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Donor Genetic and Non-Genetic Factors Affecting Red Blood Cell Transfusion Effectiveness

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Abbreviations table:

Hb	hemoglobin
RBCs	red blood cells
SNP	single nucleotide polymorphism
G6PD	Glucose 6-phosphate deficiency
REDS	Recipient Epidemiology and Donor Evaluation Study
NIH	National Institutes of Health
NHLBI	National Heart Lung and Blood Institute
FDA	Food and Drug Administration
IQR	interquartile range
US	United States
CI	confidence interval

ABSTRACT

Background: Red blood cell (RBC) transfusion effectiveness varies due to donor, component, and recipient factors. Prior studies identified characteristics associated with variation in hemoglobin increments following transfusion. We extended these observations, examining donor genetic and non-genetic factors affecting transfusion effectiveness.

Methods: This is a multicenter retrospective study of 46,705 patients, and 102,043 evaluable RBC transfusions from 2013-2016 across 12 hospitals. Transfusion effectiveness was defined as hemoglobin, bilirubin, or creatinine increments following single RBC unit transfusion. Models incorporated a subset of donors with data on single nucleotide polymorphisms associated with osmotic and oxidative hemolysis *in vitro*. Mixed modelling accounting for repeated transfusion episodes identified predictors of transfusion effectiveness.

Results: Blood donor (sex, Rh status, fingerstick hemoglobin, smoking), component (storage duration, gamma irradiation, leukoreduction, apheresis collection, storage solution), and recipient (sex, body mass index, race, age) characteristics were associated with hemoglobin and bilirubin but not creatinine increments following RBC transfusions. Increased storage duration was associated with increased bilirubin and decreased hemoglobin increments, suggestive of *in vivo* hemolysis following transfusion. Donor *G6PD*-deficiency and polymorphisms in *SEC14L4*, *HBA2*, and *MYO9B* genes were associated with decreased hemoglobin increments. Donor *G6PD*-

deficiency and polymorphisms in *SEC14L4* were associated with increased transfusion requirements in the subsequent 48 hours.

Conclusion: Donor genetic and other factors, such as RBC storage duration, affect transfusion effectiveness as defined by decreased hemoglobin or increased bilirubin increments. Addressing these factors will provide a precision medicine approach to improve patient outcomes, particularly for chronically-transfused RBC recipients, who would most benefit from more effective transfusion products.

INTRODUCTION

Red blood cell (RBC) transfusions can be a critical life-saving intervention for hospitalized and chronically transfused patients. In contrast to pharmaceutical-grade drugs, there is substantial variability in hemoglobin dose and RBC quality among units of blood (1, 2). Blood donors are a heterogenous population for which biologic, genetic, and behavioral variables interact with various manufacturing and storage conditions to contribute to variation in the hemoglobin dose of packed RBC units and their responses to cold storage. For logistical reasons, RBCs are refrigerator-stored for up to 6 weeks in the United States, resulting in altered RBC biology (i.e., the “storage lesion”) (3, 4). A subset of storage-damaged RBCs is cleared from the circulation after transfusion releasing large amounts of hemoglobin, which is primarily catabolized into iron and bilirubin (2, 5, 6). Finally, intravascular hemolysis may affect kidney function and exacerbate other human diseases (7, 8). Thus, because these laboratory values are often measured in hospitalized patients requiring transfusion, we assessed transfusion effectiveness by the change in hemoglobin, bilirubin, or creatinine levels following transfusion.

Despite substantial inter-subject variability in how RBCs tolerate refrigerated storage, there is considerable intra-subject reproducibility in storage quality (i.e., some donors are “good storers” whereas others are “poor storers”) (2, 9-12). It is increasingly recognized that blood donor demographics, social behaviors, and genetic factors play a role in stored blood product quality. A key focus of transfusion medicine research is to understand the role of donor genetics in observed RBC storage variability (4, 13). Glucose-6-phosphate dehydrogenase (G6PD) deficiency, an X-linked disorder affecting

the ability of RBCs to handle oxidant stress, is the most common human enzymopathy, affecting ~400 million people worldwide (14, 15). Blood donor G6PD deficiency increases the susceptibility of RBCs to the storage lesion, resulting in decreased post-transfusion recovery of transfused RBCs (12). More recently, several other novel and known genetic polymorphisms were found to be associated with measures of osmotic and oxidative hemolysis *in vitro*, and are predicted to affect transfusion effectiveness (16). However, translating findings from *in vitro* studies of blood donors and component manufacturing practices to clinically-relevant outcomes in transfused patients is challenging (17, 18). Thus, in this retrospective study, we utilized a large linked “vein-to-vein” database to identify predictors of transfusion effectiveness following single RBC unit transfusions. In a cohort of approximately 50,000 evaluable patients across 12 academic and community hospitals participating in the National Heart Lung and Blood Institute (NHLBI) Recipient Epidemiology and Donor Evaluation Study-III (REDS-III), we examined associations between donor, component, and recipient characteristics, including donor genetic polymorphisms, and RBC transfusion effectiveness.

RESULTS

Number of transfusion events and demographics

We identified 46,705 and 5,354 patients, with 102,043 and 6,168 evaluable single RBC unit transfusion episodes, in the full cohort and the RBC-Omics study subset (in which donor genetic polymorphism information was available) (19), respectively, which allowed for examining the hemoglobin increment as an outcome (Figure 1). In addition, there were 10,899 patients with 19,205 single RBC unit transfusion episodes and 32,416 patients with 64,051 single RBC unit transfusion episodes in whom bilirubin and creatinine increments were examined, respectively. The RBC-Omics study subset was similar to the full cohort except for race, because non-white donors were preferentially recruited in the REDS-III RBC-Omics study (e.g., mean of 2.8% black donors in the full cohort and 6.8% in the RBC-Omics subset; Table 1). Furthermore, RBC units transfused from the RBC-Omics subset were more likely to be from whole blood rather than apheresis donations as compared to the full cohort (91.2% vs 76.6% by whole blood). Recipients with bilirubin measures available were more likely than those with hemoglobin measures available to receive irradiated RBCs (52.2% vs 24.2%), concomitant plasma (6.5% vs 4.0%), and platelets (13.0% vs 5.9%). Finally, recipients with creatinine measures available were very similar to those with hemoglobin measures (Table 1).

