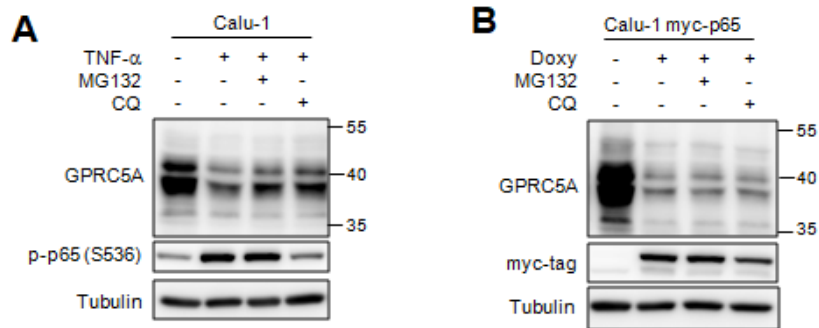


Supplementary Information of

**NFκB Induces Epigenetic Repression of GPRC5A in Lung Epithelial Cells to
Promote Neoplasia**

Supplementary Data



Supplementary Figure 1 Related to Figure 4

Proteasome and autophagy inhibitors do not affect the repression of GPRC5A by TNF α and p65. **A**, Calu-1 cells were treated with TNF α (10 ng/ml) for 3 days and MG132 (10 μ M) or chloroquine (CQ) (100 μ M) were added to the medium for the last 6 hours, cells were lysed by RIPA. GPRC5A protein levels were analyzed by western blotting. **B**, Calu-1 cells expressed myc-p65 under the control of doxycycline (doxy)-inducible promoter (TRE2) were treated with doxycycline (100 ng/ml) to induce p65 expression for 48h. MG132 (10 μ M) or chloroquine (100 μ M) were added to the medium for the last 6 hours, cells were lysed by RIPA and GPRC5A protein levels were analyzed by western blotting.

A

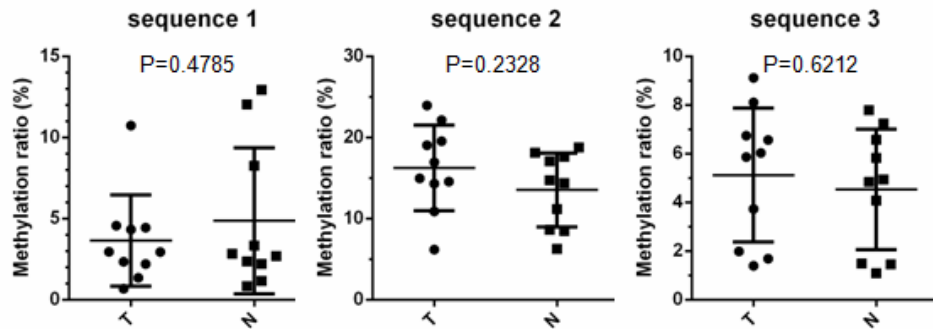
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CCGTAAGGAGCTGGTCTGCTCCGGCTCCCTCTCGTAGCCGGCGCCCTGGGACCAGCGCGTGAGCACGTCT
CGGAGGAGTCCGATGCGC TGGGCAGGGCCGGGTGGTCACCCCGCCCAGCTTCCAGGGG

CpG island 2

GAATAGGCGTGTCCTCTCCCTCGACCCCTCCCCCTCCTTGTCCCTCTGCTCACCCCTCGCTCGTTCCCTCCC
TCCGGCGAGGGCCGCCTTTATAACAACCTGCTCAGAGTGCGAGGGCGGGATAGCTGTCCAAGGTCTCCCCCA
GCACTGAGGAGCTCGCCTGCTGCCCTCTTGCGCGCGGGAAGCAGCACCAAGTTCACGGCCAACGCCTTGGC
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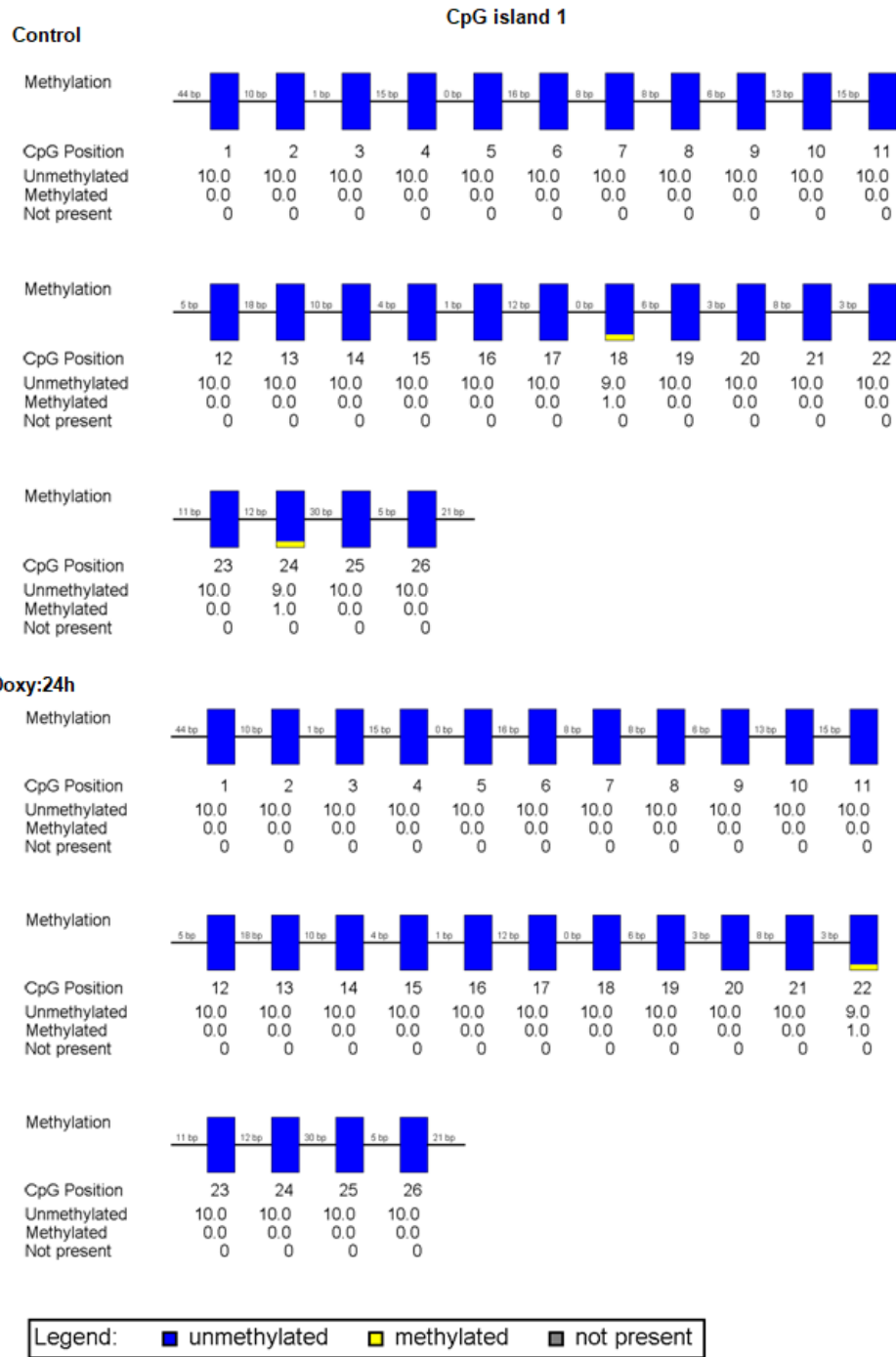
B



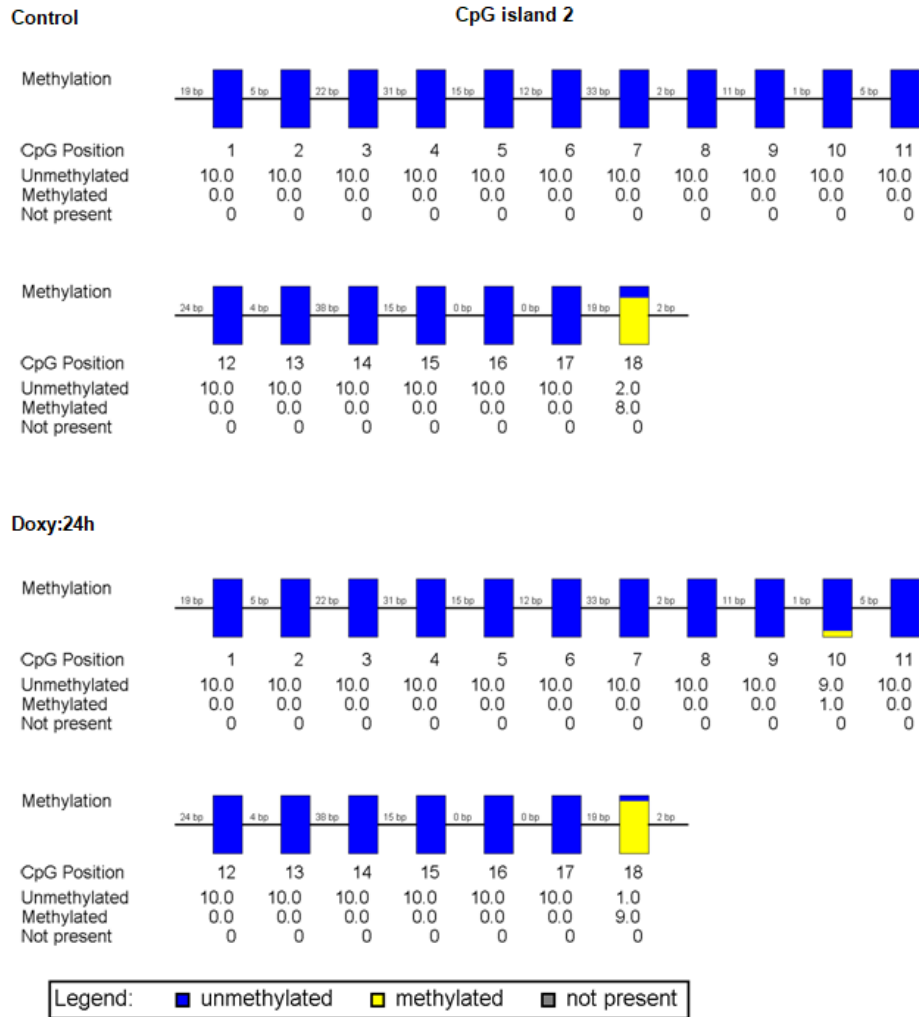
Supplementary Figure 2 Related to Figure 7

DNA methylation at the GPRC5A promoter shows no difference between lung cancer and adjacent normal tissues. **A**, three sequences (red color) in 2 CpG islands were chosen to perform DNA methylation sequencing. Sequence 1 and 2 were located in CpG island 1, includes 6 CpG sites respectively. Sequence 3 was located in CpG island 2, includes 5 CpG sites. All three sequences were indicated with red color. **B**, DNA were extracted from ten lung cancer and corresponding adjacent normal tissues, DNA methylation levels on these three sequences were analyzed by bisulfite PCR; T, tumor; N, normal.

A

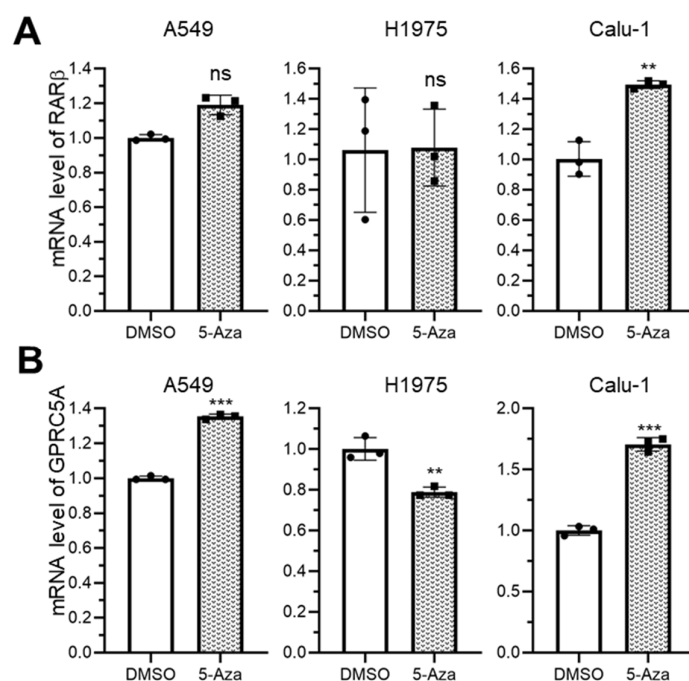


B



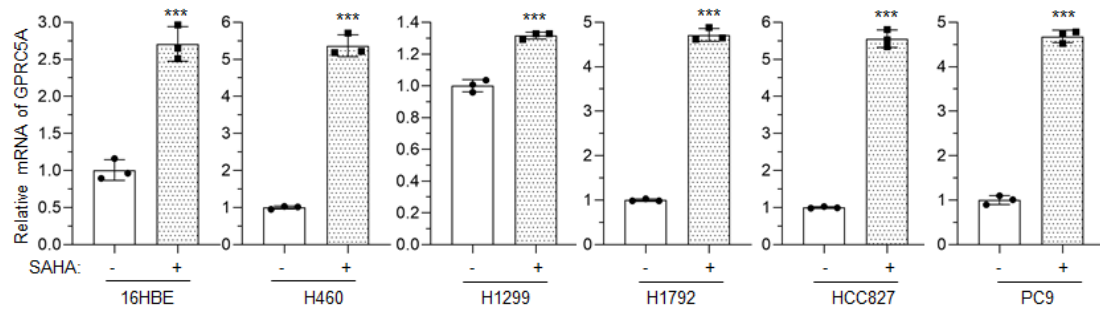
Supplementary Figure 3 Related to Figure 7

Overexpression of p65 in Calu-1 cells do not change the DNA methylation status at the GPRC5A promoter. Calu-1 cells expressed myc-p65 under the control of doxycycline-inducible promoter (TRE2) were treated with DMSO as negative control or doxycycline (300 ng/ml) to induce p65 expression for 24 h. The methylation status of the CpG island 1 (A) and CpG island 2 (B) at the GPRC5A promoter were analyzed by bisulfite sequencing PCR.



