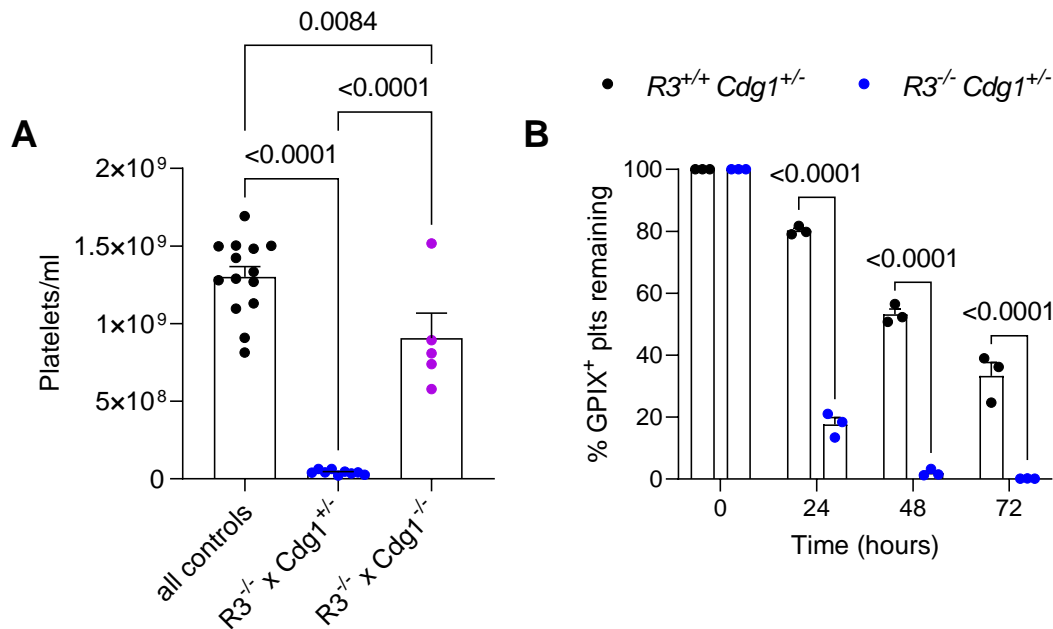
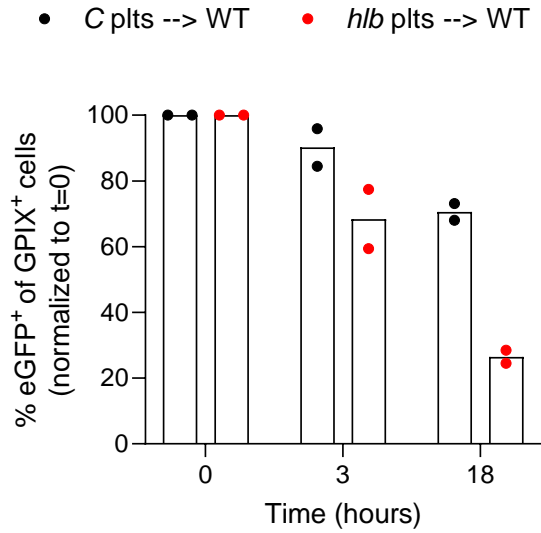


