *Nucleic Acids Res*.
527 2010;38(16):e164.

528



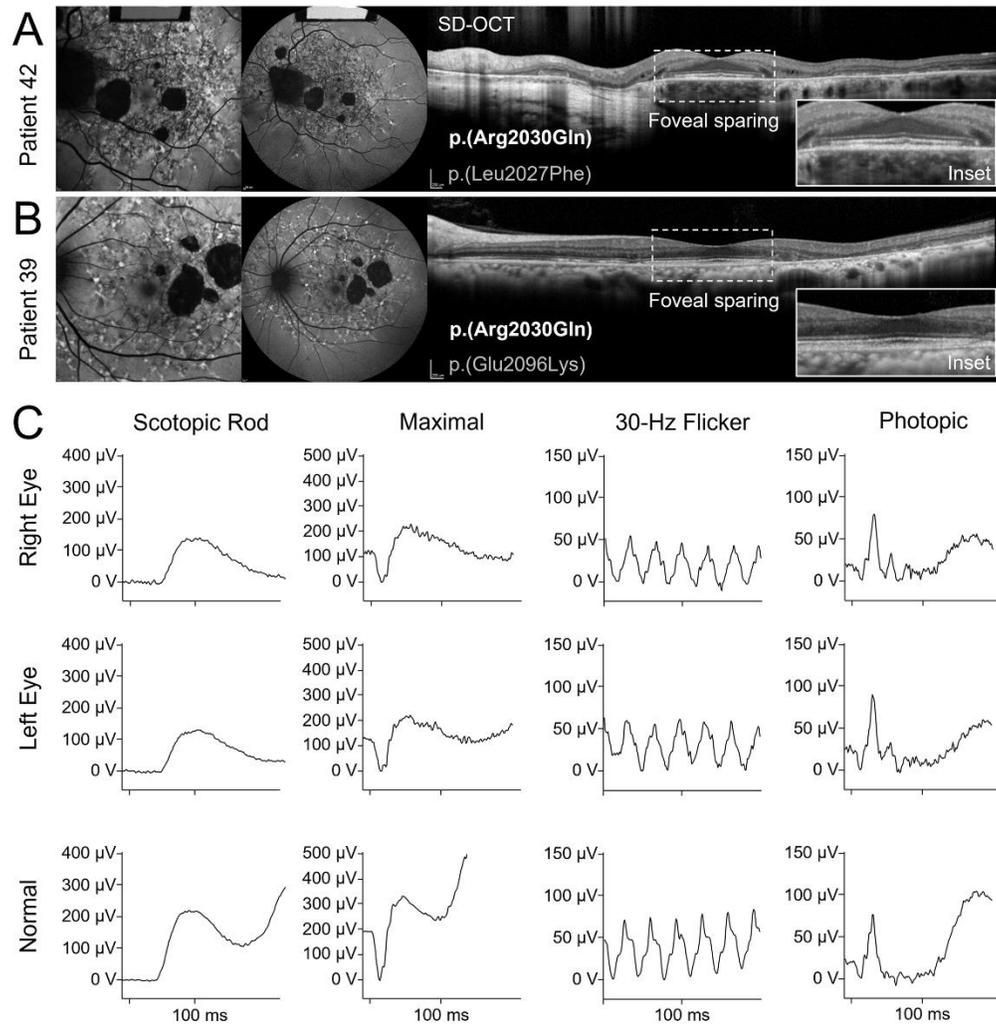
529

530 **Figure 1:** Clinical characteristics of four prognostic outcomes observed in 112 patients
 531 (>50 years of age) with ABCA4 disease. Each prognosis was defined according to
 532 observable spatial progression of disease features detected at the most recent visit in
 533 each patient. (A) Representative autofluorescence images and clinical descriptions of
 534 patients in each Prognosis classification. Extramacular development of flecks in Prognosis
 535 2 are indicated by yellow arrow heads. The position of the optic nerve is encircled by the
 536 dotted yellow line. (B) Ridgeline plots of the distribution of ages at which visual symptoms
 537 of all patients were first reported for each prognosis category (bandwidth = 5.83). (C)
 538 Density plots of the best-corrected visual acuity (BCVA) of the least-impaired eye of all
 539 patients (bandwidth = 0.246). BCVA were presented in logMAR units with corresponding
 540 Snellen equivalents (20/20, counting fingers (CF) and hand motion (HM), red arrows)
 541 provided. (D) Proportion of full-field electroretinogram (ffERG) groupings according to the
 542 classification by Lois et al.(37) for each prognosis category. Group I, normal responses;
 543 Group II, attenuation of cone responses; Group III, attenuation of cone and rod
 544 responses.