Supplementary Figure 4 Related to Figure 7

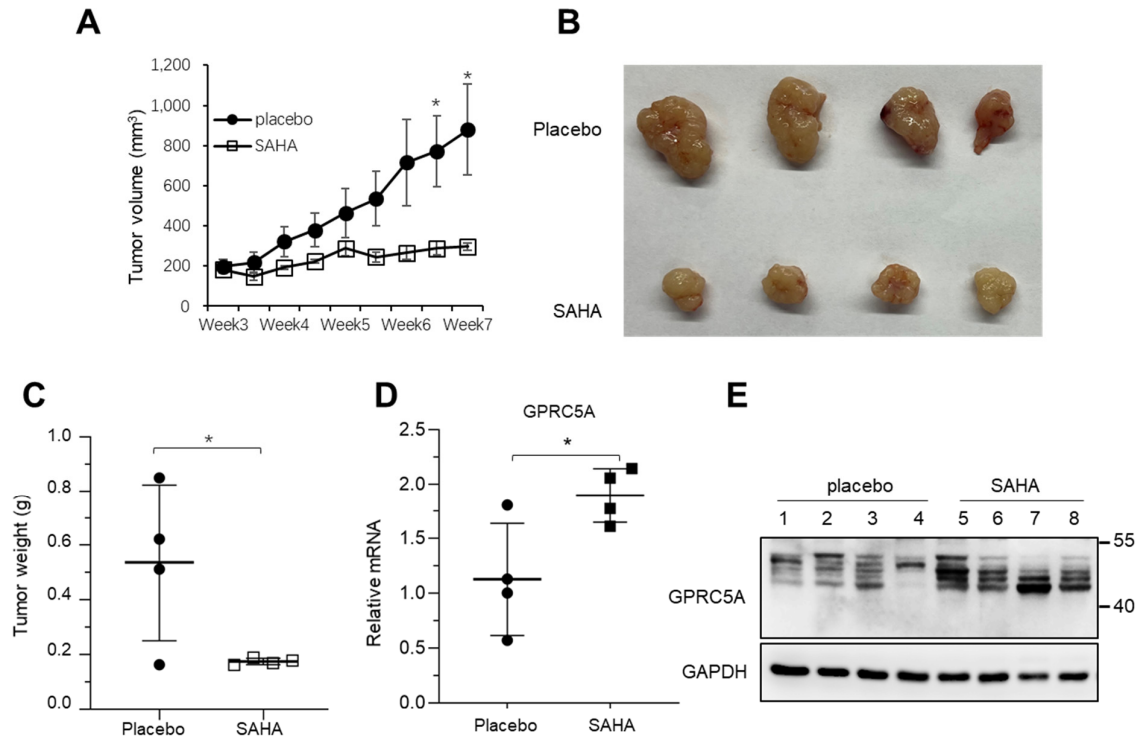
5-Aza-dc treatment has limited effects on the levels of RAR β and GPRC5A mRNA in three NSCLC cells. A549, H1975 and Calu-1 cells were treated with or without 5-Aza-dc (1 μ M, 4 days). RAR β and GPRC5A mRNA levels were analyzed by quantitative PCR. * p <0.05; ** p <0.01; *** p <0.001.



Supplementary Figure 5 Related to Figure 7

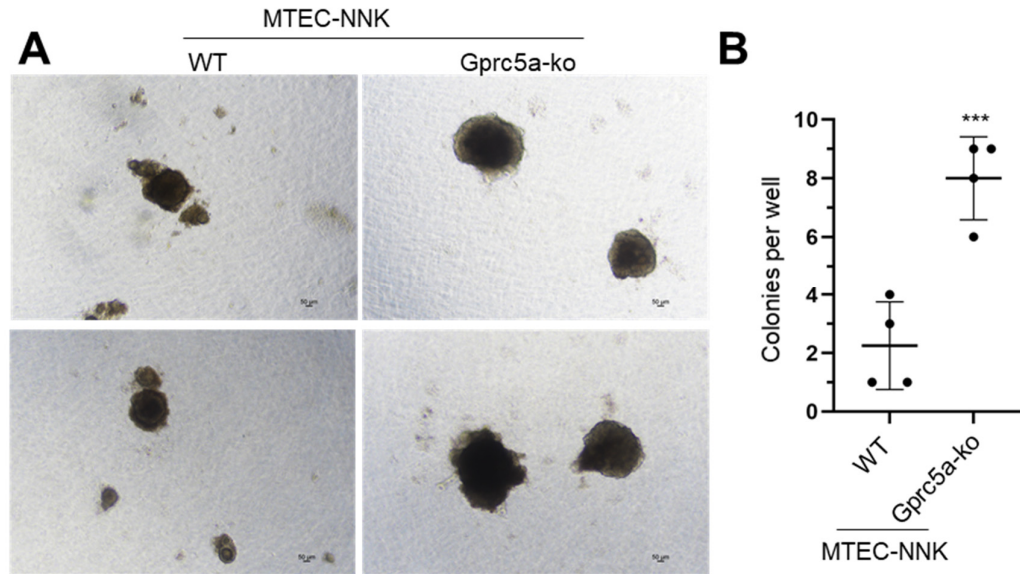
SAHA treatment increase GPRC5A mRNA levels. Normal human bronchial epithelial cell line (16HBE) and multiple human NSCLC cell lines were treated with or without SAHA (2.5 μ M, 24 hours), GPRC5A mRNA levels were analyzed by quantitative PCR.

* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.



Supplementary Figure 6

SAHA treatment increases GPRC5A expression and inhibits tumor formation in vivo. A549 cells were injected subcutaneously into nude mice; mice were separated to two groups randomly. After three weeks, once group was treated with placebo and the other group was treated with SAHA. Tumor volumes were measured twice a week. After additional four weeks, mice were sacrificed. Tumor weights were measured, and GPRC5A mRNA and protein expression levels were analyzed by qPCR and western blot. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.



Supplementary Figure 7

NNK-treated Gprc5a-ko MTEC cells have stronger colony-formation capability than NNK-treated wild type METC cells in soft-agar assay. Wild type (WT) and Gprc5a-ko (KO) MTEC cells were repeatedly treated with NNK (100 pM) for 10 passages, leading to MTEC-KO-NNK10 and MTEC-WT-NNK10 cells. Five hundred cells per well (12 well plate) were seeded in soft agar with quadruplicates. After 3 weeks, the numbers and sizes of colonies were measured under microscope. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

Supplementary Table 1. The primer sequence used in quantity real time PCR

Gene	Forward primer	Reverse primer
mGprc5a	ACCACAGACTTTGTGACCTGG	CGAGTGCAAACATGCAAGCC
mAtcb	GGCTGTATTCCCCTCCATCG	CCAGTTGGTAACAATGCCATGT
hGPRC5A	CTCACTCTCCCGATCCTCGT	CAGTCCGATGATGAAGGCGAA
hRelA	ATGTGGAGATCATTGAGCAGC	CCTGGTCCTGTGTAGCCATT
hclAP2	TTTCCGTGGCTCTTATTCAAAC	GCACAGTGGTAGGAACCTTCTCAT
hRAR β	TCCGAAAAGCTCACCAGGAAA	GGCCAGTTCACCTGAATTTGTCC
hGAPDH	TGCACCACCAACTGCTTAGC	GGCATGGACTGTGGTCATGAG

Supplementary Table 2. The primer sequence used in ChIP assay

Gene	Forward primer	Reverse primer
hGPRC5A	TGGAAGCTGGAATAGGCGTGT	GTGAAGCTTGGTGCTGCTTCC
hGAPDH	TACTAGCGGTTTTACGGGCG	TCGAACAGGAGGAGCAGAGAGCGA
hIKB α	GACGACCCCAATTCAAATCG	TCAGGCTCGGGGAATTTCC

Uncropped/unedited gel document

Song et al (Submission to *JCI Insight*, August 2022)

NF- κ B Represses Retinoic Acid Receptor-Mediated Transactivation of GPRC5A in Lung Epithelial Cells for Neoplasia

Note: This project was started from 2007 in Dr. Reuben Lotan's laboratory at the University of Texas MD Anderson Cancer Center. It was carried out by Dr. Xiaofeng Ye (co-first author of this manuscript) as part of his PhD thesis under the direct supervision of Dr. Jiong Deng (research assistant professor at that time). Both of them were working on the GPRC5A project at Dr. Lotan's group (please see reference below). Unfortunately, Dr. Lotan (1946-2011) passed away due to a terrible car accident in 2011, and his laboratory was closed 2-month later after this tragic accident. Dr. Ye graduated by summarizing his study/data on GPRC5A, including this uncompleted project, in his PhD thesis; he then performed a short period of post-doc study at Dr. Binhua P. Zhou's laboratory working on cancer metastasis. Dr. Deng went back to Shanghai Jiao-Tong University School of Medicine (2010-2021) to continue the GPCR5A study, including this project in his laboratory. The first author (Dr. Hongyong Song) is a post-doctoral fellow at Dr. Deng's group and he is the major driving force to finally complete this project. Due to the unexpected tragic accident of Dr. Lotan and the short notice of laboratory closure, Dr. Ye was unable to retrieve the original uncropped version of blots/films for **Fig 2D, 3A, 3B, 5B, 6C and 6D** despite many efforts.

To resolve these issues, we have repeated these experiments (for Fig 2D, 3A, 3B, 5B, 6C, and 6D) in the last three months. We have obtained similar results. Now we have presented these uncropped/unedited images in this uncropped/unedited blot documents.

Reference:

Xiaofeng Ye, Qingguo Tao, Yafan Wang, Yijun Cheng, **Reuben Lotan**. Mechanisms underlying the induction of the putative human tumor suppressor GPRC5A by retinoic acid. *Cancer Biol Ther*. **2009** May;8(10):951-62.

Qingguo Tao, Junya Fujimoto, Taoyan Men, **Xiaofeng Ye**, **Jiong Deng**, Ludovic Lacroix, John L Clifford, Li Mao, Carolyn S Van Pelt, J Jack Lee, Dafna Lotan, **Reuben Lotan**. Identification of the retinoic acid-inducible Gprc5a as a new lung tumor suppressor gene. *J Natl Cancer Inst*. **2007** Nov 21;99(22):1668-82.

Jiong Deng, Junya Fujimoto, **Xiao-Feng Ye**, Tao-Yan Men, Carolyn S Van Pelt, Yu-Long Chen, Xiao-Feng Lin, Humam Kadara, Qingguo Tao, Dafna Lotan, **Reuben Lotan**. Knockout of the tumor suppressor gene Gprc5a in mice leads to NF-kappaB activation in airway epithelium and promotes lung inflammation and tumorigenesis. *Cancer Prev Res (Phila)*. **2010** Apr;3(4):424-37.

Small Airway Epithelial Cells

Fig 2A

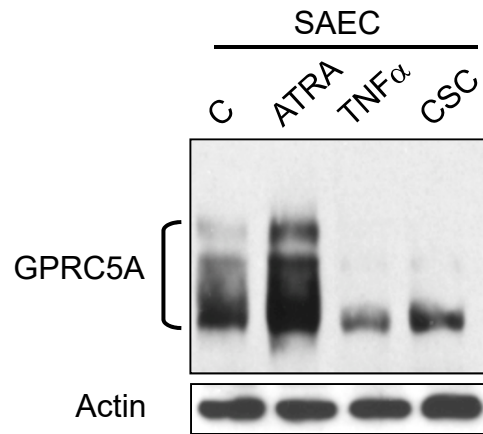
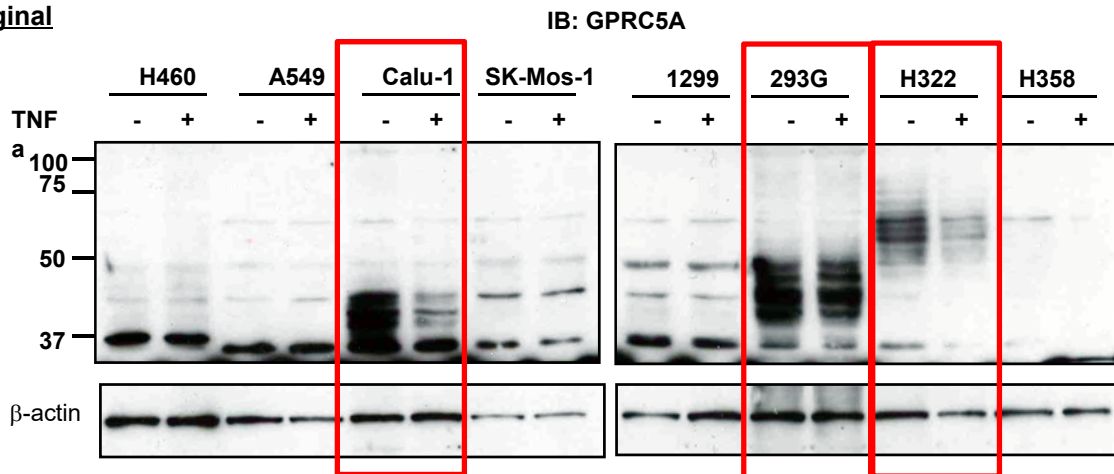