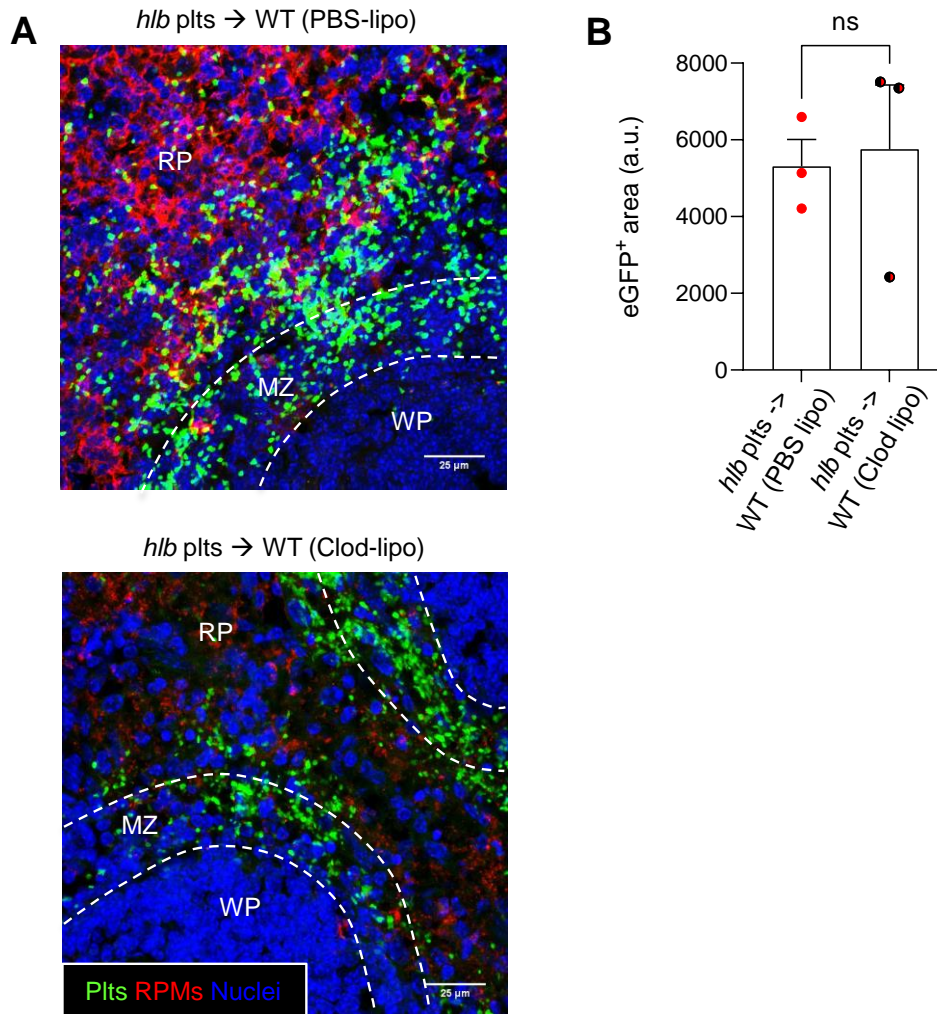
## Supplemental Figures



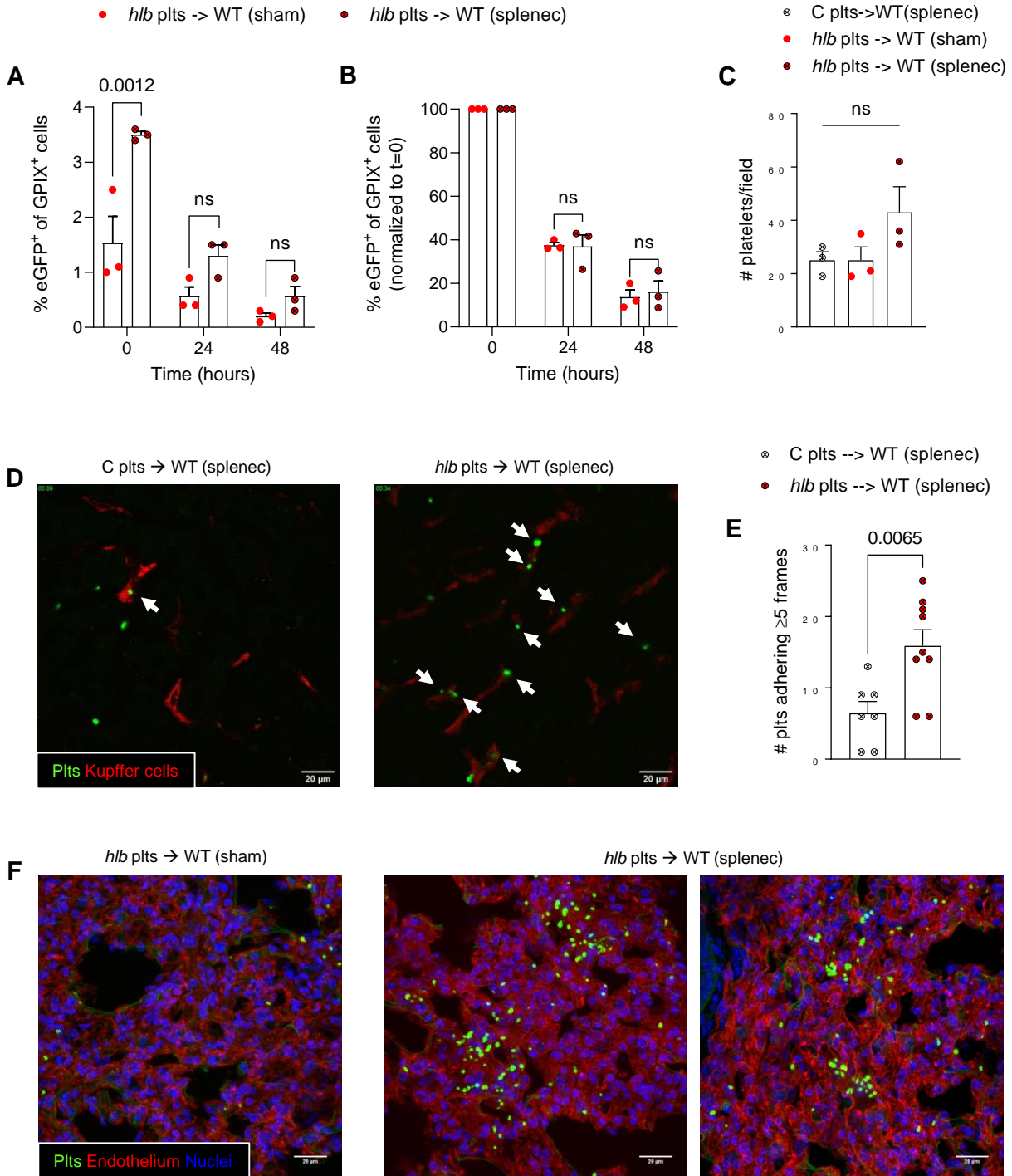
**Supplemental Figure 1: Severely reduced platelet counts and platelet lifespan in  $R3^{-/-} \times Cdg1^{+/-}$  mice.** (A) Circulating platelet counts were determined by flow cytometry using whole blood from  $R3^{-/-} \times Cdg1^{+/-}$ ,  $R3^{-/-} \times Cdg1^{-/-}$  and mixed control mice (n=5-14). (B) Mice were injected I.V. with anti-GPIX-AF488 antibody (2.5  $\mu$ g/mouse) on day 0 to label all circulating platelets, and then the percentage of GPIX-AF488<sup>+</sup> platelets remaining was determined every 24 hrs by flow cytometry (n=3). Data shown as mean  $\pm$  SEM. Statistical significance