Bivariate associations confirm factors associated with hemoglobin and bilirubin increment

The mean \pm SD pre-transfusion hemoglobin, bilirubin, and creatinine levels were 7.35 ± 0.84 g/dL, 0.67 ± 0.34 mg/dL, and 0.86 ± 0.29 mg/dL and were measured at 4 (IQR, 2-

8), 8 (IQR, 4-15), and 6 (IQR, 3-10) hours before transfusion, respectively (Table 1). The post-transfusion hemoglobin, bilirubin, and creatinine levels were measured at 21 (IQR, 18-25), 15 (8-19), and 67 (IQR, 61-72) hours after transfusion, respectively. The mean \pm SD hemoglobin, bilirubin, and creatinine increments were 0.96 ± 0.84 g/dL, 0.19 ± 0.44 mg/dL, and -0.01 ± 0.29 mg/dL, respectively. The timing and hemoglobin levels in the RBC-Omics study subset closely paralleled those of the full hemoglobin cohort. The bivariate analyses of blood donor, component, and recipient characteristics (Supplementary Table 1) were mostly consistent with prior studies (1), with recipient characteristics such as sex and BMI having strong associations with hemoglobin increments. Donor race and ethnicity were not associated with hemoglobin or bilirubin increments. Significant associations were consistent between the full hemoglobin cohort and the RBC-Omics study subset.

Because it is expected that prolonged refrigerator storage prior to transfusion is associated with an approximately 10% decrease in post-transfusion RBC recovery on average (9), we hypothesized that storage duration would decrease the hemoglobin increment; and indeed, interpolation from the parameter estimates of the bivariate analysis (Supplementary Table 1) confirmed a 8% decrease in hemoglobin increment by the end of the 6-week allowable storage duration. Furthermore, prior studies suggested that extravascular hemolysis following clearance of the storage-damaged RBCs would result in increased circulating bilirubin levels (2). Bivariate analysis supported this finding with a ~ 0.03 mg/dL observed increase in bilirubin increment per week of storage (Supplementary Table 1). No donor factors were associated with the change in creatinine following transfusion. In addition, the parameter estimates for

component and recipient factors associated with the creatinine change following transfusion were small and clinically insignificant. Sensitivity analyses examining relative rather than absolute changes in creatinine levels after transfusion yielded similar findings (results not shown). Thus, we concluded that single RBC transfusions have limited effect on creatinine levels, and multivariable models exploring change in creatinine as an outcome are not reported.

Multivariable analysis suggests that increased storage duration is associated with hemolysis

On multivariable analysis, donor factors predicted to increase the amount of hemoglobin in a unit (e.g., male donor, increased donor hemoglobin, tobacco use; Table 2) (1, 20) were associated with both increased hemoglobin and bilirubin increments. Furthermore, component characteristics associated with decreased amounts of hemoglobin in a unit (e.g., apheresis collection and leukoreduction) (21, 22) were associated with both decreased hemoglobin and bilirubin increments. In contrast, longer RBC storage duration was associated with decreased hemoglobin increment but increased bilirubin increment (Figure 2), suggesting that storage-damaged RBCs are cleared from the circulation and hemoglobin derived from these lysed RBCs is catabolized to bilirubin. In addition, there was a significant interaction term between storage duration and irradiation such that irradiation was associated with a negative effect on hemoglobin increment after 19 days of storage. Finally, recipient factors associated with increased blood volume (e.g., male gender and BMI) were associated with a decreased hemoglobin increment. Other recipient demographic factors (e.g., age, race, receipt of

concomitant platelets or plasma, issue location) were associated with both hemoglobin and bilirubin increments.

RBC units from donors with G6PD-deficiency are associated with decreased hemoglobin increment

Given that a subset of the blood donors in our linked database had associated genotype data from the REDS-III RBC-Omics Study (Figure 1), we examined SNPs identified in that study that were associated with oxidative and osmotic hemolysis of stored RBCs *in vitro* (16). The first candidate SNP explored was in the *G6PD* gene. A prior study suggests that, based on current FDA criteria, RBCs from *G6PD*-deficient donors would not meet the requirements for suitable storage quality and are associated with a 6.8% reduction in post-transfusion recovery by the end of the allowable storage period, as compared to RBCs from donors with the wild type *G6PD* allele (12). Thus, to validate the models, we examined *G6PD*-deficiency in the linked database with respect to hemoglobin increment. Compared to the wild-type allele (n=3458), donor *G6PD*-deficiency (n=43) in male donors was strongly associated with a decreased mean \pm SD hemoglobin increment in transfused recipients (1.02 ± 0.30 g/dL vs. 0.76 ± 0.23 g/dL, $p<0.0001$; Figure 3) in our multivariable model accounting for other donor, component, and recipient factors. In parallel to prior studies (1), RBC units from *G6PD*-normal female donors exhibited a lower hemoglobin increment than *G6PD*-normal males (0.93 ± 0.30 g/dL vs. 1.02 ± 0.30 g/dL, $p<0.0001$; Figure 3). Smaller hemoglobin increments associated with RBC units from female donors heterozygous for *G6PD*-deficiency (n=46) compared to those with wild-type alleles (n=2584) were also statistically significant (0.81 ± 0.35 g/dL vs. 0.93 ± 0.30 g/dL, $p=0.04$; Figure 3). No relatively rare

female homozygous *G6PD*-deficient donors were enrolled in the RBC-Omics study. Bilirubin changes were not analyzed due to the insufficient number of transfusion episodes in the bilirubin model when incorporating only units from donors participating in the RBC-Omics Study. Finally, relative to RBC units from wild-type allele donors, RBC units from *G6PD*-deficient donors were associated with increased incidence (OR 2.35 [95% CI 1.16, 4.76]; p=0.017; n=43) and number of additional RBC transfusions (1.14 [95% CI 0.75, 1.52] vs. 0.69 [95% CI 0.65, 0.73] RBC units; p=0.03) within 48 hours of the index transfusion event.