Fig 2B

Original



revised

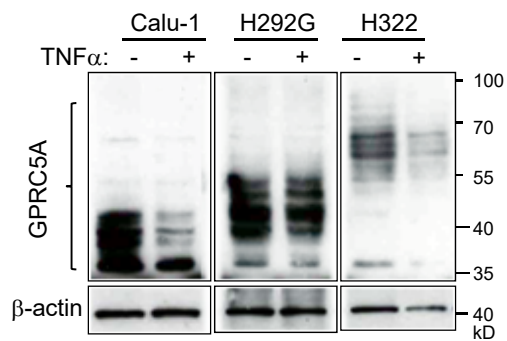


Fig 2F

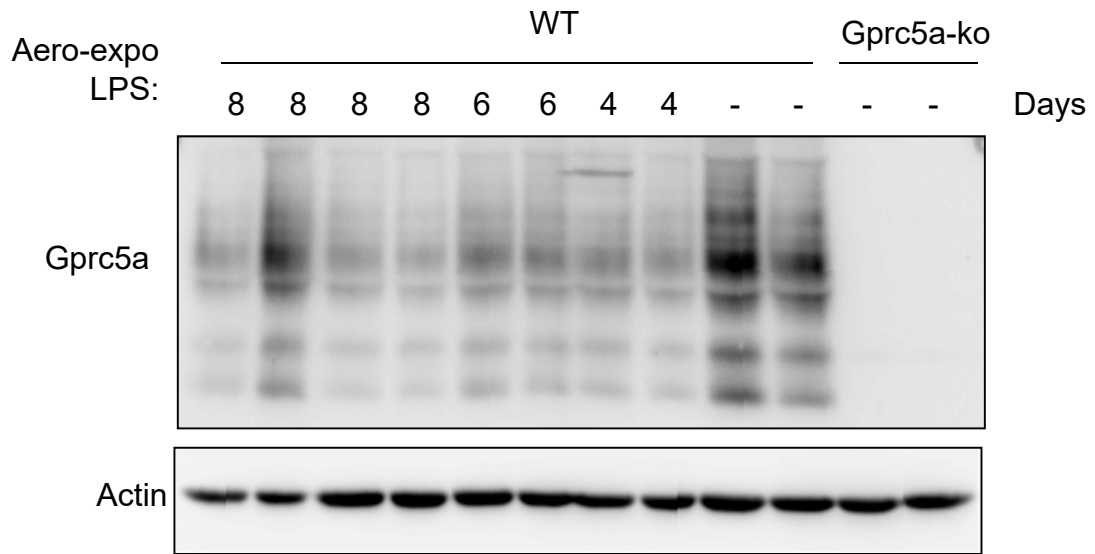


Fig 3D

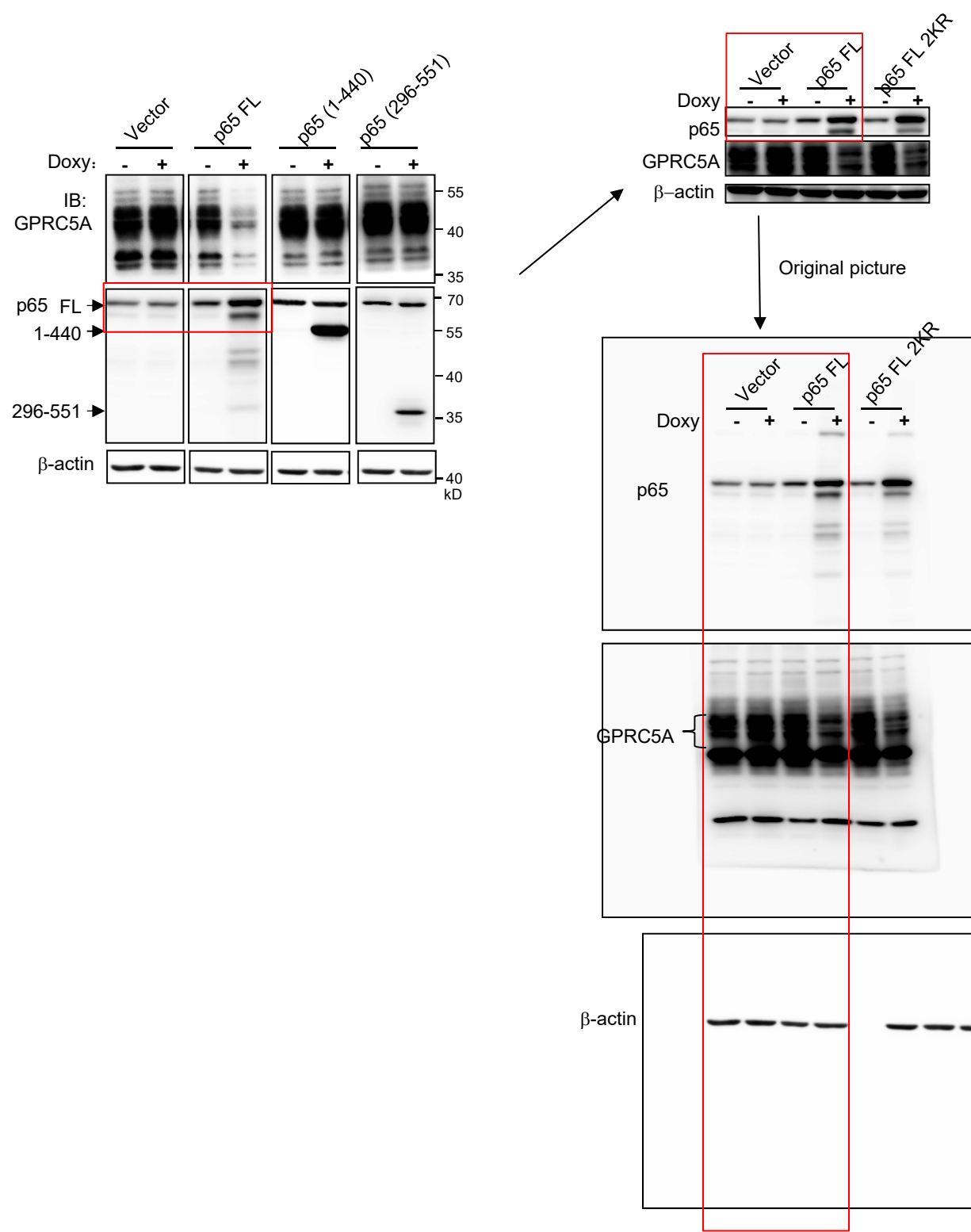


Fig 3D

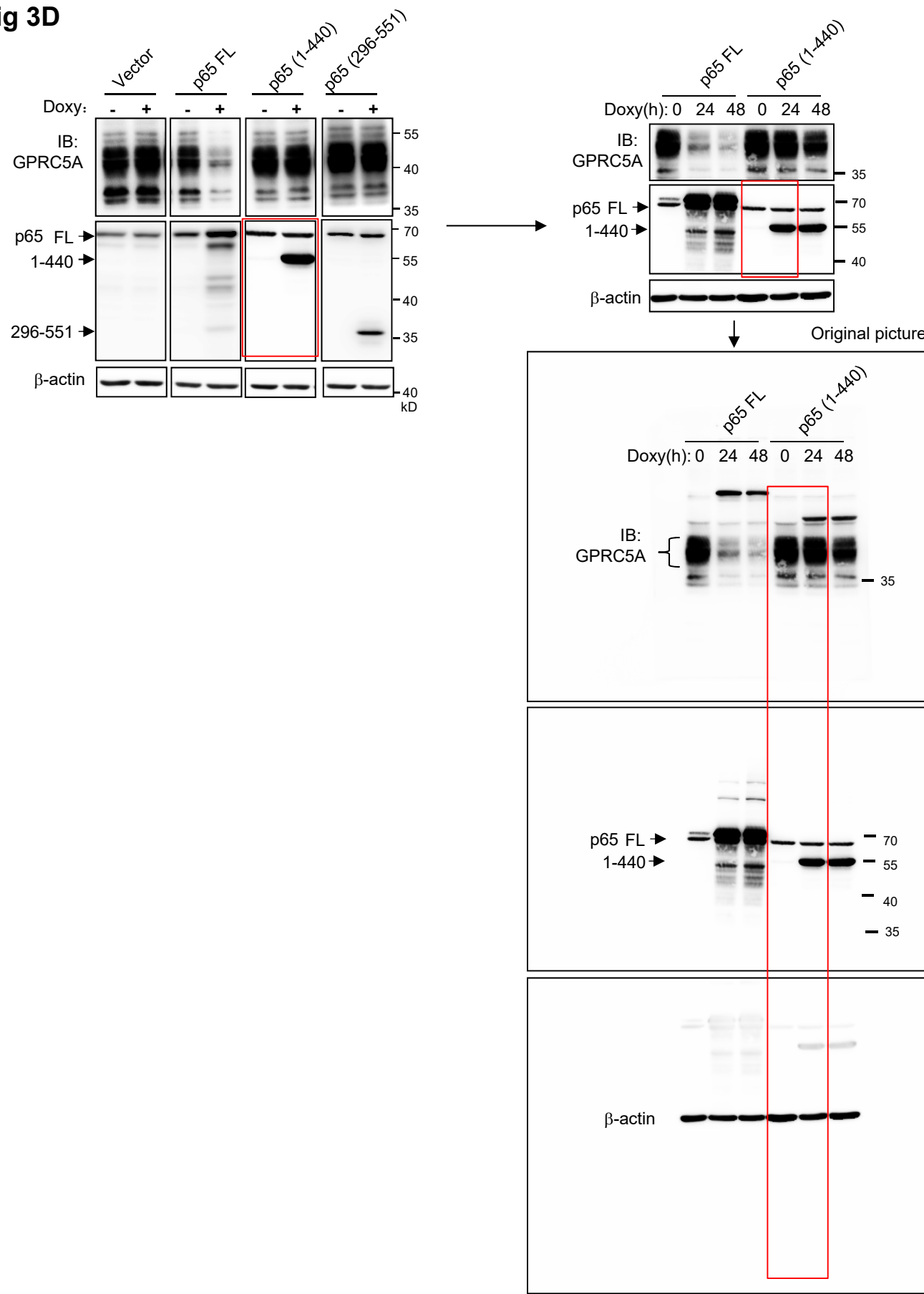
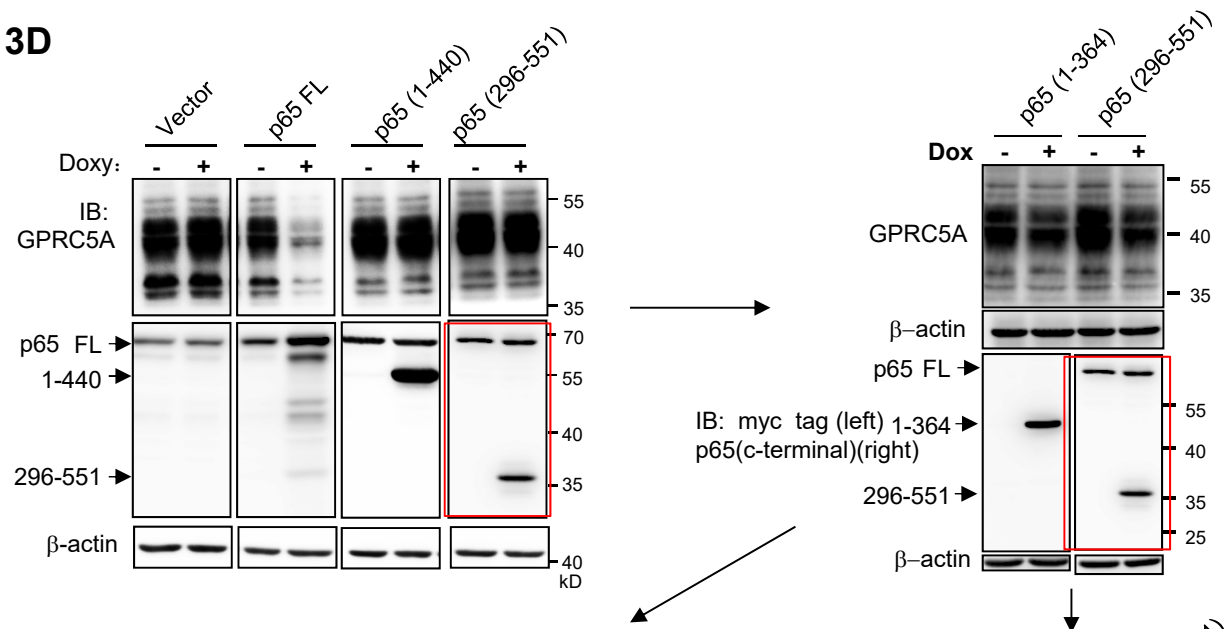
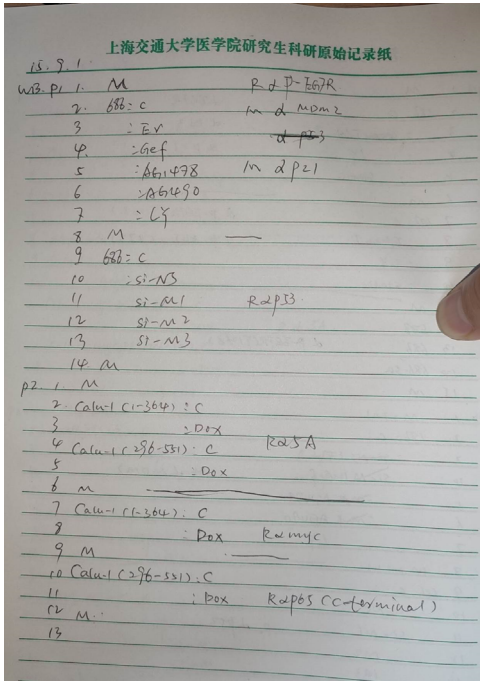


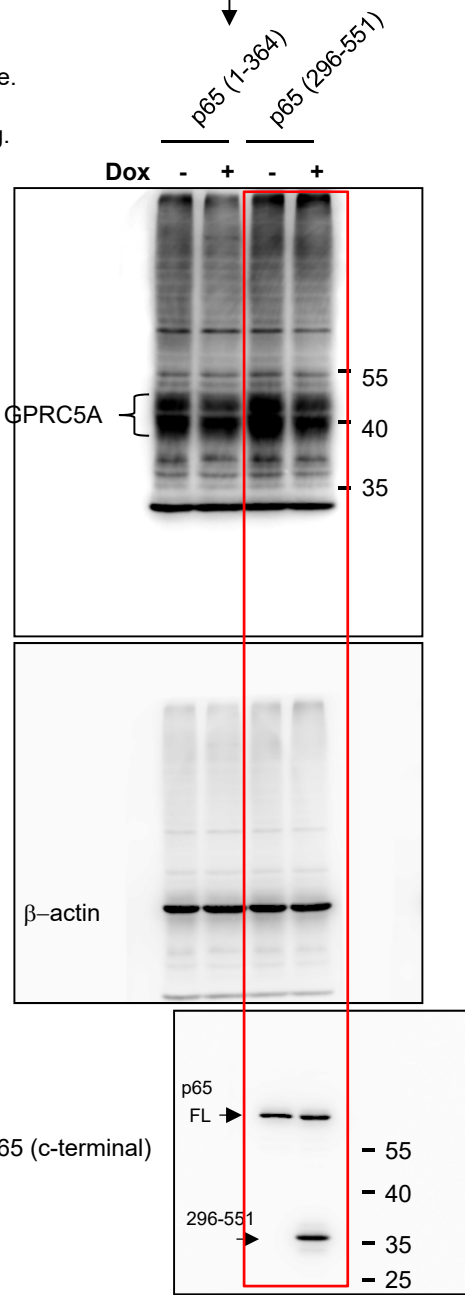
Fig 3D



GPRC5A and p65 blot were run in the same gel, loading the same volume of sample. And then were cropped to blot with different Ab. P65(1-364) were blotted with myc-tag Ab, and because p65(296-551) without any tag. so p65(296-551) were blotted with p65 Ab (target its c-terminal). The following picture was the original experimental record for this blot.



Original picture



β-actin

IB: p65 (c-terminal)