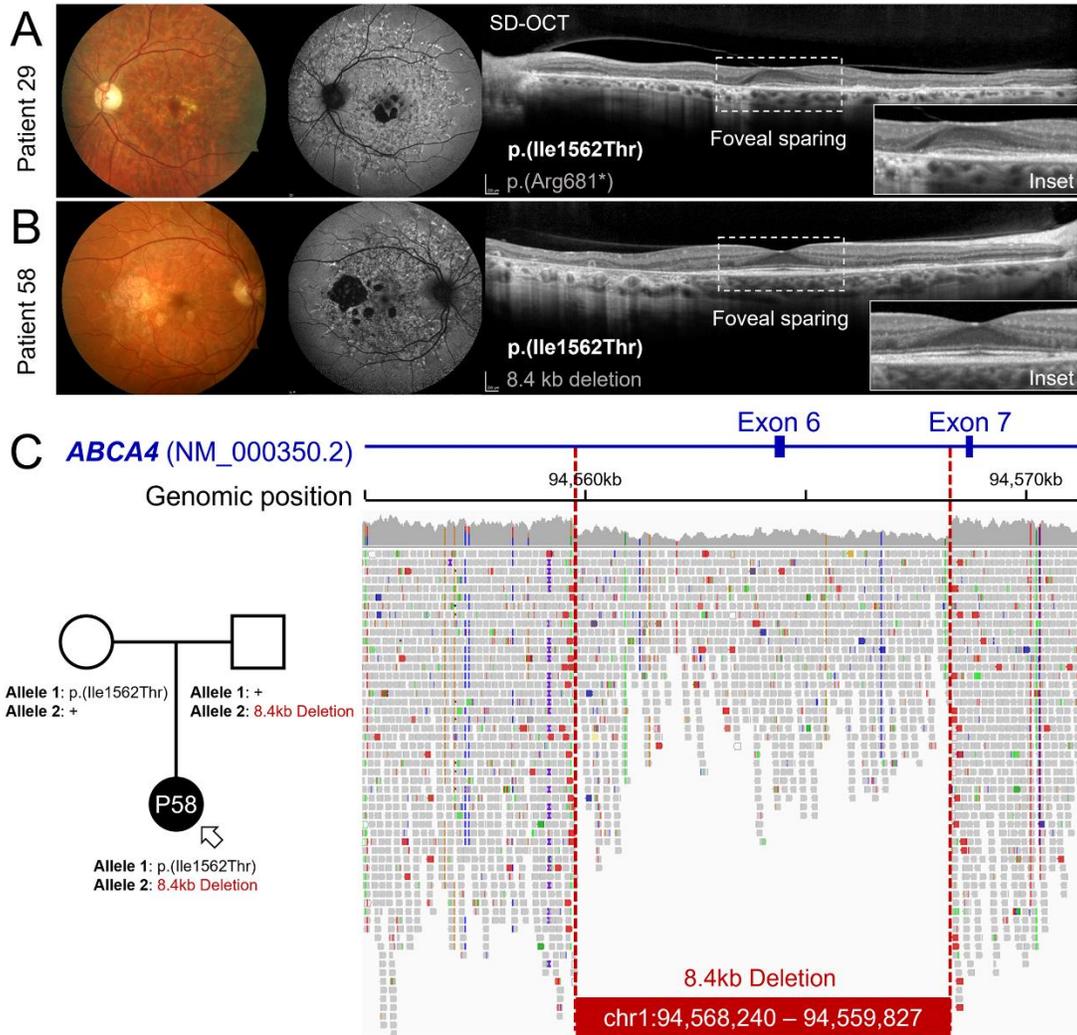
545

546 **Figure 2:** Classification and phenotypic characterization of mild *ABCA4* alleles. (A) Mild
 547 *ABCA4* alleles identified in patients with mild prognoses included p.(Gly1961Glu) and two
 548 hypomorphic allele sub-groups: frequent hypomorphs which consisted of p.(Asn1868Ile)
 549 and rare hypomorphs which consisted of p.(A1038V), c.4253+43G>A, p.(Pro1486Leu),
 550 p.(Thr1526Met), p.(Ile1562Thr), p.(Arg2030Gln). (B) Horizontal spectral domain-optical
 551 coherence tomography (SD-OCT) scan showing structural preservation of the fovea in
 552 Patient 22, an allele-specific sub-phenotype common amongst p.(Asn1868Ile) genotypes.
 553 (C) Scatter plot of average age of onset (years) versus average best-corrected visual
 554 acuity (BCVA) of the least deteriorated eye in patients within all patients/genotypes with
 555 p.(Gly1961Glu) (blue), frequent hypomorph (p.(Asn1868Ile) (green), rare hypomorph
 556 (green) and all other allele combinations (black) in the study cohort. Horizontal and
 557 vertical bars represent +/-95% confidence intervals. BCVA are provided as logMAR units
 558 with corresponding Snellen equivalents listed above the axis. (D) Survival analysis
 559 showing the probability of the least affected eye retaining better than Snellen 20/400 in
 560 patients with p.(Gly1961Glu) (blue curve), rare and frequent hypomorphic alleles (green
 561 curve) and all other patients (black curves). Color-matched dotted lines represent 95%
 562 confidence intervals for each individual curve.