was determined using one-way (A) or two-way (B) ANOVA with Bonferroni correction for multiple comparisons.



**Supplemental Figure 2: Rapid clearance of transfused eGFP<sup>+</sup> *R3<sup>h1b/h1b</sup>* x *Cdg1<sup>+/-</sup>* platelets in WT recipient mice.** Platelets from eGFP<sup>+</sup> *R3<sup>+/+</sup>* x *Cdg1<sup>+/-</sup>* (C) or eGFP<sup>+</sup> *R3<sup>h1b/h1b</sup>* x *Cdg1<sup>+/-</sup>* (*h1b*) mice were transfused into recipient WT mice (n=2). The percentage of eGFP<sup>+</sup> platelets of all GPIIX<sup>+</sup> platelets was determined at 20 mins (t=0), 3 hrs and 18 hrs after transfusion, and normalized to t=0.

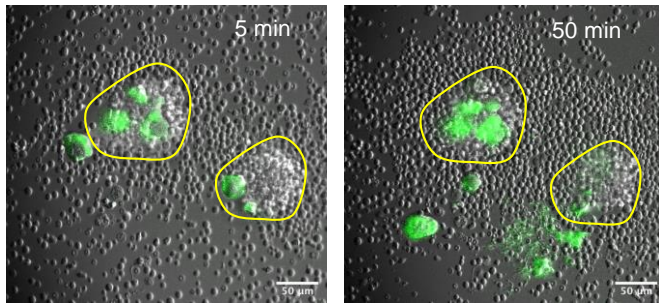
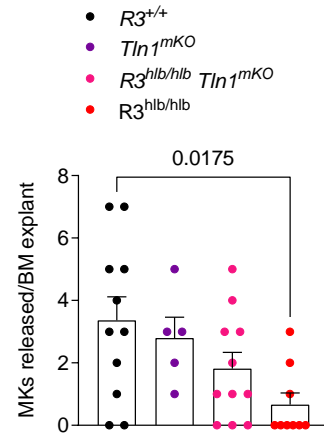