Fig 3E

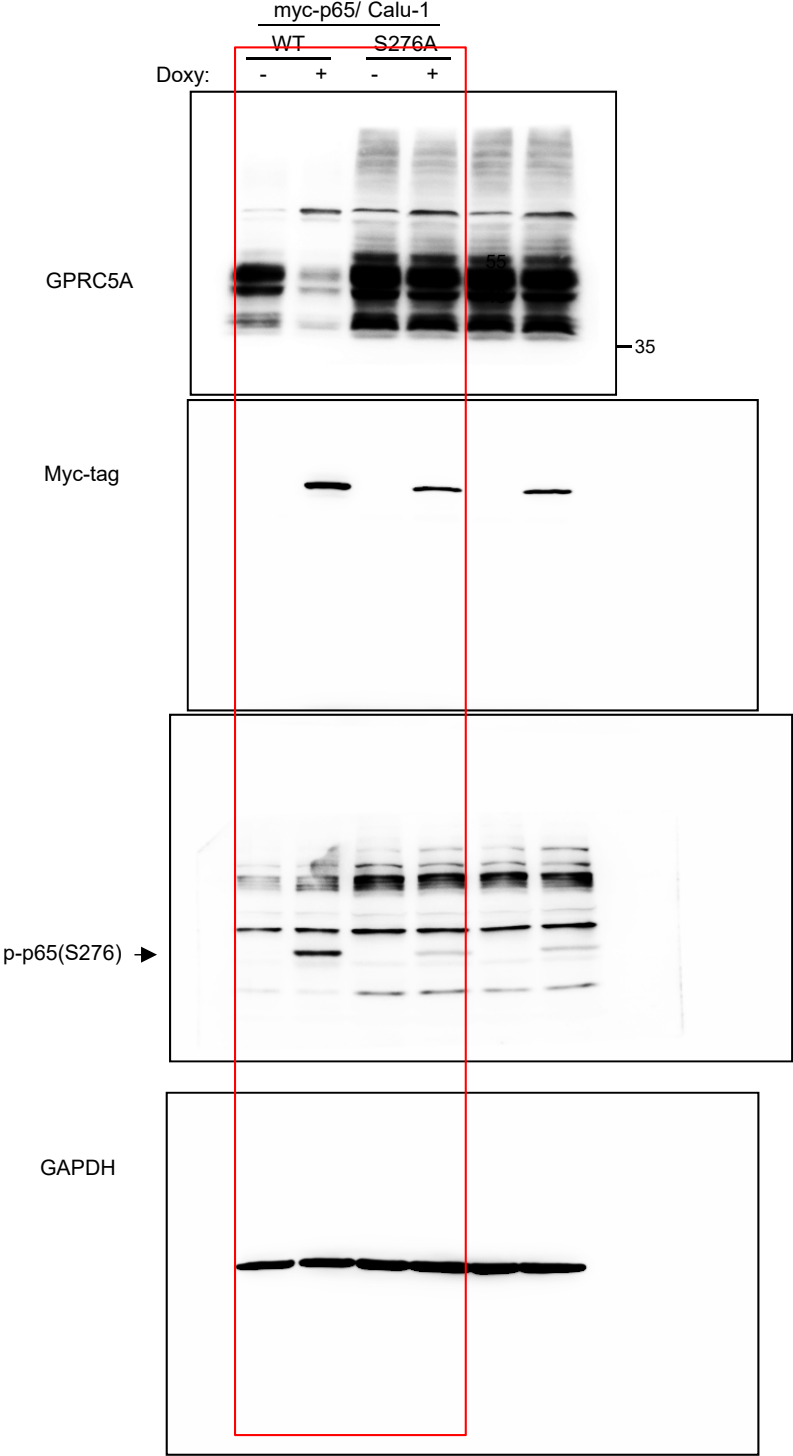


Fig 4

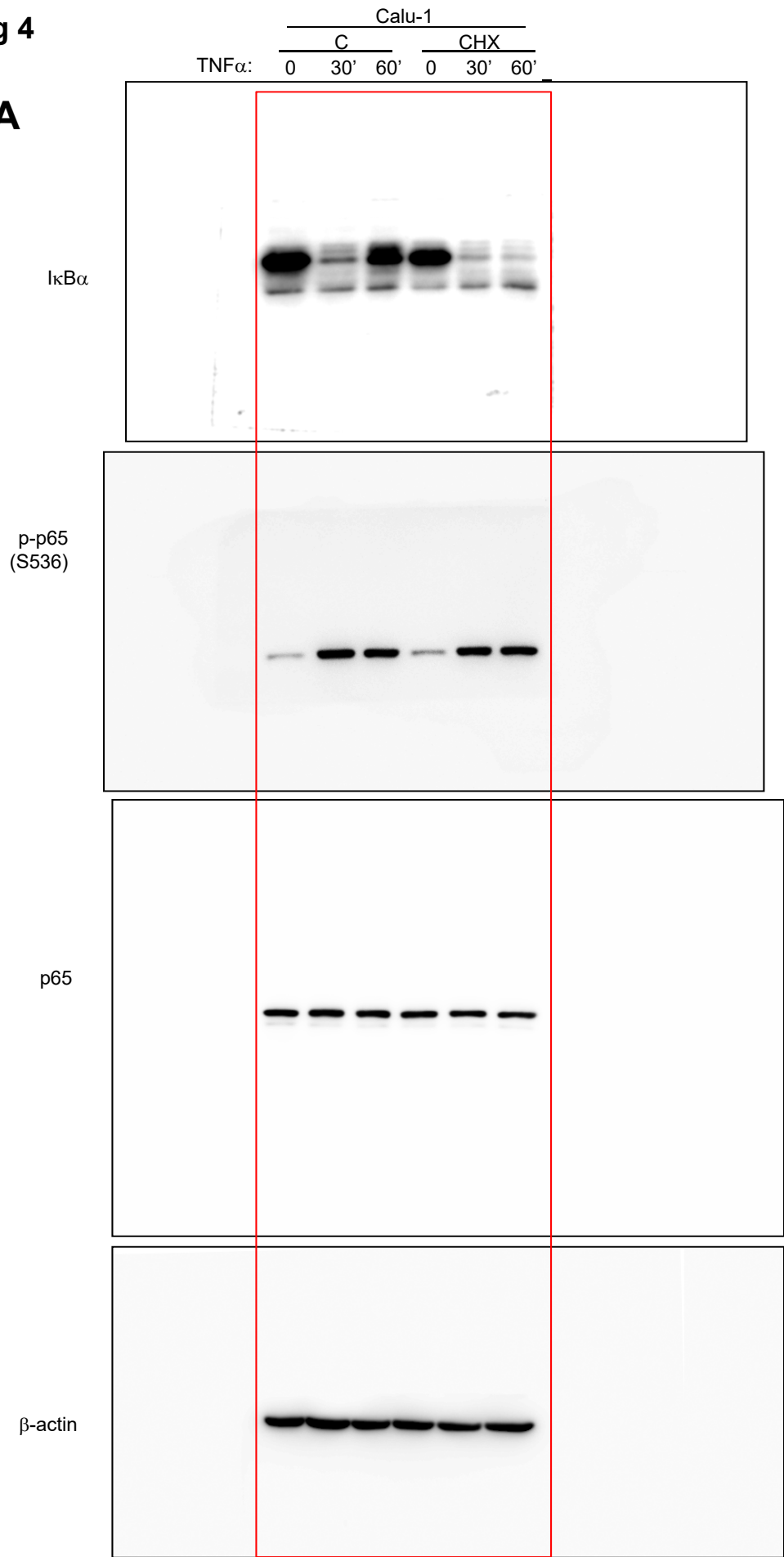
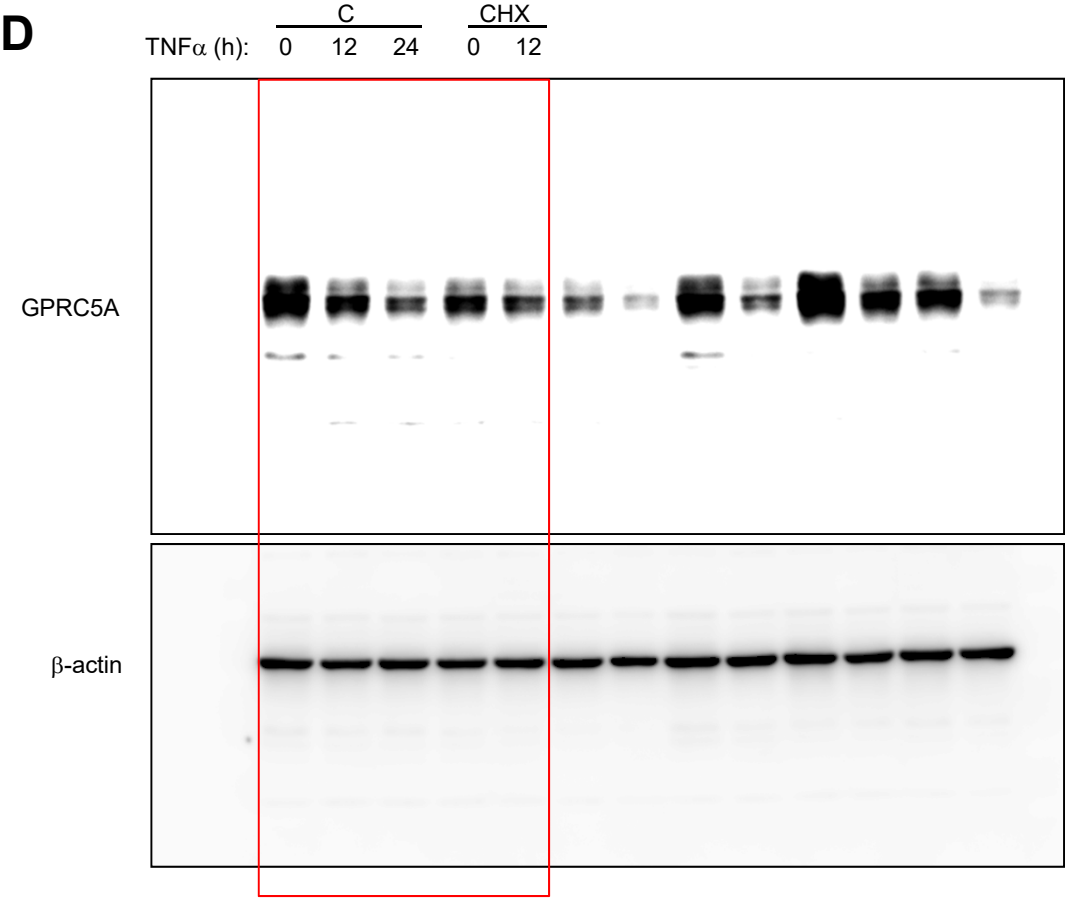


Fig 4



D

	Calu-1			
ATRA:	-	-	+	+
TNF α :	-	+	-	+

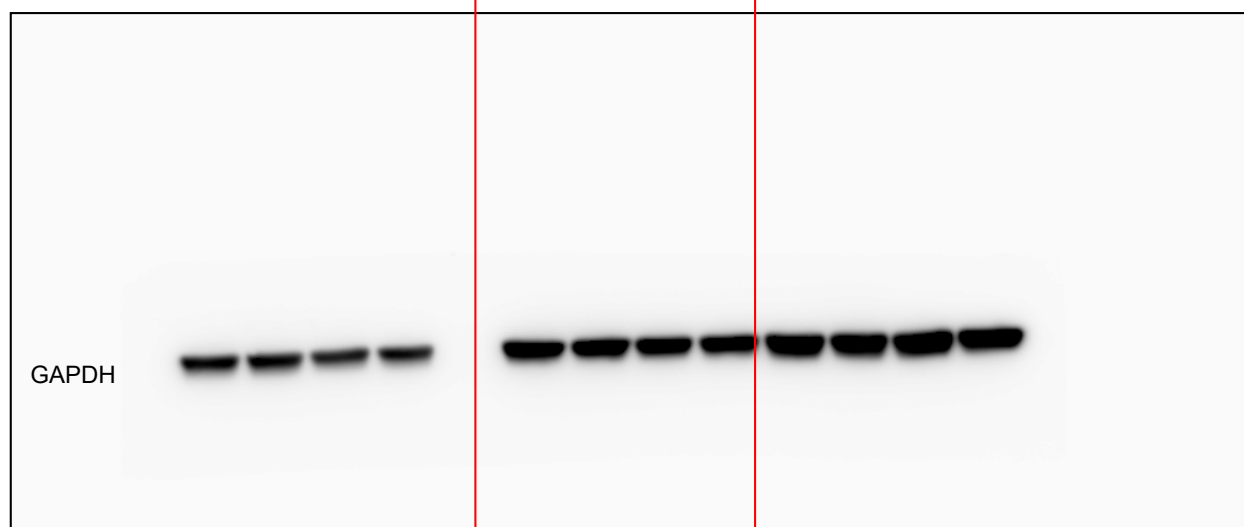
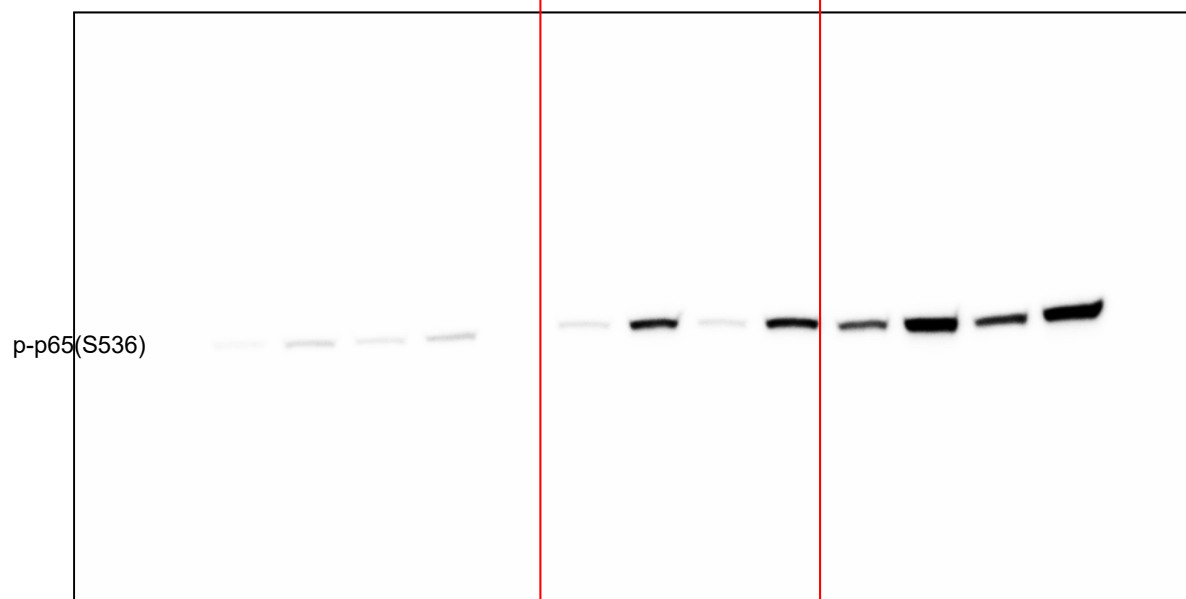
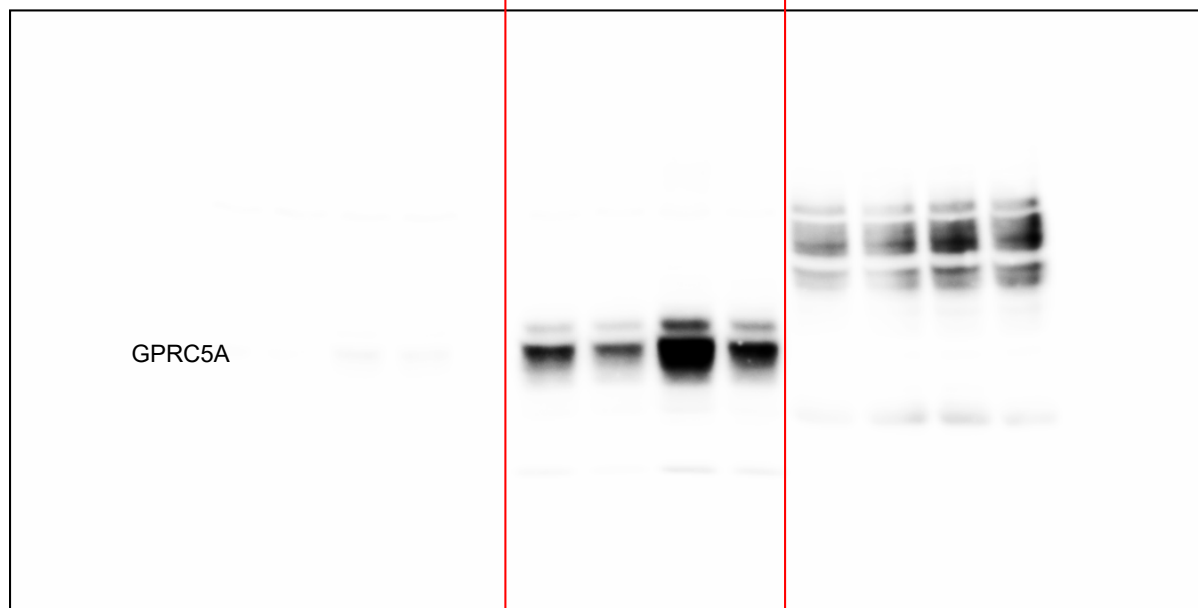