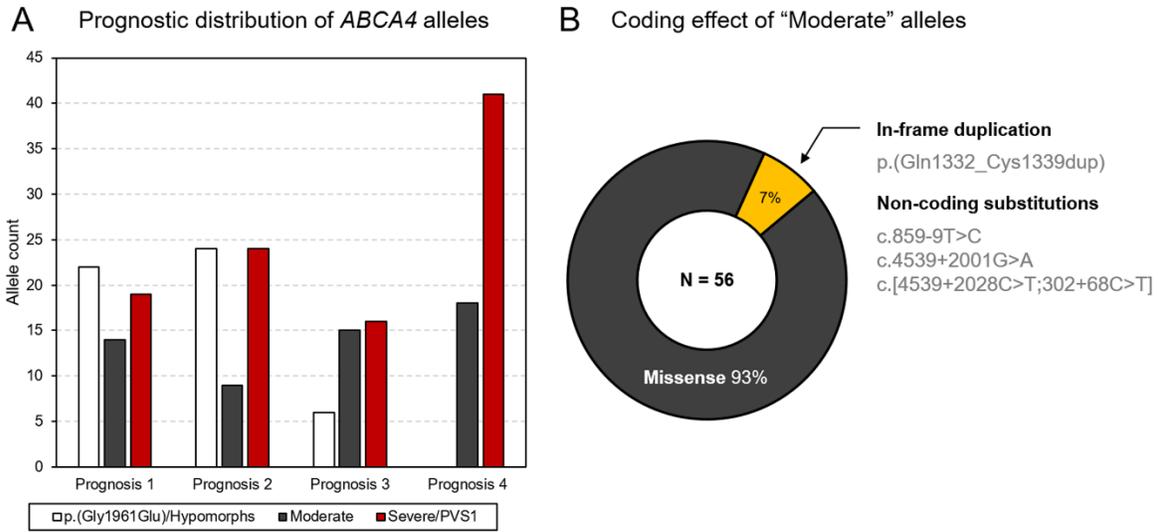
563

564 **Figure 3:** Retinal phenotype of the rare hypomorph p.(Arg2030Gln) variant of ABCA4
 565 disease. Macular 30° autofluorescence, 55° autofluorescence images and horizontal
 566 spectral domain-optical coherence tomography (SD-OCT) scans of the (A) left eye of
 567 Patient 42 and (B) left eye of Patient 40. SD-OCT scans with enlarged insets of the fovea
 568 show preservation of outer retinal layers resulting in 20/20 vision in the eyes of both
 569 patients. Unimpaired full-field scotopic (dark-adapted 0.01 rod), maximal (dark-adapted
 570 3.0 combined rod and cone), 30 Hz flicker and photopic (light-adapted 3.0 single flash
 571 cone) electroretinogram responses of the right and left eyes of Patient 39 and
 572 representative waveforms from an age-matched healthy control eye.