**Supplemental Figure 3: Macrophage depletion does not impair splenic sequestration of *Rasa3<sup>h1b/h1b</sup>* platelets.** (A) IF images of spleen cryosections from WT mice injected with either PBS (PBS-lipo) or Clodronate-containing (Clod-lipo) liposomes, and subsequently transfused with eGFP<sup>+</sup> *R3<sup>h1b/h1b</sup>* × *Cdg1<sup>+/-</sup>* (*h1b*) platelets (24 hrs post-liposomes). Cryosections were stained for RPMs with anti-F4/80-AF647 (red) and nuclei with DAPI (blue), and platelets were visualized by eGFP intensity (green). Scale bars equal 25 μm. (B) Platelet (eGFP<sup>+</sup>) area was determined in spleen cryosections using Fiji, and expressed as area in arbitrary units (a.u.) (n=3). Data shown as mean ± SEM. Statistical significance was determined using Student's t-test, ns=not significantly different.

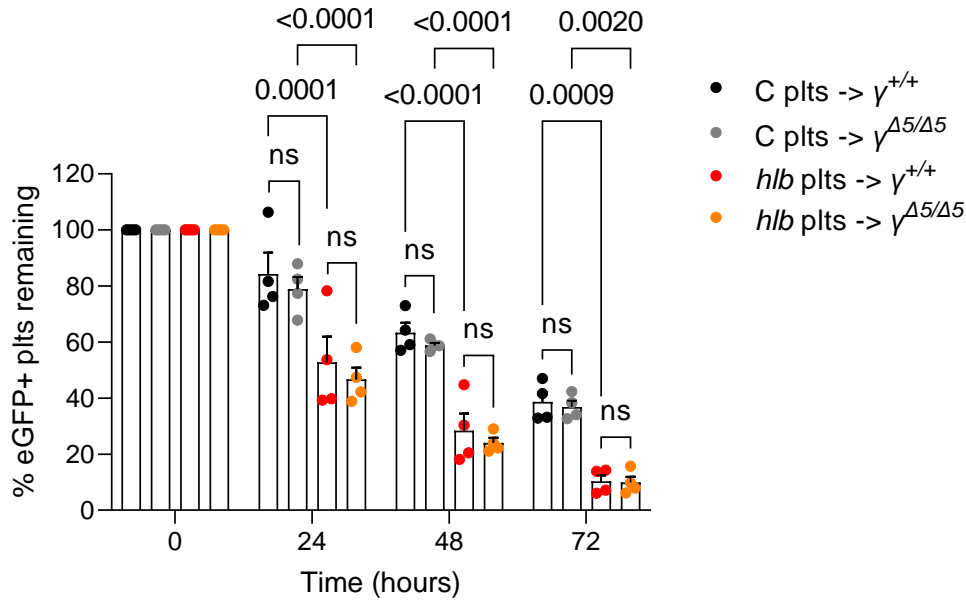


**Supplemental Figure 4: Secondary liver and lung clearance of *Rasa3<sup>h1b/h1b</sup>* platelets compensates in the absence of the spleen. (A)** Recovery and survival of transfused eGFP<sup>+</sup> *h1b* platelets in splenectomized (splenec) or sham-operated (sham) WT recipient mice at 20 mins (t=0), 24 hrs and 48 hrs post-transfusion (n=3). **(B)** Data from (a) normalized to t=0 for each genotype. **(C)** Platelets were quantified in immunostained liver cryosections from WT (splenec) or WT (sham) mice transfused with C or *h1b* platelets

(n=3 mice). **(D)** Still frames from time-lapse confocal videos acquired in the liver of WT (splenec) recipients transfused with eGFP<sup>+</sup> C or *h/b* platelets 18-24 hrs post platelet transfusion. Platelets were visualized by eGFP fluorescence (green), and Kupffer cells were labeled *in vivo* by I.V. administration of anti-F4/80-AF647 antibody (2.5 µg/mouse; red). Scale bars equal 20 µm. Arrowheads denote adherent platelets in contact with Kupffer cells within liver sinusoids. **(E)** Quantification of platelet adhesion in liver sinusoids from intravital confocal videos (3-5 different 5-min videos were analyzed for n=2 mice per group). **(F)** Representative images of lung cryosections from WT (sham) or WT (splenec) mice transfused with *h/b* platelets. Platelets are visualized by eGFP fluorescence (green), endothelium was stained with anti-CD31-AF647 antibody (red), and nuclei with DAPI (blue). Scale bars equal 20 µm. Data shown as mean ± SEM. Statistical significance was determined using two-way (A,B) or one-way (C) ANOVA with Bonferroni correction for multiple comparisons, or Student's t-test (E), ns=not significantly different.