Fig 6 E

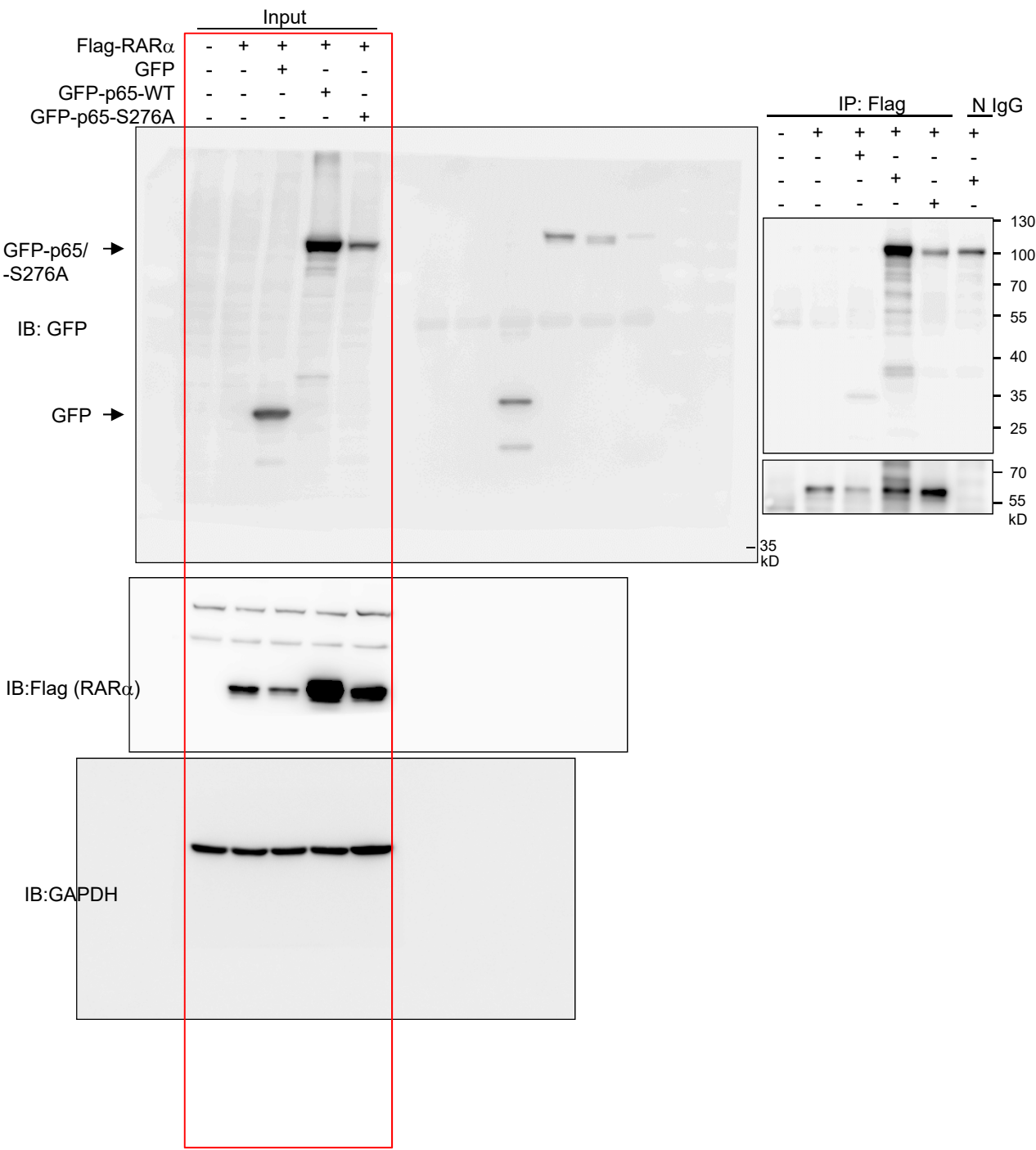


Fig 6 E

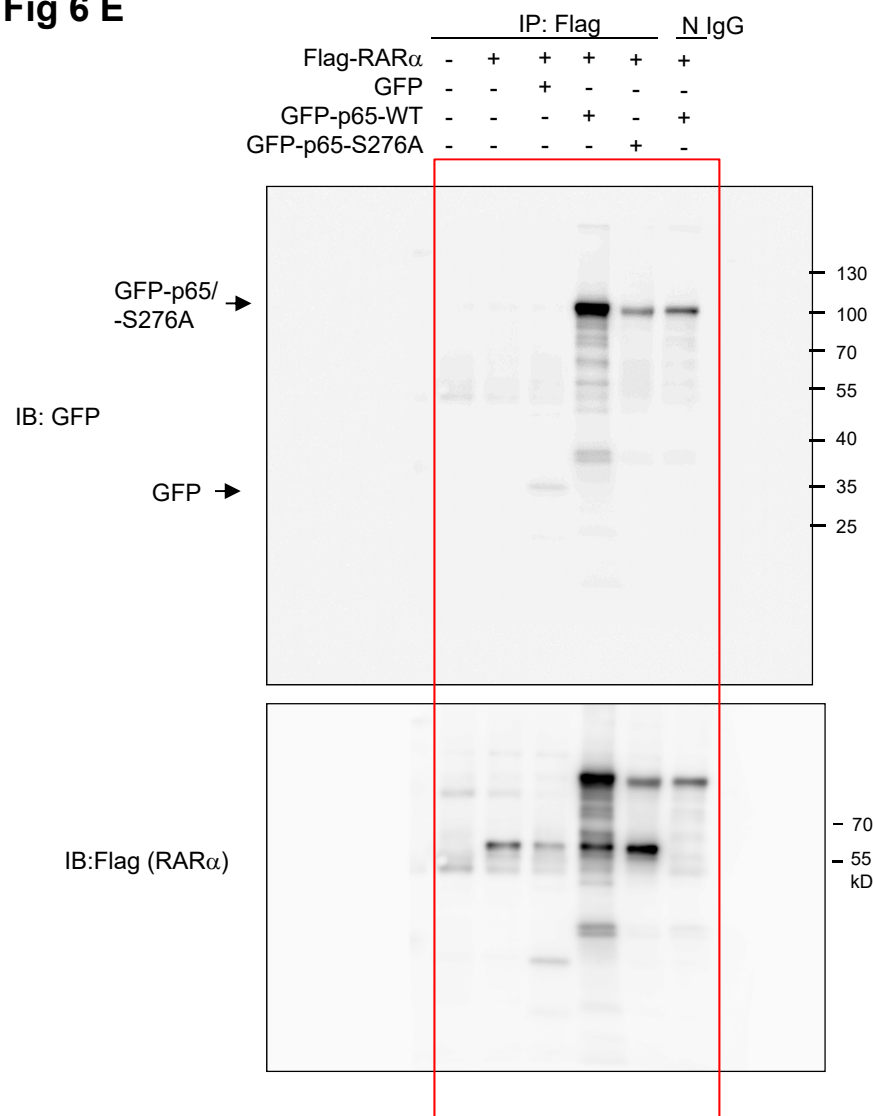


Fig 7

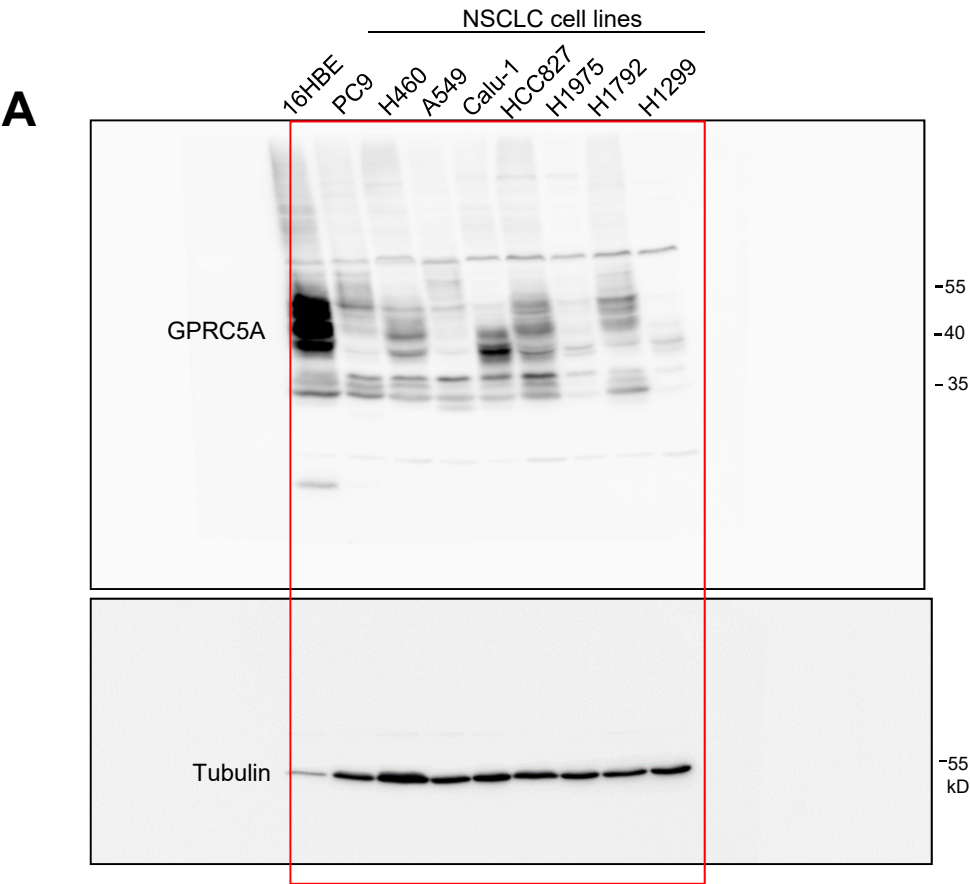


Fig 7

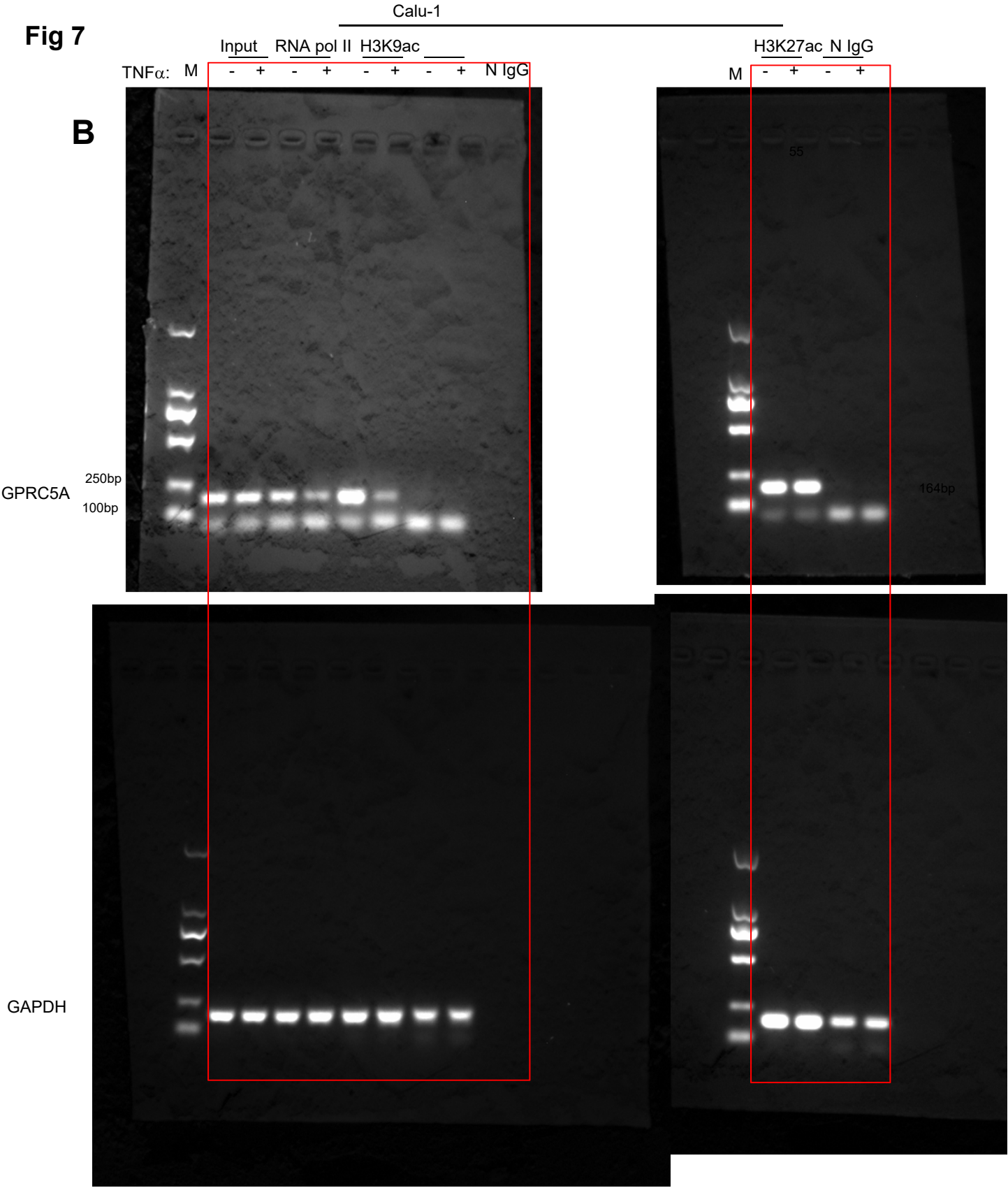


Fig 7

C