Other donor SNPs are associated with hemoglobin increments in recipients

We explored the effects of other donor SNPs on transfusion effectiveness, as defined by the recipient hemoglobin increment (bilirubin increment models were not considered due to sample size limitations), in our multivariable model adjusting for donor, component, and recipient factors. We first compared allele frequency in all RBC-Omics donors with that of donors represented in our linked database to test whether there was selection bias (Supplementary Table 2); the relative prevalence of minor alleles was similar for the SNPs in all RBC-Omics donors with those present in single RBC unit transfusion episodes. Of the 22 genes associated with osmotic or oxidative hemolysis in the RBC-Omics study (Supplementary Table 3), those that were significantly associated with the hemoglobin increment in our study were *SEC14 Like 4* (*SEC14L4*), *Myosin IXB* (*MYO9B*) and *Hemoglobin, Alpha 2* (*HBA2*) (Table 3). Although mutations in *G6PD* and *HBA2* are associated with anemia and human disease (15, 23), *SEC14L4* and *MYO9B* are not previously known to affect RBC function. Relative to RBC-units from homozygous dominant donors, RBC units from homozygous recessive donors were

associated with increased incidence in the receipt of additional RBC transfusions within 48 hours of the index transfusion event for *SEC14L4* (OR 1.54 [95% CI 1.03, 2.30]; p=0.035; n=247), but not for *HBA2* (OR 2.46 [95% CI 0.40, 15.2]; p=0.34; n=10) or *MYO9B* (OR 1.13 [95% CI 0.94, 1.35]; p=0.20; n=1,113). In parallel, the number of additional RBC transfusions for recipients of index units from homozygous recessive donors compared to homozygous dominant donors was increased for *SEC14L4* (0.95 [95% CI 0.72, 1.17] vs. 0.70 [95% CI 0.64, 0.75] RBC units; p=0.03), but not for *HBA2* (1.12 [95% CI -0.15, 2.40] vs. 0.65 [95% CI 0.61, 0.69] RBC units; p=0.46) or *MYO9B* (0.60 [95% CI 0.51, 0.70] vs. 0.62 [95% CI 0.55, 0.69] RBC units; p=0.76).

Recognizing that blood component modifications have been associated with increased rates of hemolysis (1, 24, 25), we additionally examined the association of unit irradiation in the subset of SNPs associated with *in vitro* hemolysis. Of those, four genes were associated with significant decrements in hemoglobin levels for the homozygous recessive allele for only irradiated units (Supplementary Table 4). Among irradiated units, recessive alleles for *SEC14L4* again correlated with reduced hemoglobin increments as did SNPs for *Hexokinase 1 (HK1)*, for which mutations are associated with a non-spherocytic hemolytic anemia (26), and *Aquaporin 1 (AQP1)*, an integral membrane protein regulating RBC volume in response to tonicity (27). Given the relative frequency of the homozygous recessive alleles for *SEC14L4* and *MYO9B* in blood donors and transfused units, we examined our primary model covariates stratifying by SNP allele for each of these two genes and found that donor, component, and patient characteristics were similar for recipients of single unit RBC transfusions,

thereby supporting a true effect of these gene variants rather than confounding (Supplementary Tables 5a and 5b).

DISCUSSION

We examined the role of blood donor, component, and recipient characteristics, including donor genetic polymorphisms, on three laboratory parameters that we defined as measures of RBC transfusion effectiveness. In this large, linked, “vein-to-vein” database, we first examined previously described donor, component, and recipient factors associated with increments in hemoglobin (1, 20) and bilirubin (2, 11) to establish the validity of our models and to determine covariates that should be included in the final multivariable model. Validated by other studies (1, 2, 11, 20), hemoglobin and bilirubin increments correlated positively with factors associated with increased RBC unit mass. In contrast, prolonged RBC storage was associated with increased bilirubin and reduced hemoglobin increments in transfused patients, findings indicative of hemolysis (1, 2, 11). We did not observe any clinically significant associations in the creatinine increment following transfusion, suggesting that variability in transfused RBC products have limited impact on this gross measure of kidney function. We next sought to examine novel associations between donor SNPs associated with *in vitro* hemolysis measures on our outcomes of transfusion effectiveness. Given prior evidence supporting a negative effect of *G6PD* deficiency on post-transfusion recovery (12) and transfusion effectiveness in patients with sickle cell disease (28), we first examined *G6PD* deficiency in our more limited RBC-Omics Study cohort. Transfusion of single RBC units from donors with *G6PD* deficiency not only decreased the hemoglobin

increment but was also associated with an increased need for subsequent transfusion in the ensuing 48 hours. Thus, this represents the first demonstration that transfusion of blood from donors with *G6PD* deficiency has a clinical impact. We extended these results to additional novel SNPs and discovered that polymorphisms in the *SEC14L4*, *MYO9B*, and *HBA2* genes are associated with a reduced hemoglobin increment. The potential clinical relevance of these findings is highlighted by the significant increase in the need for additional RBC transfusions within 48 hours of the index transfusion event for single unit RBC transfusions from donors with a polymorphism in *SEC14L4*. Thus, these novel findings may ultimately lead to improved precision patient blood management approaches for decreasing RBC transfusions in vulnerable patient populations.

Allogeneic RBC units are complex biological products prepared from donated blood with significant variability in hemoglobin dose and quality. In parallel to other recent studies, we identified changes in the hemoglobin increment related to individual blood donor, component manufacturing, and recipient factors (1, 29). Some of these differences are due to differences in the RBC mass of the collected blood. For example, males have higher hemoglobin levels than females; in this study both male sex and higher donor hemoglobin levels at collection were associated with a larger hemoglobin increment. In contrast, manufacturing practices such as apheresis collection and leukoreduction are associated with a smaller hemoglobin increment due to reductions in overall RBC mass collected (for apheresis RBC) or due to loss of RBC during component processing (due to leukoreduction) (21, 22). Although some donor and component characteristics individually played a small role in hemoglobin

increments, they remained relevant to the variation observed in clinical practice even after multivariable adjustment. For example, an irradiated unit of RBCs from a male donor with G6PD deficiency stored for 42 days would be associated with a median hemoglobin increment that is 0.51 g/dL lower than a non-irradiated unit from a male donor without G6PD deficiency transfused on the first day of storage. Assuming an average single unit transfusion is associated with a 1 g/dL increase in hemoglobin, this represents an approximately 50% decrease in the effective dose of circulating hemoglobin.