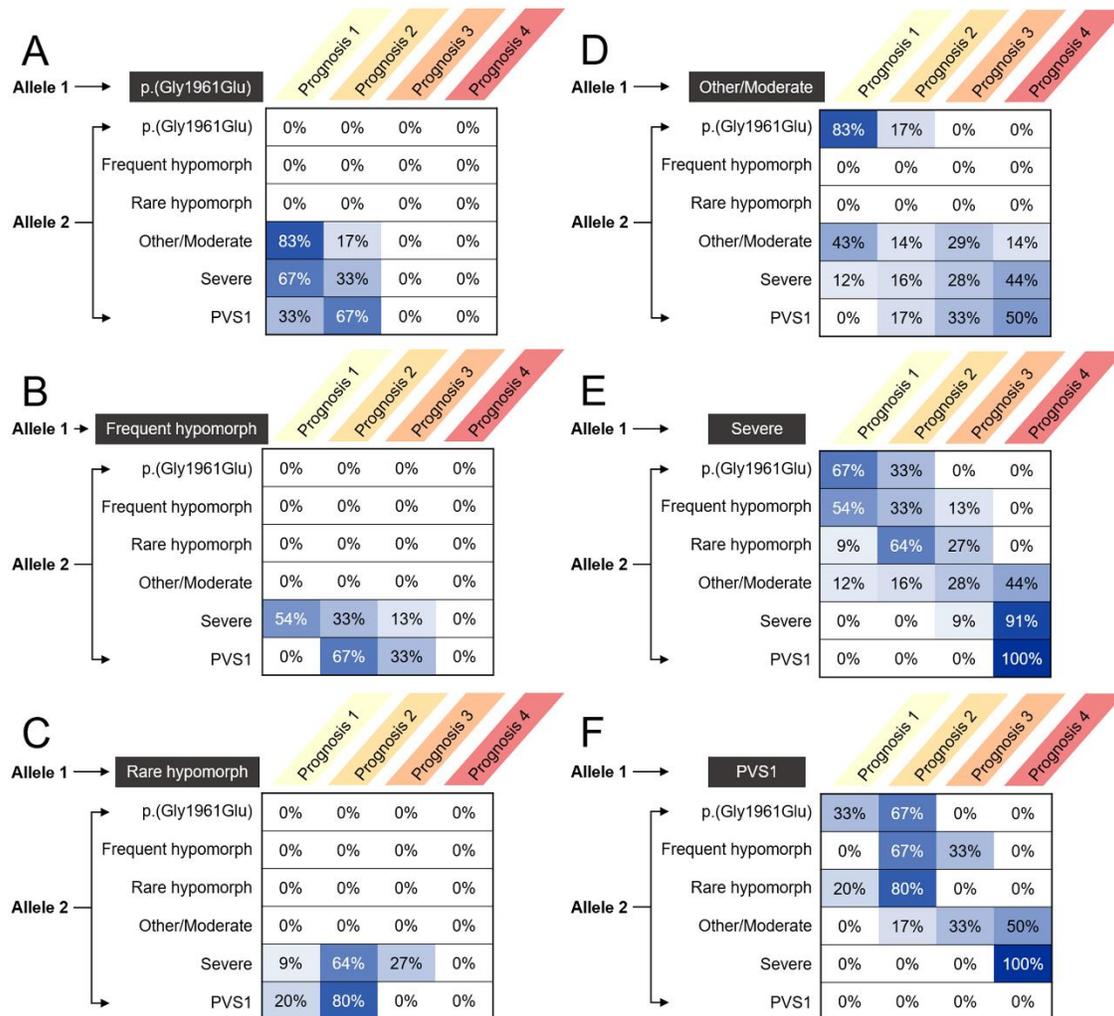
573

574 **Figure 4:** Retinal phenotype of the rare hypomorph p.(Ile1562Thr) variant of ABCA4
 575 disease. Color fundus photographs, autofluorescence images and horizontal spectral
 576 domain-optical coherence tomography (SD-OCT) of the (A) left eye of Patient 29 and (B)
 577 right eye of Patient 58. SD-OCT scans with enlarged insets of the fovea show
 578 preservation of outer retinal resulting in unimpaired 20/20 vision in the eyes of both
 579 patients. (C) Pedigree showing segregation of the p.(Ile1562Thr) and large 8.4 kb deletion
 580 alleles in Patient 58. Pileup of whole genome sequencing reads showing the approximate
 581 size and genomic position of the *ABCA4* deletion which spans the entire length of exon
 582 6.



583

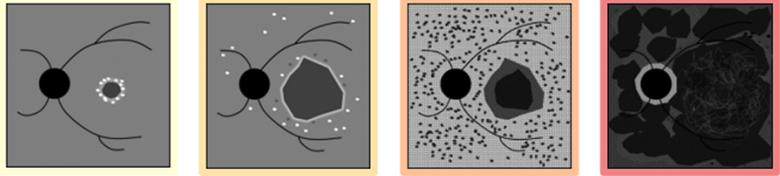
584 **Figure 5:** Clinical and genetic characteristics of moderate ABCA4 alleles in the study. (A)
 585 Distribution of p.(Gly1961Glu) and hypomorphs (white bars), moderate (gray bars) and
 586 Severe/PVS1 (red bars) alleles across Prognosis categories. (B) Coding effect of alleles
 587 designated as "Moderate" in patients with ABCA4 disease.



588