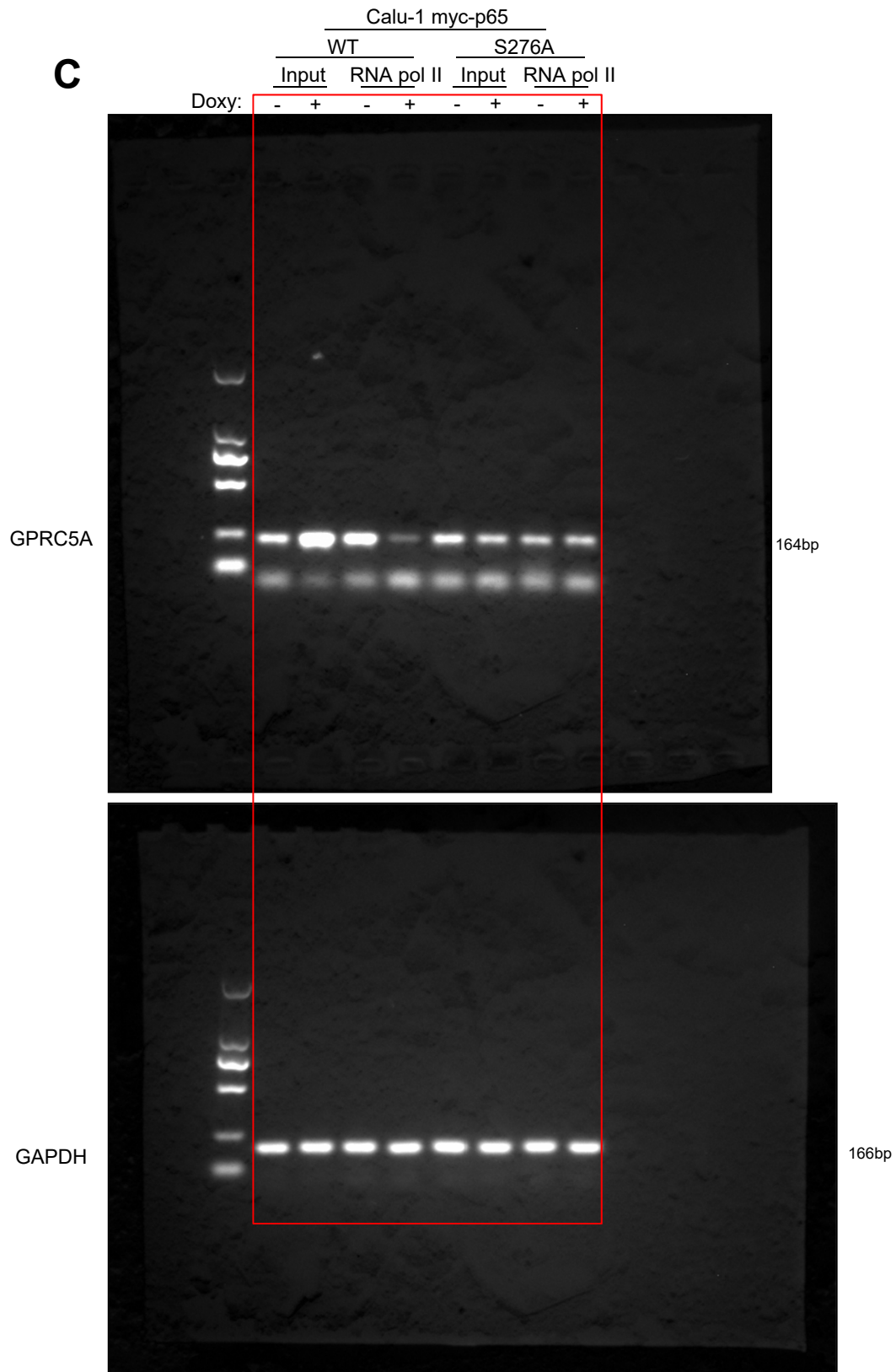


Fig 7

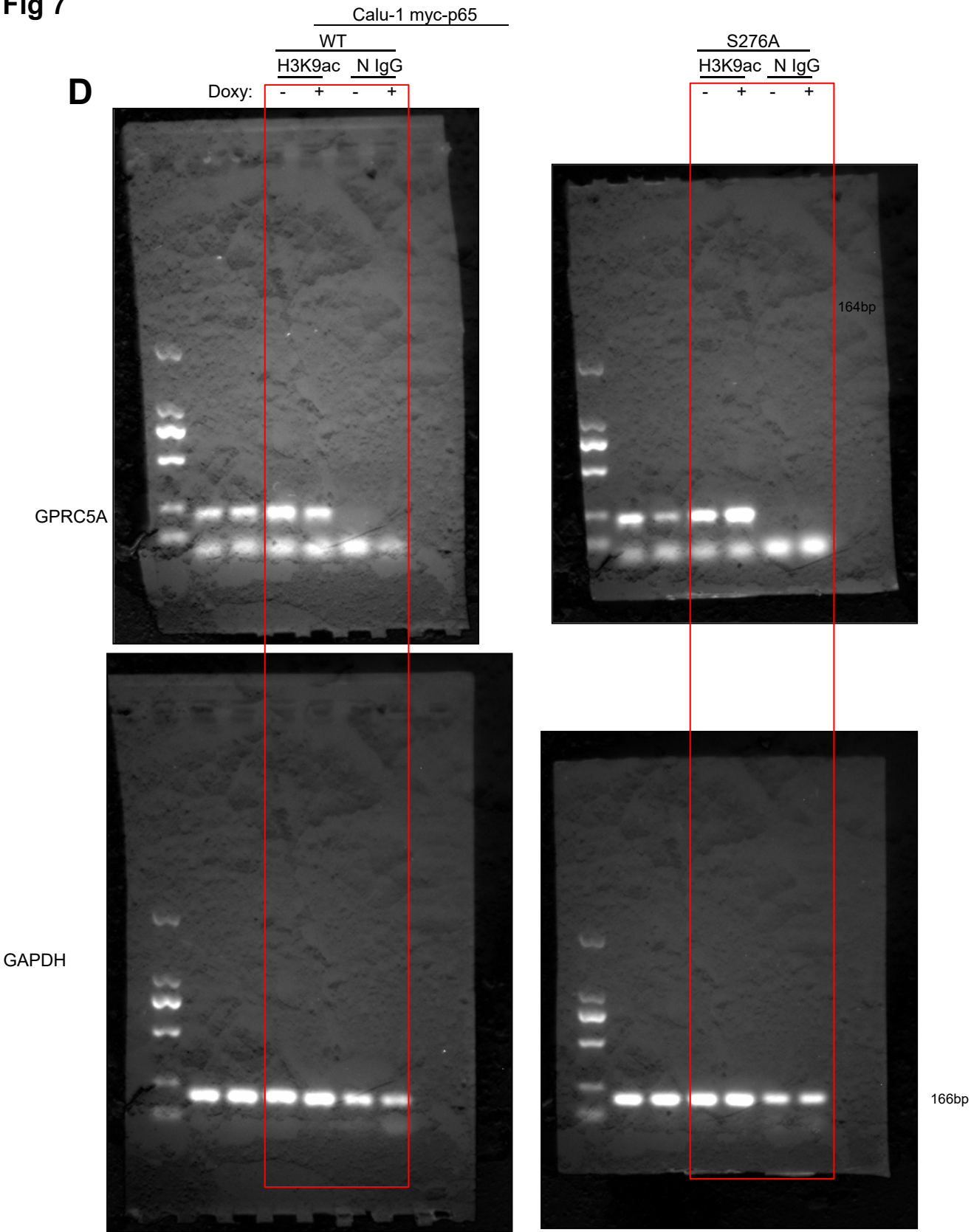


Fig 7

E

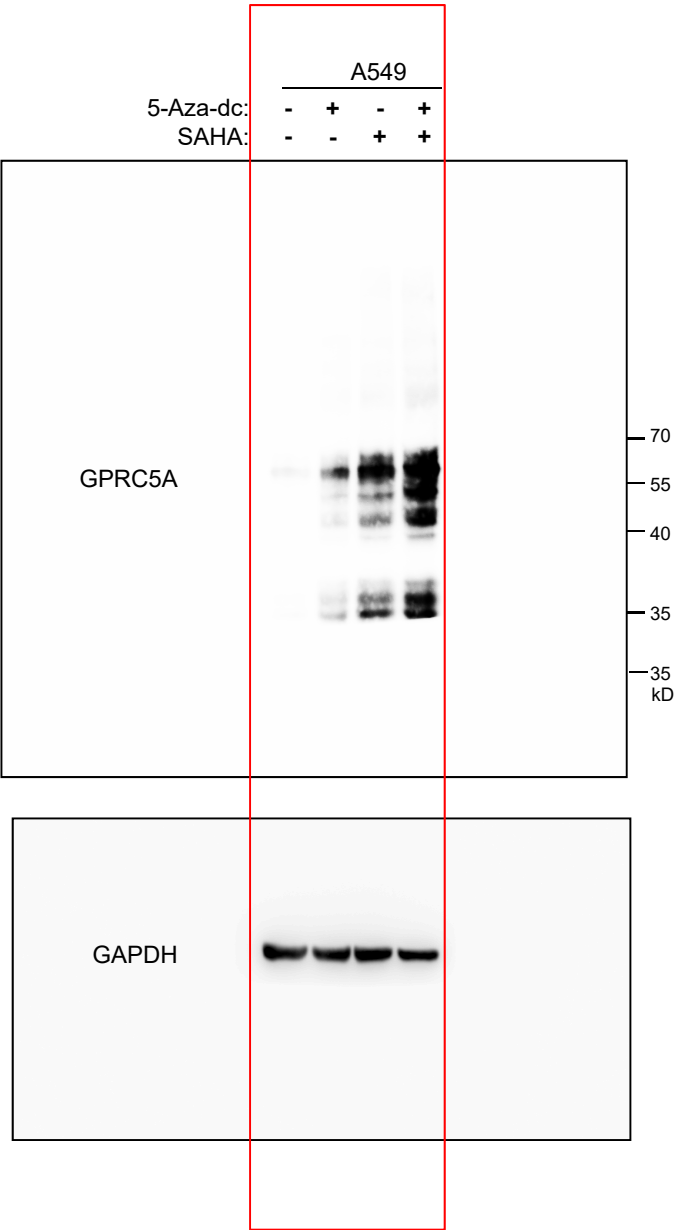


Fig 7

F

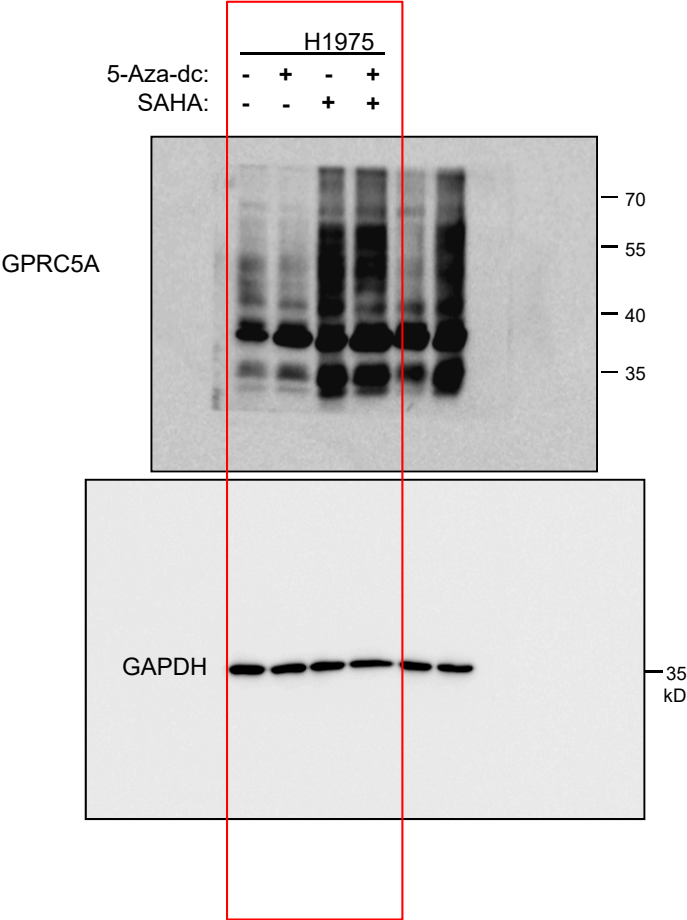


Fig 7

G