Although not previously demonstrated, findings of both increased bilirubin and hemoglobin increments (e.g., with transfusion of male-derived or whole blood RBC units) are not unexpected and likely represent clearance of a larger mass of damaged RBCs in units with higher doses of hemoglobin. RBC storage duration was the exception to the aforementioned correlation, with prolonged storage being strongly associated with a *reduced* hemoglobin and an *increased* bilirubin increment. This particular relationship was previously demonstrated in healthy volunteers transfused with autologous RBC units (2). Although several randomized controlled trials (30-34) did not observe significant improvements in outcomes of patient who received fresher RBC transfusions, it is increasingly recognized that, due to significant donor variability, the chronological storage age of RBC units does not necessarily indicate the metabolic age of the RBCs transfused (35). This is the first retrospective study to demonstrate an effect of storage age on post-transfusion bilirubin levels, and this approach may be useful for identifying patient populations and clinical outcomes that may be affected by hemolysis from transfused RBCs. Indeed, because increases in bilirubin correlate with

other markers of hemolysis, such as serum iron and non-transferrin bound iron (2), these findings may be particularly relevant for chronically transfused patients for whom overall iron burden results in harm (36-38).

The REDS-III RBC-Omics Study identified multiple novel donor genetic polymorphisms associated with hemolysis *in vitro* after prolonged storage (16). Many of the SNPs identified were in genes associated with hemoglobinopathies, enzymopathies, and membranopathies, which could plausibly affect RBC recovery following storage and transfusion. Due to sample size limitations, we focused our analyses on the hemoglobin increment as the main outcome measure for the genetic variants identified in the RBC-Omics Study. Using multivariable analyses, we found a smaller hemoglobin increment in two of the five genome-wide significant loci for oxidative hemolysis: *G6PD* and *SEC14L4*. Out of eighteen loci for osmotic hemolysis examined for decreased hemoglobin increment, we observed two significant associations: *HBA2* and *MYO9B*. Although SNPs in *G6PD* and *HBA2* are associated with human disease, clinical phenotypes for *SEC14L4* and *MYO9B* are not yet described. A sensitivity analysis of relevant donor, component, and patient characteristics did not identify any obvious differences between the alleles of these two novel SNPs (Supplementary Tables 5a and 5b). Interestingly, and requiring further validation, transfusion of RBC units from *G6PD* heterozygous female donors was associated with a decreased hemoglobin increment, suggesting that female heterozygotes may have an intermediary phenotype for oxidative hemolysis both *in vitro* and *in vivo*. In parallel with decreased hemoglobin increments, we found an increase in the likelihood and number of subsequent transfusions in patients receiving a single RBC unit from donors with polymorphisms in

G6PD or *SEC14L4*. Although other clinical consequences of these SNPs are unclear and will require additional mechanistic investigations, it is notable that the above four SNPs were among seven genetic variants that correlated with *in vivo* hemolysis in a study in sickle cell patients (16).

Additional factors may further influence the impact of donor genetic polymorphisms associated with oxidative or osmotic hemolysis. Several studies found that manufacturing method, gamma irradiation, and storage solution influence hemolysis rates. For example, gamma irradiation increases plasma free hemoglobin in the supernatant of RBC units by disrupting RBC membrane integrity, which results in decreased RBC recovery following transfusion, particularly after longer storage durations (24, 25, 39-42). Component modifications may further increase hemolysis in RBC units from donors with particular genetic variants. Thus, we hypothesized that donor genetic polymorphisms and component factors may have “multi-hit” interactions on RBC quality. In this setting, the first “hit” is a particular donor genetic polymorphism and the second “hit” is gamma irradiation (Supplementary Table 4). Because recessive alleles for *G6PD* or *HBA2* are relatively infrequent in this donor population, these could not be examined robustly. However, several SNPs, including the homozygous recessive allele for *SEC14L4*, were associated with a decreased hemoglobin increment following transfusion of irradiated RBC units. Although speculative, a cumulative effect of donor genetics and component modifications on RBC quality align with the proposed concept of the “metabolic age” of RBCs (35), which may vary significantly from changes expected with chronological storage age alone.

The current study has several limitations. The retrospective and observational nature of the study design limits the ability to infer causation for the factors associated with decreased transfusion effectiveness. Nonetheless, the models confirmed previous prospective, randomized study findings of decreased hemoglobin, and increased bilirubin increments associated with increased storage duration (2). Furthermore, RBCs from donors with *G6PD* deficiency were associated with decreased post-transfusion recovery in prospective studies (12, 28, 43). Thus, our statistical models were able to validate these prior associations and this lends support to the validity of the analytic approach. However, given the number of variables examined, the possibility of a chance result (Type I error) for any given association exists and each association will require further validation. In particular, given the lack of adjustment for a false discovery rate and multiple comparisons, the SNPs in *SEC14L4*, *MYO9B*, and *HBA2* require further confirmation as their effects on transfusion effectiveness have yet to be explored in other studies. In addition, subjects with evaluable pre- and post-transfusion bilirubin levels were clearly different from those with just evaluable hemoglobin levels (e.g., they were more likely to receive irradiated blood products and concomitant plasma and/or platelet units). These subjects were also more likely to be recipients of multi-unit RBC episodes where bleeding and additional intravenous fluids may have impacted our outcomes. In parallel with other characteristics, uncommon SNPs are also less likely to impact hemoglobin increment when occurring in multi-unit transfusion events; nevertheless, they contribute to the significant variation in hemoglobin increments observed in clinical practice in conjunction with other donor and component factors. Future studies of multi-unit transfusion events are needed in a controlled setting

including the role of platelet and plasma transfusions to demonstrate the generalizability of our findings. Although we limited analysis to pre-transfusion bilirubin levels below 1.5 mg/dL to mitigate the role of patient conditions (e.g., liver disease) from affecting the results, our findings may still be prone to unmeasured confounding. However, sensitivity analyses examining relative changes in creatinine levels and various pre-transfusion thresholds for our outcomes did not yield different results. Due to sample size limitations, our analysis was underpowered to identify associations of uncommon donor alleles which had small effect sizes; nor were we able to examine the impact of donor genetics on post-transfusion bilirubin increments. To this end, the NIH-funded Recipient Epidemiology and Donor Evaluation-IV Pediatrics (REDS-IV-P) program is planning a prospective trial in chronically transfused patients with sickle cell disease, thalassemia, or pediatric cancer to confirm and expand upon the genetic determinants of transfusion effectiveness (Improving Transfusions for Chronically Transfused Recipients; RBC-IMPACT Study). Finally, despite the associations of donor, component, and recipient variables with decreased transfusion effectiveness, as defined by a decreased hemoglobin increment or an increased bilirubin increment demonstrated by this study, we did not explore whether decreased transfusion effectiveness is associated with patient harm other than an increased need for subsequent transfusion. Our current analyses were limited to single unit episodes, and potential harmful effects may be greater when accounting for cumulative transfusion exposures. A recent study found that a limited set of donor and manufacturing characteristics correlated with smaller hemoglobin increments, which were then associated with additional RBC transfusion requirements in the patients (20). We similarly observed an increased odds