**A***R3<sup>h1b/h1b</sup> Tin1<sup>mKO</sup>***B**

**Supplemental Figure 5: Genetic deletion of Talin1 partially rescues impaired ex vivo proplatelet formation in *Rasa3<sup>h1b/h1b</sup>* MKs.** (A) Representative images of ex vivo proplatelet-forming MKs from *R3<sup>h1b/h1b</sup> x Tin1<sup>mKO</sup>* mice taken at the indicated time points after start of imaging. Samples were stained with anti-GPIX-AF488 to label MKs/platelets (green). Scale bars equal 50  $\mu$ m. Yellow lines delimit BM pieces. (B) MKs released from BM explants, expressed as number of MKs released per explant in each field of view (n=5-11). Data shown as mean  $\pm$  SEM. Statistical significance was determined using one-way ANOVA with Bonferroni correction for multiple comparisons.



**Supplemental Figure 6: Transfused *hlb* platelets are cleared equally fast in fibrinogen  $\gamma^{\Delta5/\Delta5}$  mice.** Platelets from eGFP<sup>+</sup> *R3*<sup>+/+</sup>  $\times$  *Cdg1*<sup>+/-</sup> (C) or eGFP<sup>+</sup> *R3*<sup>hlb/hlb</sup>  $\times$  *Cdg1*<sup>+/-</sup> (*hlb*) mice were transfused into recipient fibrinogen  $\gamma^{\Delta5/\Delta5}$  or littermate control mice ( $n=4$ ). The percentage of eGFP<sup>+</sup> platelets of all GPIX<sup>+</sup> platelets was determined at 20 mins (t=0) and every 24 hrs thereafter, and normalized to t=0. Data shown as mean  $\pm$  SEM. Statistical significance was determined using two-way ANOVA with Bonferroni correction for multiple comparisons, ns=not significantly different.

## Supplemental Videos

Supplemental Video 1: 3D reconstruction of confocal Z stack from  $R3^{+/+}$  BM explant. MKs and platelets are labeled in green, endothelium in red, nuclei in blue.

Supplemental Video 2: 3D reconstruction of confocal Z stack from  $R3^{hlb/hlb}$  BM explant. MKs and platelets are labeled in green, endothelium in red, nuclei in blue.

Supplemental Video 3: Time lapse imaging of ex vivo proplatelet formation in BM explants from a  $R3^{+/+}$  mouse. MKs are labeled in green, overlaid with brightfield.

Supplemental Video 4: Time lapse imaging of ex vivo proplatelet formation in BM explants from a  $R3^{hlb/hlb}$  mouse. MKs are labeled in green, overlaid with brightfield.

Supplemental Video 5: Time lapse spleen IVM using 2-photon microscopy in a WT mouse transfused with eGFP<sup>+</sup>  $R3^{+/+}$  x  $Cdg1^{+/-}$  ("C") platelets. Platelets are green, red pulp macrophages are labeled in red.

Supplemental Video 6: Time lapse spleen IVM using 2-photon microscopy in a WT mouse transfused with eGFP<sup>+</sup>  $R3^{+/+}$  x  $Cdg1^{+/-}$  ("*hlb*") platelets. Platelets are green, red pulp macrophages are labeled in red.

Supplemental Video 7: Time lapse liver IVM using laser scanning confocal in a WT mouse transfused with eGFP<sup>+</sup>  $R3^{+/+}$  x  $Cdg1^{+/-}$  ("C") platelets. Platelets are green, Kupffer cells are labeled in red.

Supplemental Video 8: Time lapse liver IVM using laser scanning confocal in a WT mouse transfused with eGFP<sup>+</sup>  $R3^{+/+}$  x  $Cdg1^{+/-}$  ("*hlb*") platelets. Platelets are green, Kupffer cells are labeled in red.

Supplemental Video 9: 3D reconstruction of confocal Z stack from  $R3^{hlb/hlb}$  x  $Tln1^{mKO}$  BM explant. MKs and platelets are labeled in green, endothelium in red, nuclei in blue.