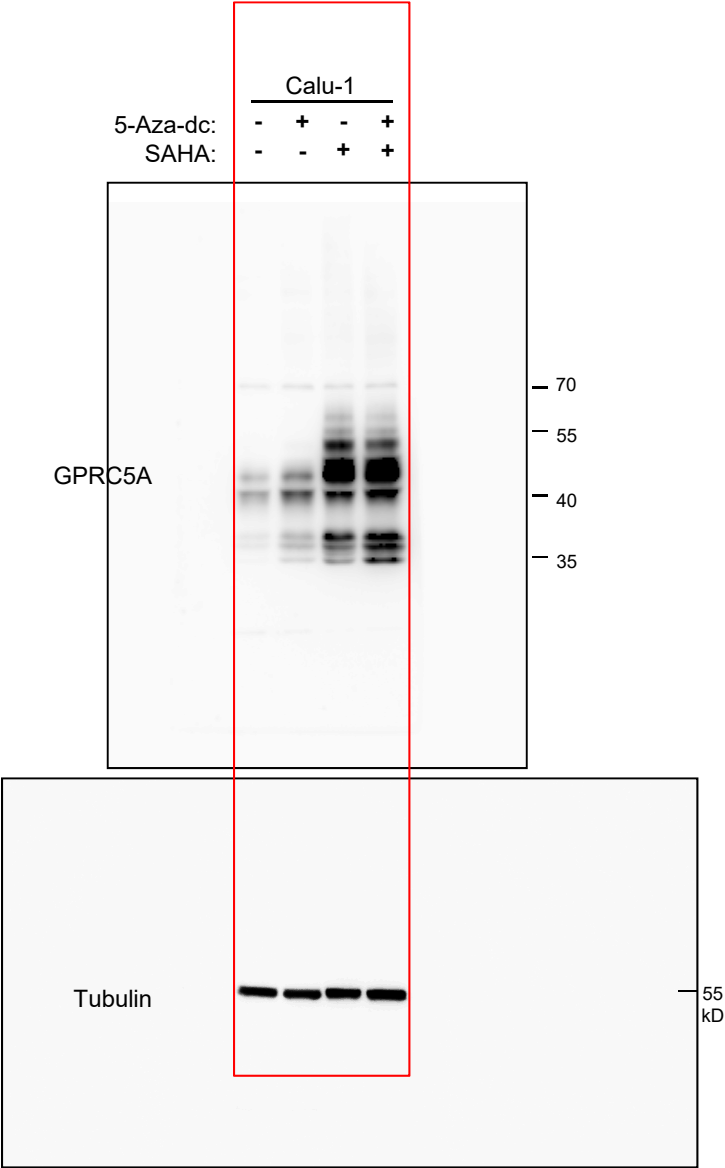


Fig 7

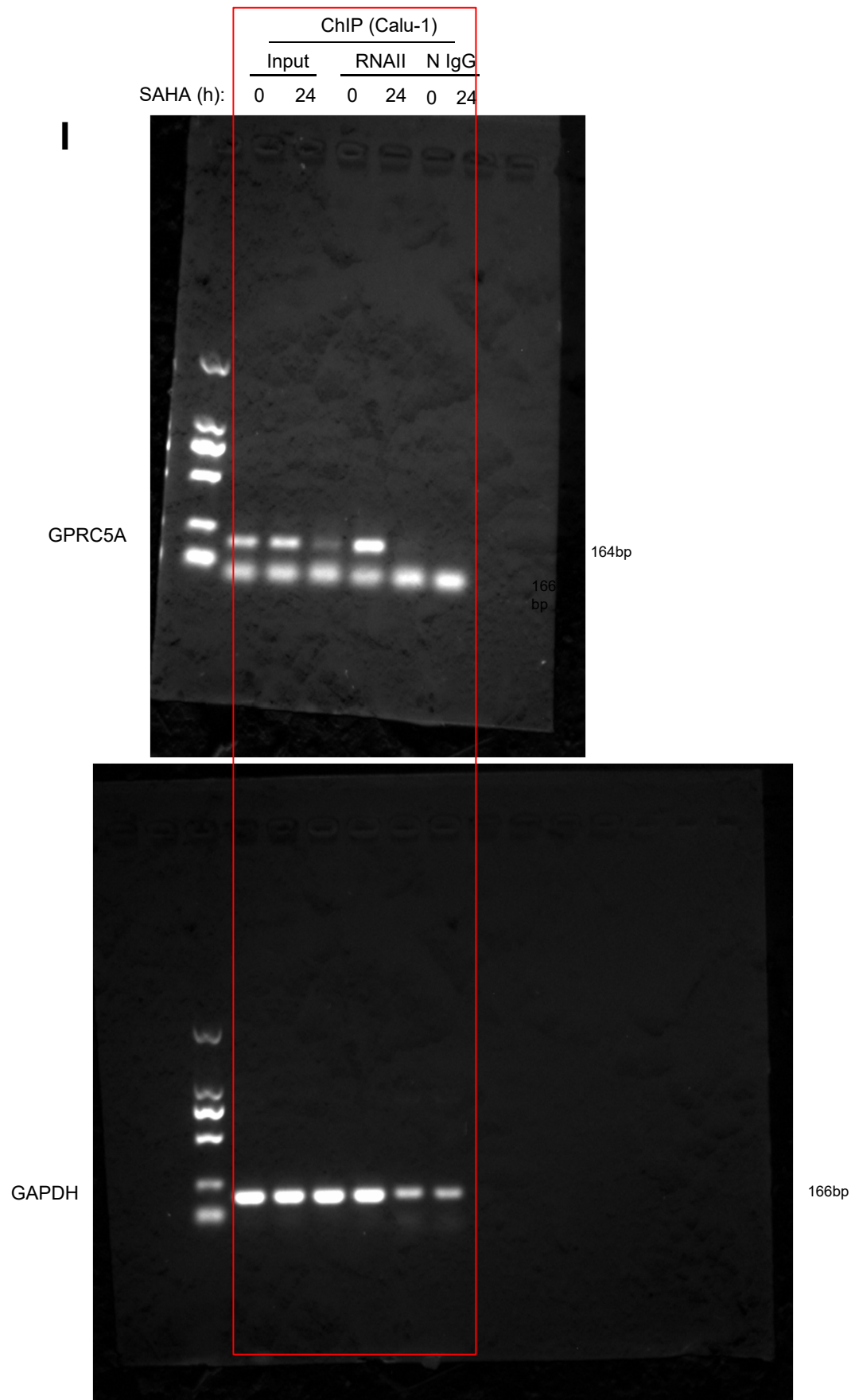


Fig 7

J

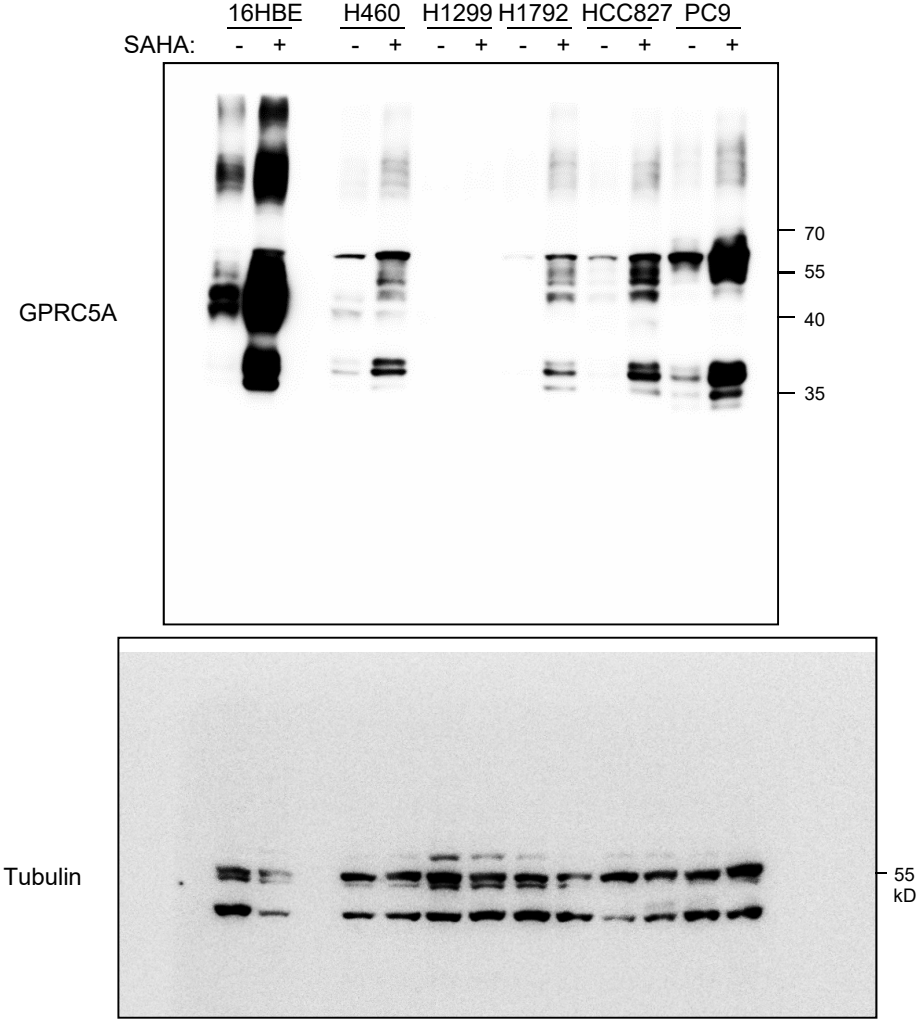


Fig 7

K

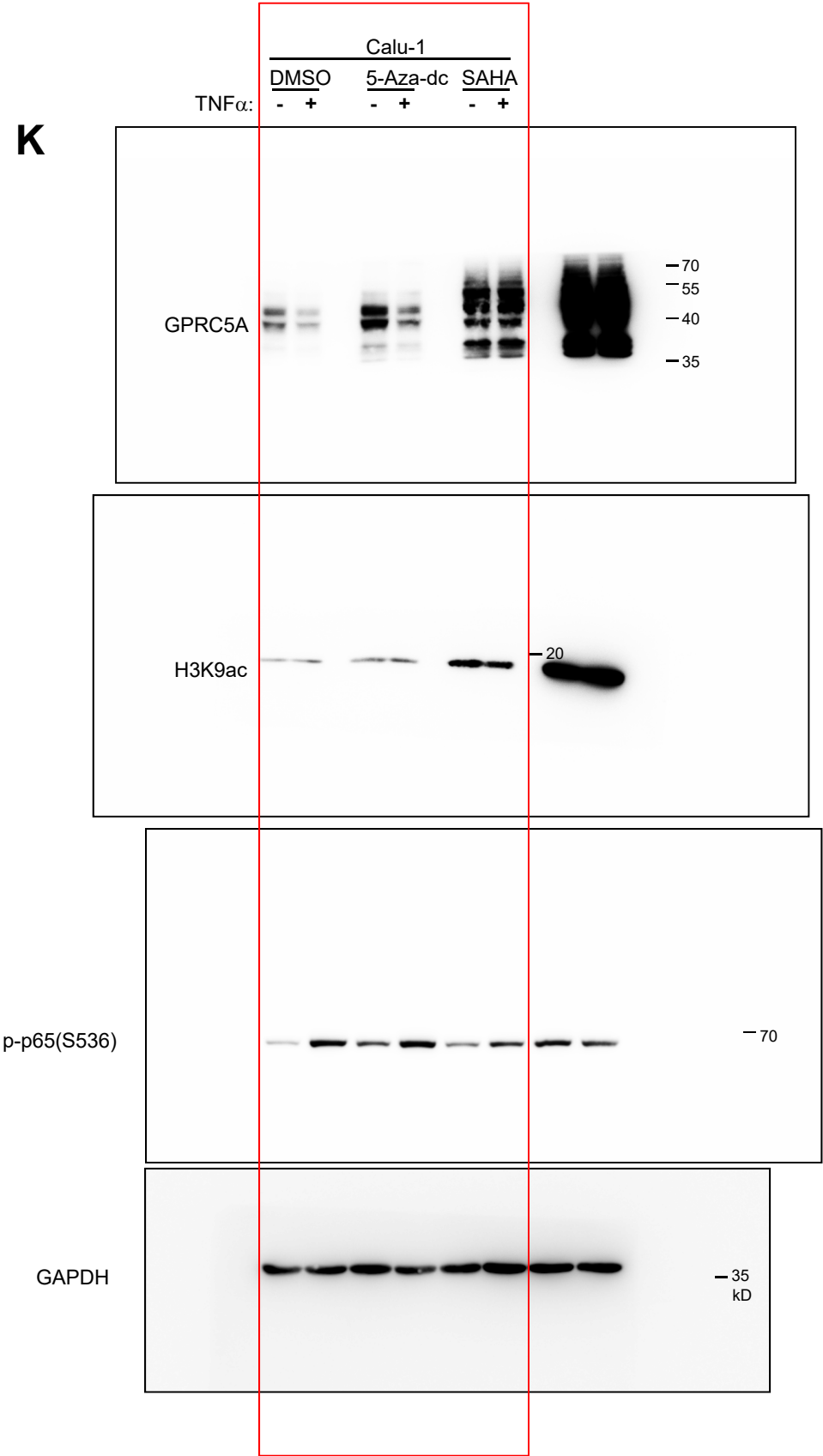
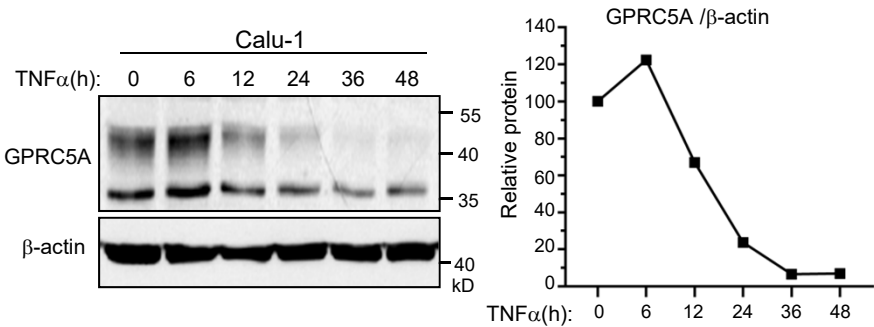