of requiring a subsequent transfusion in patients transfused a single RBC unit from a donor with polymorphisms in *G6PD* or *SEC14L4*. Nonetheless, we believe that future studies focusing on specific patient populations (e.g., chronically transfused patients with hemoglobinopathies) and specific outcome measures (e.g., iron overload) can better address the issue of potential harm.

In conclusion, we found significant associations between blood donor (e.g., sex, Rh status, fingerstick hemoglobin, tobacco use), component (e.g., storage duration, gamma irradiation, leukoreduction, apheresis collection, storage solution), and recipient (e.g., sex, race, BMI, age) characteristics, including specific donor genetic polymorphisms in four genes (e.g., *G6PD*, *SEC14L4*, *HBA2*, and *MYO9B*), and effectiveness measures following single-unit RBC transfusion events. The strong associations between increased storage duration and donor G6PD deficiency on transfusion effectiveness support the validity of this approach for identifying factors that influence transfusion effectiveness. These findings highlight the power of large linked “vein-to-vein” databases that link blood donor and recipient outcome information. Furthermore, the current models expand upon previous models examining variables affecting the hemoglobin increment (1) and are the first to describe the impact of donor, component, and recipient factors on bilirubin increment following transfusion, a laboratory measure of hemolysis *in vivo*. These findings support the concept that donor factors, including genetic variables, can modulate RBC stability in storage, and could provide relevant donor selection criteria for improving transfusion outcomes in selected populations. For example, many blood donors are already genotyped and phenotypically-matched blood is targeted to select populations to prevent

alloimmunization (44). In similar fashion, a precision transfusion medicine approach may benefit chronically-transfused patients, who may be at increased risk of harm from transfusion of RBCs from donors with G6PD deficiency or other SNPs (28, 45). Thus, identifying genetically-susceptible RBC units, which may result in suboptimal transfusion responses, or conversely RBC units that better tolerate cold storage, could provide a strategy more advanced than the current approach using a random distribution of donors and RBC products irrespective of donor genotype or short- and long-term transfusion requirements of the recipient.

METHODS

General Study Design

We conducted a retrospective cohort study using electronic health records from the NHLBI REDS-III program available as public use data through BioLINCC (46, 47). The database included blood donor, component, and recipient data collected at 12 academic and community hospitals from four geographically dispersed regions in the United States (Connecticut, Pennsylvania, Wisconsin, and California) for the 4-year period from January 1, 2013 to December 31, 2016. Genotype data from the subset of blood donors who participated in the REDS-III RBC-Omics study (19) were included as part of the linked dataset.

Study Population and Definitions

Data regarding blood donor demographics (e.g., age, sex, race, ethnicity, ABO/Rh), collection date and methods (e.g., whole blood or apheresis), as well as component manufacturing characteristics (e.g., timing of leukoreduction, additive solution, irradiation status) were extracted for each RBC unit collected. Available donor genetic polymorphism data from the REDS-III RBC-Omics study (19) was linked to issued RBC units using donor identification numbers. Among transfusion recipients, we included all adult patients who received a single RBC unit during one or more transfusion episodes between January 1, 2013 and December 30, 2016. Recipient details included age, sex, race, ethnicity, and body mass index (BMI), along with storage age, and blood product issue date, time, and location of transfused RBCs. Information on concomitant transfusion of other blood components (e.g., platelets, plasma) during the RBC transfusion episode was included. We collected hemoglobin, creatinine, and bilirubin

levels measured by the corresponding clinical laboratory prior to and following each RBC transfusion event.

Transfusion Exposures and Outcome Measures

All single RBC unit transfusion episodes were included in this analysis. A RBC unit transfusion episode was defined as any single RBC transfusion from a single donor with both informative pre- and post-transfusion laboratory measures and without any other RBC units transfused in the intervening period between lab draws. The three outcome measures of interest were hemoglobin, total bilirubin, and creatinine increments following a single RBC unit transfusion episode. These outcomes were defined as the difference between the post-transfusion and pre-transfusion levels. Pre-transfusion thresholds for these measures were chosen to limit patient confounding (e.g., underlying hepatic or renal disease). For pre-transfusion hemoglobin, the value used was the most proximal hemoglobin measurements prior to RBC transfusion, but at most 24 hours prior to transfusion. For post-transfusion hemoglobin (g/dL), the laboratory measure nearest to 24-hours post-transfusion, but between 12- and 36-hours following transfusion was based on prior studies. For pre-transfusion bilirubin (mg/dL), the most proximal total bilirubin measurement prior to transfusion, but at most seven days prior to transfusion was used. Furthermore, to exclude patients with pre-existing liver disease, the pre-transfusion bilirubin had to be below 1.5 mg/dL for the transfusion to be considered. For post-transfusion bilirubin, the most proximal laboratory measure to the transfusion, but at most 24-hours post-transfusion was used based on clinical trial data (2, 11, 32). For pre-transfusion creatinine (mg/dL), the most proximal measurement

prior to the transfusion time, but at most 24-hours prior to transfusion was used. Furthermore, to exclude patients with pre-existing kidney disease, the pre-transfusion creatinine had to be below 1.5 mg/dL for the transfusion to be considered. Finally, to assess for changes in kidney function, the maximum creatinine level most proximal to 72-hours post-transfusion, but at most 96 hours post-transfusion was derived from acute kidney injury criteria (48).