589 **Figure 6:** Prognostic probabilities (%) of all possible combinations for each allele class:
 590 (A) p.(Gly1961Glu), (B) Frequent hypomorph, (C) Rare hypomorph, (D) Moderate, (E)
 591 Severe and (F) PVS1. Percentages represent the observed fraction of patients across
 592 each Prognosis category for a given Allele 1 and Allele 2 combination.

ABCA4 Genotype-Phenotype
Correlation Matrix



Allele 1	Allele 2	Prognosis 1	Prognosis 2	Prognosis 3	Prognosis 4
p.(Gly1961Glu)	Moderate	83%	17%	0%	0%
	Severe	67%	33%	0%	0%
	PVS1	33%	67%	0%	0%
Frequent hypomorph	Severe	54%	33%	13%	0%
	PVS1	0%	67%	33%	0%
Rare hypomorph	PVS1	20%	80%	0%	0%
	Severe	9%	64%	27%	0%
Moderate	Moderate	43%	14%	29%	14%
	Severe	12%	16%	28%	44%
	PVS1	0%	17%	33%	50%
Severe	Severe	0%	0%	9%	91%
	PVS1	0%	0%	0%	100%

593

594 **Figure 7:** Genotype-phenotype correlation matrix based on the long-term prognostic
 595 outcomes of 112 genetically confirmed patients with ABCA4 disease patients.
 596 Percentages represent the observed fraction of patients across each Prognosis category
 597 for a given Allele 1 and Allele 2 combination. For the list of unclassified variants, see Table
 598 1 or Supplementary Table 2.