Figure 2D

Old data

Figure 2D



New data

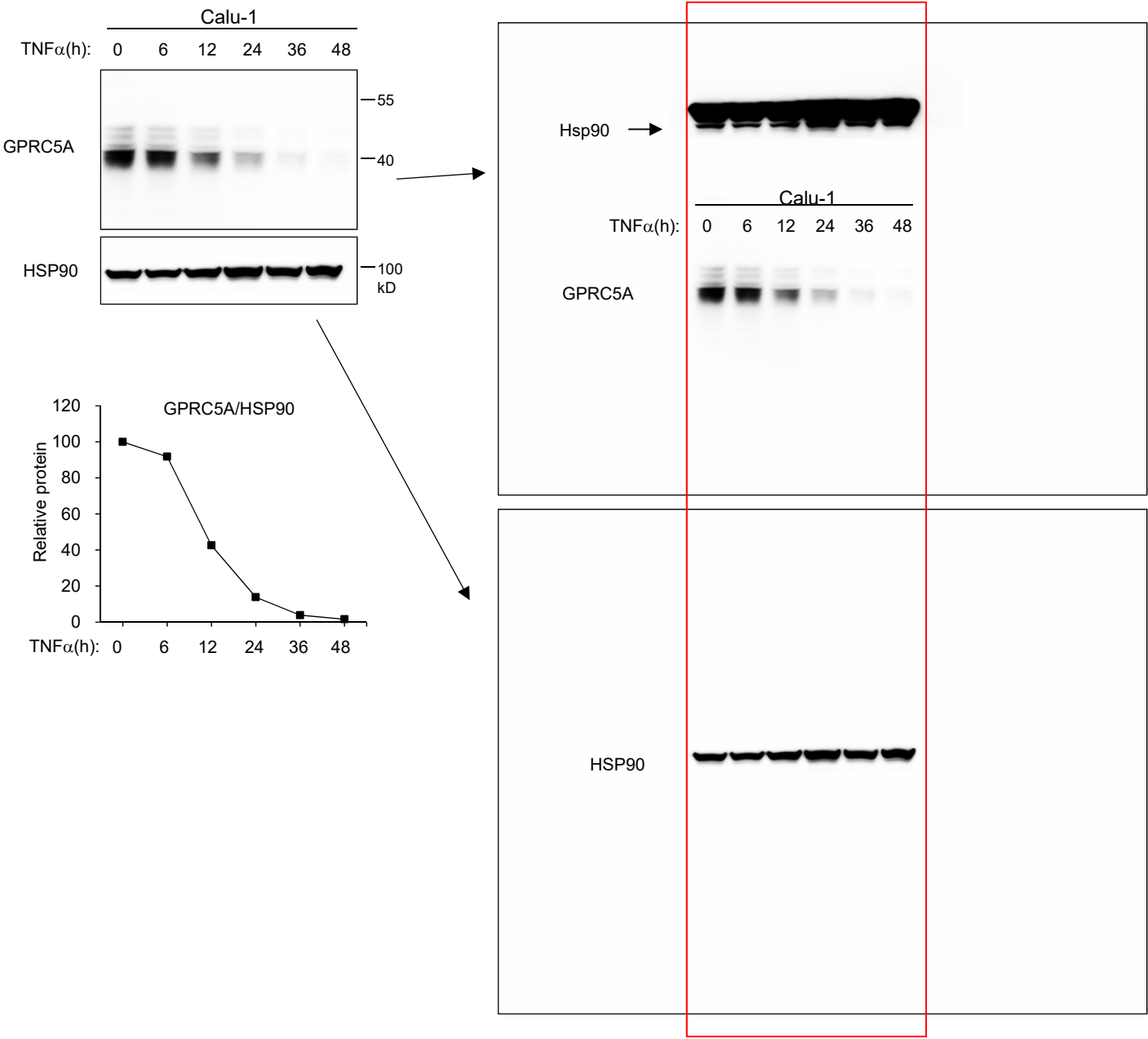


Figure 3A

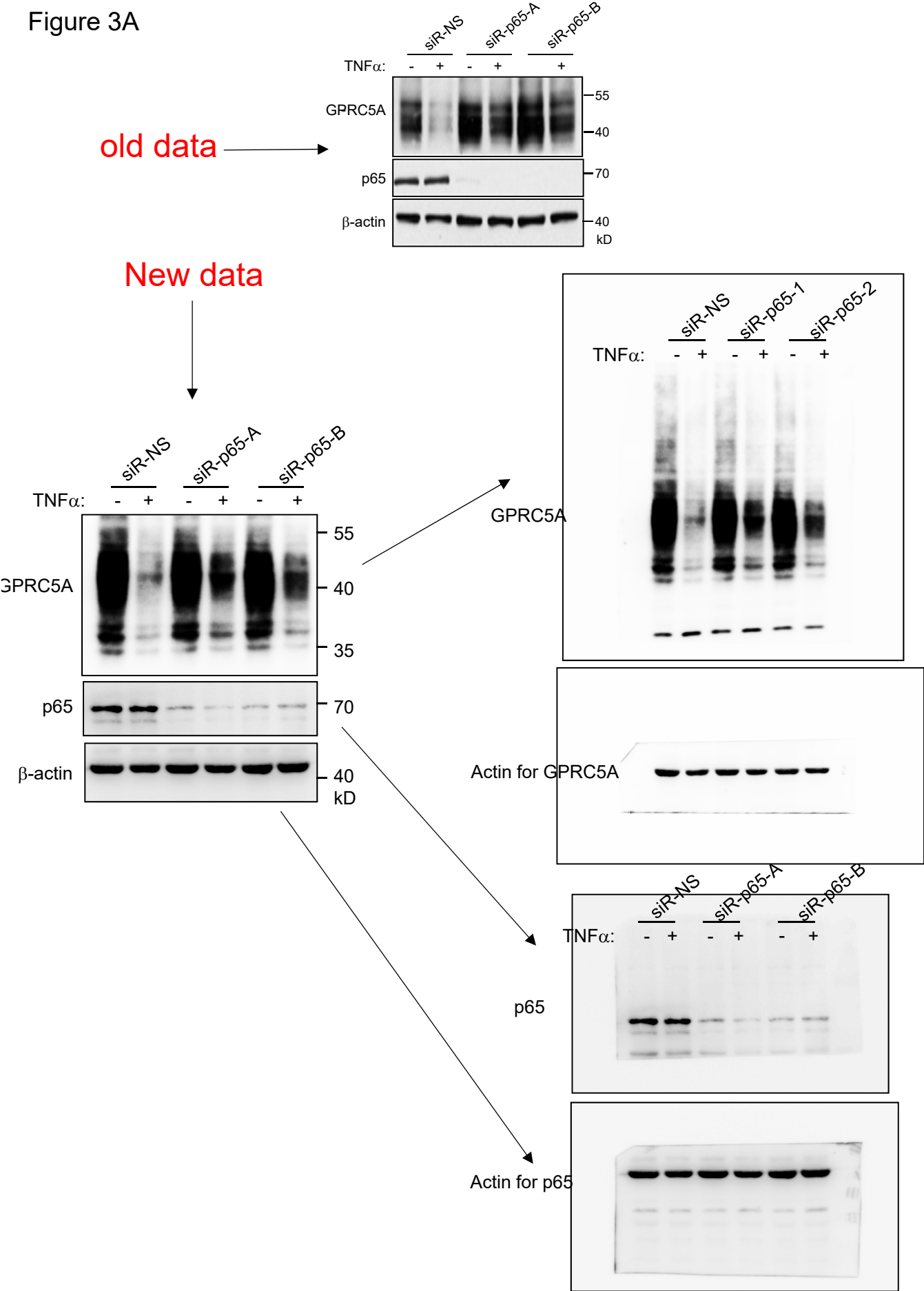


Figure 3B

old data

New data

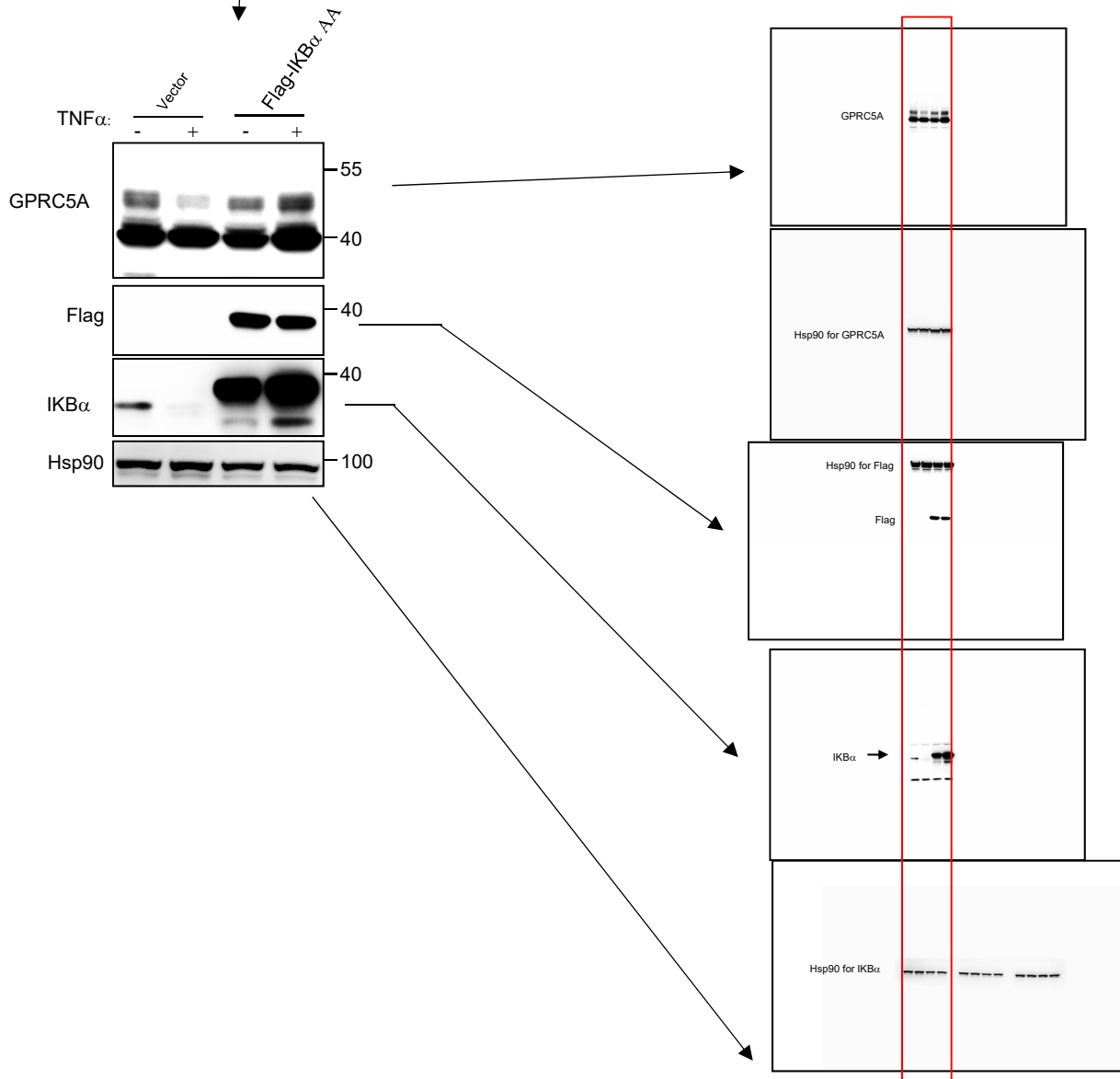
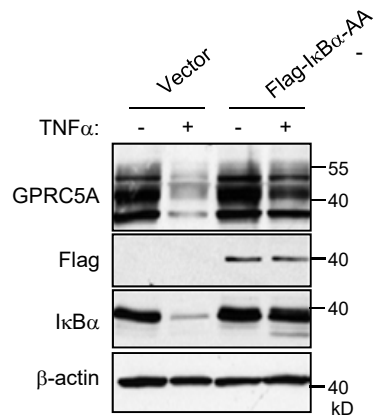
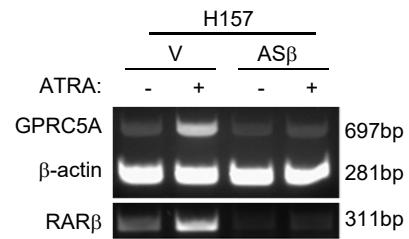


Figure 5

C

Old data



New data

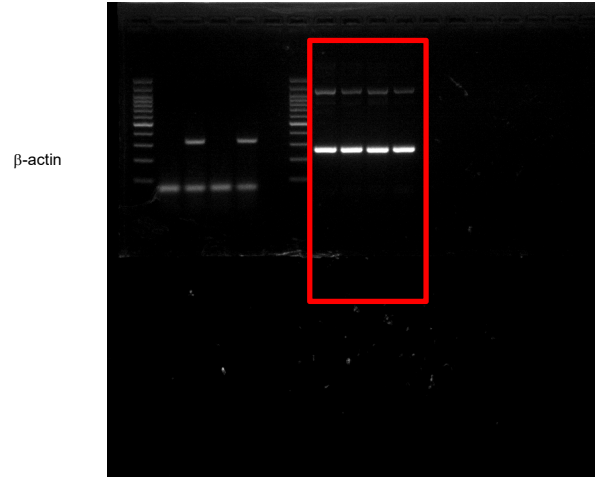
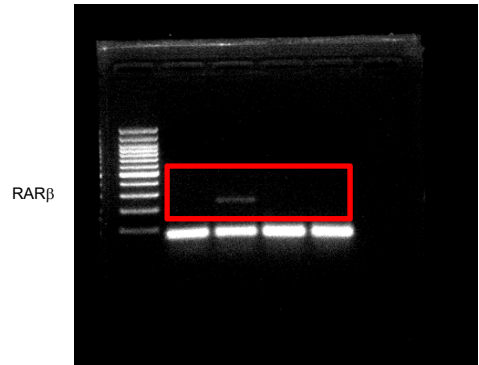
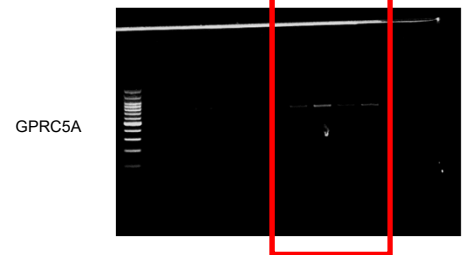
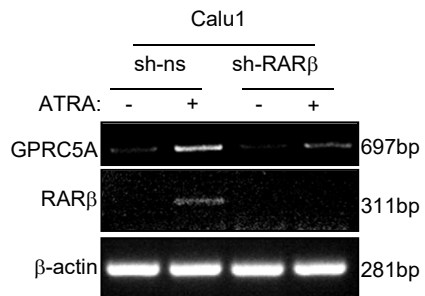


Figure 5B

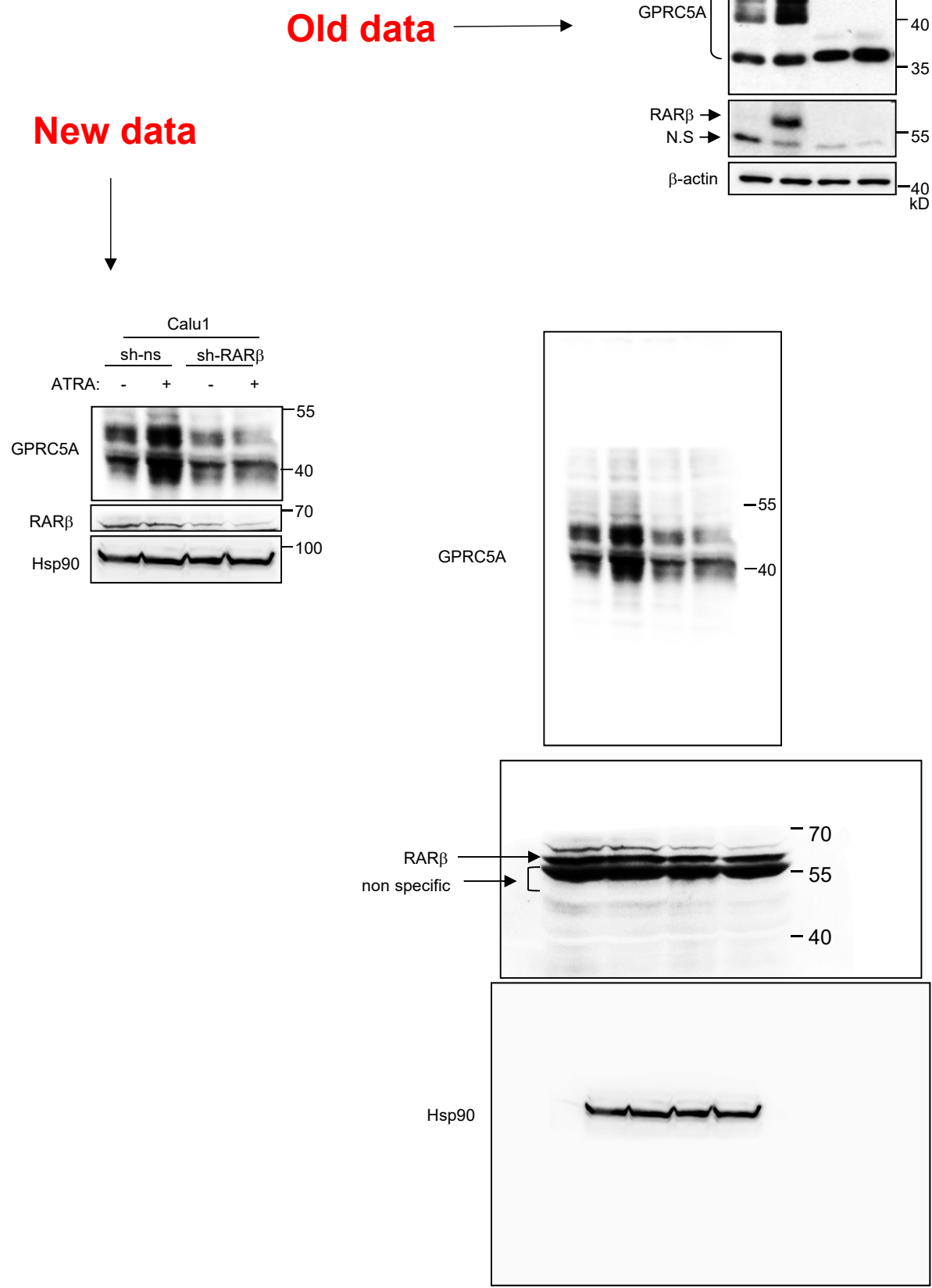
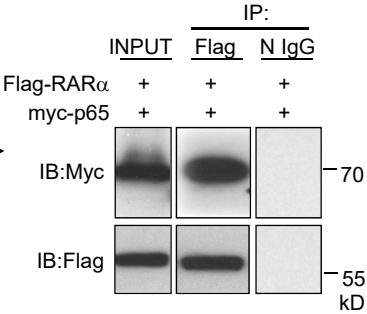


Figure 6C

Old data



New data

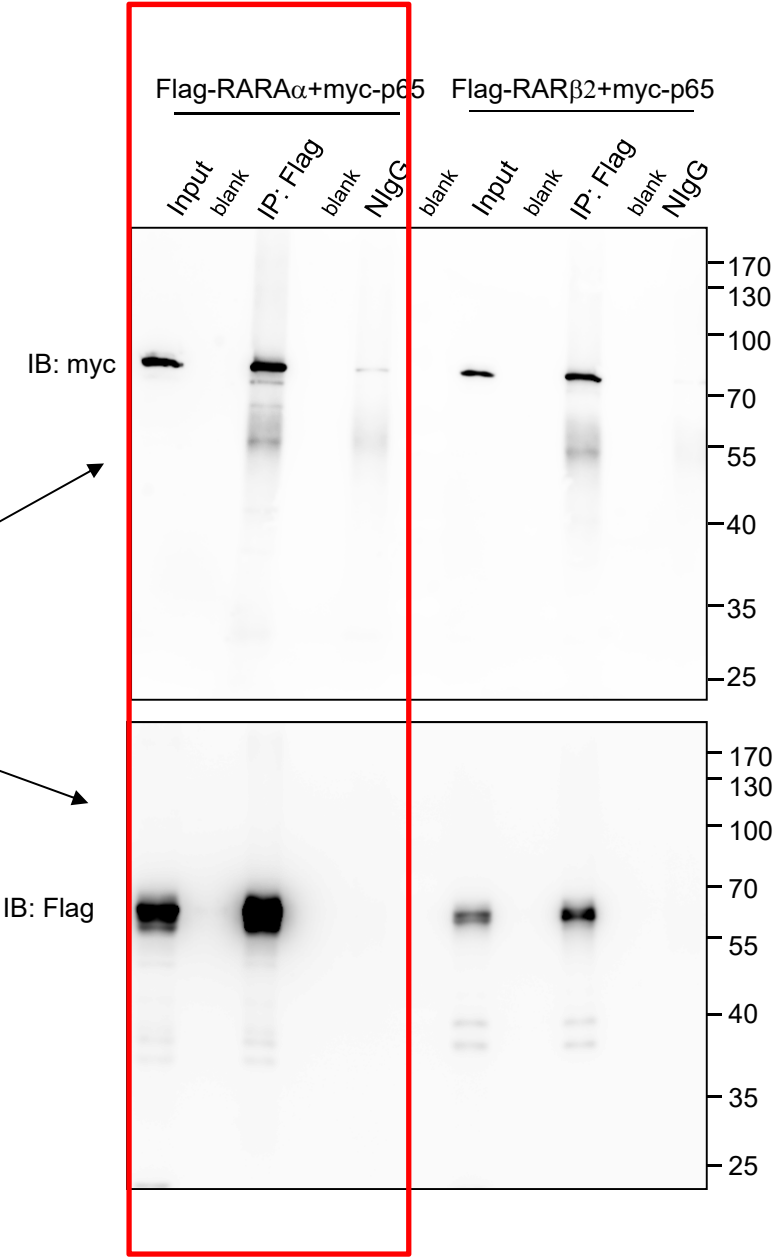
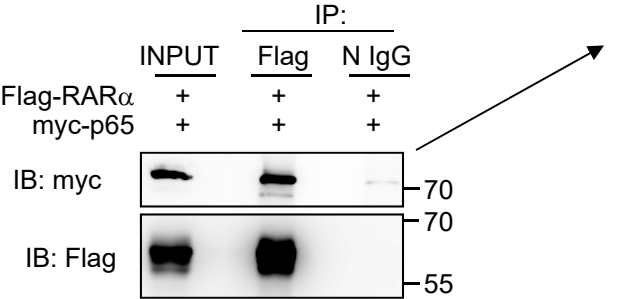
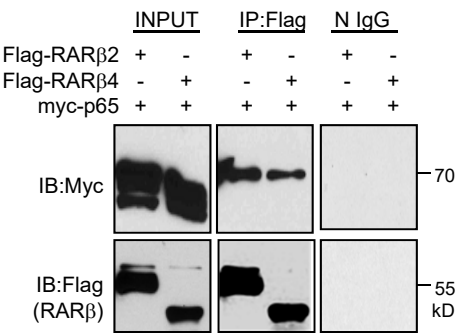
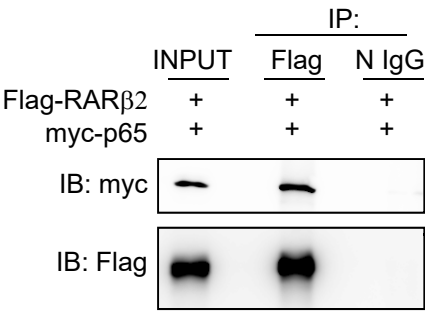


Figure 6D

Old data →



New data



IB: myc

IB: Flag

Flag-RARAα+myc-p65

Flag-RARβ2+myc-p65