Statistical Methods

We assessed the univariable association of all *a priori* selected donor, component, and recipient covariates with the outcomes using linear regression. Any covariate that was associated ($\alpha = 0.2$) with the outcome was included in a multivariable linear mixed effects model accounting for repeated transfusion recipients. Additional *a priori* selected interaction terms were included in the multivariable models based on hypotheses of modifiers of clinical outcomes (e.g., gamma irradiation * storage duration). Cook's D was used to detect outliers (threshold < 0.1) and stepwise model selection on the fixed effects ($\alpha = 0.05$) was used to select the final full covariate model for each outcome. Initial models were fit on the subset of the data that had no missing observations for all covariates. Once the final model covariates were selected, the model including just those covariates was reassessed on the larger data set, excluding observations with missing values for only the selected covariates to maximize sample size. We tested for multicollinearity in the final models using generalized variance inflation factor values adjusted for the number of regressors as well as performing additional outlier assessment. Fixed marginal effects from multivariable regression models were used to

correlate delta hemoglobin and bilirubin levels by week of storage duration. For the subset of recipients of RBC units from REDS-III RBC-Omics donors (19), we examined alleles of donor genetic polymorphisms associated with measures of osmotic or oxidative hemolysis *in vitro*. Accounting for the same donor, component, and recipient factors, multivariable linear regression assessed associations between alleles of donor single nucleotide polymorphisms (SNPs) and the post-transfusion hemoglobin increment. For donor SNPs associated with the post-transfusion hemoglobin increment, we further examined the incidence and number of subsequent RBC transfusions occurring within 48 hours of the index transfusion event using multivariable logistic regression. Sample size estimates for different allele frequencies and effect sizes were calculated (Supplementary Table 6). Two-sided p-values less than 0.05 were considered to be statistically significant. Analyses were performed in R 4.0.3 (R Core Team, 2020)(49, 50) and Stata Version 14.1 (StataCorp, College Station, TX). R package lme4 was used to fit linear mixed effects models (49).

Study approval

Approval for data collection had been obtained from the Institutional Review Board at each participating institution.

Conflict of interest: The authors have no relevant conflicts of interest to disclose.

Author Contributions:

NHR, SR, RC, BC, RG, SK, AM, SS, BS, SAS MPB, EAH conceived of and designed the study. NHR, SR, HQ, CP, FF, GP, BC, TK, SK, AM, BS, MPB, EAH acquired and prepared the data. NHR, SR, CP, FF, GP, EAH performed statistical analyses. NHR, SR, FF, GP, RC, BC, MG, RG, BH, JH, TK, SK, AM, SS, BS, SAS, MPB, EAH interpreted data. NHR, SR, EAH wrote the manuscript. All authors contributed to the critical revision of the manuscript for important intellectual content.

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REFERENCES

1. Roubinian NH, Plimier C, Woo JP, Lee C, Bruhn R, Liu VX, Escobar GJ, Kleinman SH, Triulzi DJ, Murphy EL, et al. Effect of donor, component, and recipient characteristics on hemoglobin increments following red blood cell transfusion. *Blood*. 2019;134(13):1003-13.
2. Rapido F, Brittenham GM, Bandyopadhyay S, La Carpia F, L'Acqua C, McMahon DJ, Rebbaa A, Wojczyk BS, Netterwald J, Wang H, et al. Prolonged red cell storage before transfusion increases extravascular hemolysis. *J Clin Invest*. 2017;127(1):375-82.
3. Zimring JC. Established and theoretical factors to consider in assessing the red cell storage lesion. *Blood*. 2015;125(14):2185-90.
4. Glynn SA, Klein HG, and Ness PM. The red blood cell storage lesion: the end of the beginning. *Transfusion*. 2016;56(6):1462-8.
5. Roussel C, Morel A, Dussiot M, Marin M, Colard M, Fricot-Monsinjon A, Martinez A, Chambrion C, Henry B, Casimir M, et al. Rapid clearance of storage-induced microerythrocytes alters transfusion recovery. *Blood*. 2021;137(17):2285-98.
6. Francis RO, Mahajan S, Rapido F, La Carpia F, Soffing M, Divgi C, Yeh R, Mintz A, Leslie L, Agrest I, et al. Reexamination of the chromium-51-labeled posttransfusion red blood cell recovery method. *Transfusion*. 2019;59(7):2264-75.
7. Leung N. Hematologic manifestations of kidney disease. *Semin Hematol*. 2013;50(3):207-15.

8. Gladwin MT, Kanas T, and Kim-Shapiro DB. Hemolysis and cell-free hemoglobin drive an intrinsic mechanism for human disease. *J Clin Invest.* 2012;122(4):1205-8.
9. Dumont LJ, and AuBuchon JP. Evaluation of proposed FDA criteria for the evaluation of radiolabeled red cell recovery trials. *Transfusion.* 2008;48(6):1053-60.
10. Dern RJ, Gwinn RP, and Wiorkowski JJ. Studies on the preservation of human blood. I. Variability in erythrocyte storage characteristics among healthy donors. *J Lab Clin Med.* 1966;67(6):955-65.
11. Hod EA, Brittenham GM, Billote GB, Francis RO, Ginzburg YZ, Hendrickson JE, Jhang J, Schwartz J, Sharma S, Sheth S, et al. Transfusion of human volunteers with older, stored red blood cells produces extravascular hemolysis and circulating non-transferrin-bound iron. *Blood.* 2011;118(25):6675-82.
12. Francis RO, D'Alessandro A, Eisenberger A, Soffing M, Yeh R, Coronel E, Sheikh A, Rapido F, La Carpia F, Reisz JA, et al. Donor glucose-6-phosphate dehydrogenase deficiency decreases blood quality for transfusion. *J Clin Invest.* 2020;130(5):2270-85.
13. Spitalnik SL, Triulzi D, Devine DV, Dzik WH, Eder AF, Gernsheimer T, Josephson CD, Kor DJ, Luban NL, Roubinian NH, et al. 2015 proceedings of the National Heart, Lung, and Blood Institute's State of the Science in Transfusion Medicine symposium. *Transfusion.* 2015;55(9):2282-90.