599

600 **Table 1:** Classification criteria and list of all pathogenic ABCA4 alleles in the study cohort

Allele class	Classification criteria	Alleles
p.(Gly1961Glu)	(Allele-specific)	p.(Gly1961Glu)
Frequent hypomorph	(Allele-specific)	p.(Asn1868Ile)
Rare hypomorph	Mild prognostic association and hypomorphic allele features (late-onset symptoms, foveal sparing)	p.(Ala1038Val), c.4253+43G>A, p.(Pro1486Leu), p.(Thr1526Met), p.(Ile1562Thr), p.(Arg2030Gln)
Moderate	No intrinsic indication of severity (variant effect on protein); determined by functional studies to be moderate; undetermined clinical association	p.(Arg24His), p.(Ile214Asn), p.(Leu257Arg), c.859-9T>C, p.(Trp339Gly), p.(Lys346Thr), p.(Arg508Cys), p.(Arg602Gln), p.(Gln623Arg), p.(Arg653His), p.(Phe655Cys), p.(Val675Ile), p.(Leu844Arg), p.(Thr972Asn), p.(Val989Ala) , p.(Ser1096Leu), p.(Arg1108Cys) , p.(Arg1108His), p.(Thr1112Asn), p.(Thr1117Ile), p.(Gln1332_Cys1339dup), p.(Arg1640Gln), p.(Ser1696Asn), p.(Leu1784Arg), p.(Met1882Ile), p.(Val1896Asp), p.(Arg2040Gln), p.(Arg2077Gly), p.(Arg2106Cys), p.(Arg2107His) , p.(Cys2150Tyr), p.[Cys1490Tyr;Asn1868Ile], p.([Leu257Aspfs*3,Gly1961Glu]), p.([Thr1253Met;Gly1961Glu]), c.4539+2001G>A, c.[4539+2028C>T;302+68C>T]
Severe	Observed in <i>trans</i> to hypomorphic allele in patients	p.(Arg18Trp), p.(Cys54Tyr) , p.(Gly88Arg), p.(Arg602Trp), p.(Cys641Arg), p.(Arg653Cys), c.2382+179G>A, p.(Ala848Asp), p.(Asn965Ser) , p.(Gly1203Arg), p.(Pro1380Leu) , ‡ p.Phe1417del, p.(Tyr1557Cys), p.(Arg1705Trp), p.(Phe1714Ser), p.(Leu2027Phe) , p.(Arg2077Trp), p.(Glu2096Lys), p.[Leu541Pro;Ala1038Val] , p.[Trp1408Arg;Arg1640Trp]
	Validated in functional studies in HEK293T and patient-derived cell lines	c.768G>T, † c.1100-6T>A, c.3050+5G>A, c.4253+5G>T, p.(Gln1513Arg), c.4773+3A>G, c.5196+1056A>G, c.5461-10T>C, c.6342G>A†
	Associated with Prognosis 3 and Prognosis 4	p.(Thr1019Met), ‡ p.(Ala1598Asp), ‡ p.[Gly863Ala,Gly863del;Asn1868Ile], p.[Asp1532Asn;Asn1868Ile], ‡ c.5714+5G>A
PVS1	Null or loss-of-function variants (nonsense, frameshifts, canonical +/- 1 or 2 splice sites, large multi-exonic deletions)	p.(Asn14Lysfs*38), p.(Ser84Thrfs*16), p.(Asp202Thrfs*39), c.570+331_768+4523del, p.(Leu296Cysfs*4), p.(Val521Serfs*47), p.(Trp663*), p.(Arg681*) , c.2160+1G>C, c.2587+2T>C, p.(Ser1071Cysfs*14), p.(Trp1408*), p.(Cys1502*), c.4539+1G>T, c.4540-2A>G, p.(Leu1534Trpfs*1), c.5018+2T>C, p.(Ile1687Phefs*15), p.(Val1706*), p.(Val1764Trpfs*14), p.(Arg2030*) , p.(Ala2044Valfs*25), p.(Arg2149*), p.(Gln2220*), c.6148-698_6670del

601
602 *Footnotes:* Variants in bold were found in 2 or more patients in the study. †Synonymous variants validated
603 by Braun et al. (2013) and Sangermano et al. (2018) to have the following effects: c.768G>T
604 (p.(Leu257Valfs*17)) and c.6342G>A (p.([Val2114_Ser2129delfs*5,=])). ‡Homozygous in individual with
605 Prognosis 3 or Prognosis 4. The PVS1 classification was used in accordance with the ACMG/AMP
606 Standards and Guidelines for the Interpretation of Sequence Variants.(37)