14. Nkhoma ET, Poole C, Vannappagari V, Hall SA, and Beutler E. The global prevalence of glucose-6-phosphate dehydrogenase deficiency: a systematic review and meta-analysis. *Blood Cells Mol Dis.* 2009;42(3):267-78.
15. Luzzatto L, Ally M, and Notaro R. Glucose-6-phosphate dehydrogenase deficiency. *Blood.* 2020;136(11):1225-40.
16. Page GP, Kanas T, Guo YJ, Lanteri MC, Zhang X, Mast AE, Cable RG, Spencer BR, Kiss JE, Fang F, et al. Multi-ancestry genome-wide association study identifies 27 loci associated with erythrocyte osmotic fragility, oxidative or spontaneous hemolysis following blood storage. *Journal of Clinical Investigation.* 2021;In Press.()
17. Roubinian N, Kleinman S, Murphy EL, Glynn SA, and Edgren G. Methodological considerations for linked blood donor-component-recipient analyses in transfusion medicine research. *ISBT Sci Ser.* 2020;15(1):185-93.
18. Ning S, Heddle NM, and Acker JP. Exploring donor and product factors and their impact on red cell post-transfusion outcomes. *Transfus Med Rev.* 2018;32(1):28-35.
19. Endres-Dighe SM, Guo Y, Kanas T, Lanteri M, Stone M, Spencer B, Cable RG, Kiss JE, Kleinman S, Gladwin MT, et al. Blood, sweat, and tears: Red Blood Cell-Omics study objectives, design, and recruitment activities. *Transfusion.* 2019;59(1):46-56.
20. DeSimone RA, Plimier C, Lee C, Kanas T, Cushing MM, Sachais BS, Kleinman S, Busch MP, and Roubinian NH. Additive effects of blood donor smoking and

gamma irradiation on outcome measures of red blood cell transfusion. *Transfusion*. 2020;60(6):1175-82.

21. AuBuchon JP, Elfath MD, Popovsky MA, Stromberg RR, Pickard C, Herschel L, Whitley P, McNeil D, Arnold N, and O'Connor JL. Evaluation of a new prestorage leukoreduction filter for red blood cell units. *Vox Sang*. 1997;72(2):101-6.
22. Sweeney JD. Standardization of the red cell product. *Transfus Apher Sci*. 2006;34(2):213-8.
23. Angastinotis M, and Lobitz S. Thalassemias: An Overview. *Int J Neonatal Screen*. 2019;5(1):16.
24. Davey RJ, McCoy NC, Yu M, Sullivan JA, Spiegel DM, and Leitman SF. The effect of prestorage irradiation on posttransfusion red cell survival. *Transfusion*. 1992;32(6):525-8.
25. Moroff G, Holme S, AuBuchon JP, Heaton WA, Sweeney JD, and Friedman LI. Viability and in vitro properties of AS-1 red cells after gamma irradiation. *Transfusion*. 1999;39(2):128-34.
26. Keitt AS. Hemolytic anemia with impaired hexokinase activity. *J Clin Invest*. 1969;48(11):1997-2007.
27. Crisp RL, Maltaneri RE, Vittori DC, Solari L, Gammella D, Schwartzman G, Garcia E, Rapetti MC, Donato H, and Nesse A. Red blood cell aquaporin-1 expression is decreased in hereditary spherocytosis. *Ann Hematol*. 2016;95(10):1595-601.
28. Sagiv E, Fasano RM, Luban NLC, Josephson CD, Stowell SR, Roback JD, Francis RO, and Yee MEM. Glucose-6-phosphate-dehydrogenase deficient red

blood cell units are associated with decreased posttransfusion red blood cell survival in children with sickle cell disease. *Am J Hematol.* 2018;93(5):630-4.

29. Karafin MS, Bruhn R, Roubinian NH, Chowdhury D, Qu L, Snyder EL, Murphy EL, Brambilla D, Cable RG, Hilton JF, et al. The impact of recipient factors on the lower-than-expected hemoglobin increment in transfused outpatients with hematologic diseases. *Transfusion.* 2019;59(8):2544-50.

30. Lacroix J, Hebert PC, Fergusson DA, Tinmouth A, Cook DJ, Marshall JC, Clayton L, McIntyre L, Callum J, Turgeon AF, et al. Age of transfused blood in critically ill adults. *N Engl J Med.* 2015;372(15):1410-8.

31. Spinella PC, Tucci M, Fergusson DA, Lacroix J, Hebert PC, Leteurtre S, Schechtman KB, Doctor A, Berg RA, Bockelmann T, et al. Effect of Fresh vs Standard-issue Red Blood Cell Transfusions on Multiple Organ Dysfunction Syndrome in Critically Ill Pediatric Patients: A Randomized Clinical Trial. *JAMA.* 2019;322(22):2179-90.

32. Steiner ME, Ness PM, Assmann SF, Triulzi DJ, Sloan SR, Delaney M, Granger S, Bennett-Guerrero E, Blajchman MA, Scavo V, et al. Effects of red-cell storage duration on patients undergoing cardiac surgery. *N Engl J Med.* 2015;372(15):1419-29.

33. Dhabangi A, Ainomugisha B, Cserti-Gazdewich C, Ddungu H, Kyeyune D, Musisi E, Opoka R, Stowell CP, and Dzik WH. Effect of Transfusion of Red Blood Cells With Longer vs Shorter Storage Duration on Elevated Blood Lactate Levels in Children With Severe Anemia: The TOTAL Randomized Clinical Trial. *JAMA.* 2015;314(23):2514-23.

34. Heddle NM, Cook RJ, Arnold DM, Liu Y, Barty R, Crowther MA, Devereaux PJ, Hirsh J, Warkentin TE, Webert KE, et al. Effect of Short-Term vs. Long-Term Blood Storage on Mortality after Transfusion. *N Engl J Med.* 2016;375(20):1937-45.

35. D'Alessandro A, Zimring JC, and Busch M. Chronological storage age and metabolic age of stored red blood cells: are they the same? *Transfusion.* 2019;59(5):1620-3.

36. Piga A, Longo F, Duca L, Roggero S, Vinciguerra T, Calabrese R, Hershko C, and Cappellini MD. High nontransferrin bound iron levels and heart disease in thalassemia major. *Am J Hematol.* 2009;84(1):29-33.

37. Brittenham GM, Cohen AR, McLaren CE, Martin MB, Griffith PM, Nienhuis AW, Young NS, Allen CJ, Farrell DE, and Harris JW. Hepatic iron stores and plasma ferritin concentration in patients with sickle cell anemia and thalassemia major. *Am J Hematol.* 1993;42(1):81-5.

38. Adamkiewicz TV, Abboud MR, Paley C, Olivieri N, Kirby-Allen M, Vichinsky E, Casella JF, Alvarez OA, Barredo JC, Lee MT, et al. Serum ferritin level changes in children with sickle cell disease on chronic blood transfusion are nonlinear and are associated with iron load and liver injury. *Blood.* 2009;114(21):4632-8.

39. de Korte D, Thibault L, Handke W, Harm SK, Morrison A, Fitzpatrick A, Marks DC, Yi QL, Acker JP, and Biomedical Excellence for Safer Transfusion c. Timing of gamma irradiation and blood donor sex influences in vitro characteristics of red blood cells. *Transfusion.* 2018;58(4):917-26.

40. Serrano K, Chen D, Hansen AL, Levin E, Turner TR, Kurach JD, Acker JP, and Devine DV. The effect of timing of gamma-irradiation on hemolysis and potassium release in leukoreduced red cell concentrates stored in SAGM. *Vox Sang.* 2014;106(4):379-81.

41. Harm SK, Raval JS, Cramer J, Waters JH, and Yazer MH. Haemolysis and sublethal injury of RBCs after routine blood bank manipulations. *Transfus Med.* 2012;22(3):181-5.

42. Ran Q, Hao P, Xiao Y, Zhao J, Ye X, and Li Z. Effect of irradiation and/or leucocyte filtration on RBC storage lesions. *PLoS One.* 2011;6(3):e18328.

43. Orlina AR, Josephson AM, and McDonald BJ. The poststorage viability of glucose-6-phosphate dehydrogenase-deficient erythrocytes. *J Lab Clin Med.* 1970;75(6):930-6.

44. Yazdanbakhsh K, Ware RE, and Noizat-Pirenne F. Red blood cell alloimmunization in sickle cell disease: pathophysiology, risk factors, and transfusion management. *Blood.* 2012;120(3):528-37.

45. Francis RO, Jhang J, Hendrickson JE, Zimring JC, Hod EA, and Spitalnik SL. Frequency of glucose-6-phosphate dehydrogenase-deficient red blood cell units in a metropolitan transfusion service. *Transfusion.* 2013;53(3):606-11.

46. https://biolincc.ncbi.nih.gov/studies/reds_iii/.

47. Karafin MS, Bruhn R, Westlake M, Sullivan MT, Bialkowski W, Edgren G, Roubinian NH, Hauser RG, Kor DJ, Fleischmann D, et al. Demographic and epidemiologic characterization of transfusion recipients from four US regions:

evidence from the REDS-III recipient database. *Transfusion*. 2017;57(12):2903-13.

48. Emigh BJ, Sahi SL, Teal LN, Blake JC, Heron CH, Teixeira PG, Coopwood B, Cardenas TC, Trust MD, and Brown CV. Incidence and Risk Factors for Acute Kidney Injury in Severely Injured Patients Using Current Kidney Disease: Improving Global Outcomes Definitions. *J Am Coll Surg*. 2020;231(3):326-32.

49. Bates D, Maechler M, Bolker B, and Walker S. Fitting Linear Mixed-Effects Models Using lme4. *Journal of Statistical Software*. 2015;67(1):1-48.

50. . R Core Team (2020). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>. <https://www.R-project.org/>.

FIGURE LEGENDS

Figure 1. Study flow diagram. RBC = red blood cell.

Figure 2: Effect of storage age and irradiation on hemoglobin and bilirubin increments. (A) The adjusted change in total bilirubin (N=19,205) and (B) hemoglobin (N=102,043) level between pre- and post-transfusion laboratory measures is shown stratified by week of storage, as labeled. Hemoglobin and bilirubin increments were adjusted for concomitant donor, component, and recipient factors presented in Table 2 and plotted as derivatives of the regression equation. $P<0.0001$ for linear association between storage duration and change in laboratory measure for both figure panels.

Figure 3: Adjusted hemoglobin increments stratified by gender and glucose-6-phosphate dehydrogenase (G6PD) deficiency. Adjusted change in hemoglobin level between pre- and post-transfusion measures is shown stratified by male (N=3458) and female (N=2584) gender and G6PD genotype, as labeled. As G6PD is X-linked, males labeled (-) are G6PD-deficient (N=43) and females labeled (+/-) are heterozygous (N=46). Hemoglobin is adjusted for concomitant donor, component, and recipient factors **** $P<0.0001$, * $P<0.05$ for one-way ANOVA comparison as indicated. Line represents median, boxes represent 25th to 75th percentile, and whiskers represent the 5th to 95th percentile.

Table 1: Blood donor, component, and RBC transfusion recipient characteristics

	Hemoglobin	Hb (RBC-Omics)	Bilirubin	Creatinine
N (RBC transfusion episodes)	102,043	6,168	19,205	64,051
Blood Donor Characteristics				
Male sex (%)	57.0	56.1	58.2	57.5
Female sex (%)	43.0	43.9	41.8	42.5
Age in years, mean (SD)	46 (17)	52 (15)	46 (17)	45.9 (17.4)
(+) Rh status (%)	77.8	82.1	77.8	77.6
ABO status (%)				
A	35.1	37.0	34.2	35.5
B	12.4	14.3	12.4	12.1
AB	3.3	3.0	3.2	3.1
O	49.1	45.7	50.2	49.3
Race (%)				
White	86.9	80.8	87.5	87.0
Black/African American	2.8	6.8	2.5	2.7
Asian	2.4	5.0	2.4	2.4
Other	3.2	5.3	3.2	3.2
Missing Race	4.8	2.0	4.5	4.8
Ethnicity (%)				
Hispanic/Latino	8.0	9.2	7.7	8.4
Not Hispanic/Latino	89.7	90.4	90.3	89.1
Unknown/Missing Ethnicity	2.3	0.4	1.9	2.5
Donor hemoglobin in g/dL, mean (SD)	14.5 (1.3)	14.4 (1.2)	14.5 (1.3)	14.6 (1.3)
History of tobacco use (%)	13.0	13.2	13.1	12.9
Count of prior RBC donations, median (IQR)	2 (0-6)	2 (0-7)	2 (0-8)	2 (0-7)
Blood Component Characteristics				
Gamma irradiated (%)	24.2	28.4	52.2	29.6
RBC storage age – days, median (IQR)	26 (15-36)	25 (14-36)	24 (14-35)	26 (15-36)
Apheresis-derived collection (%)	23.4	8.8	21.8	23.0
Storage solution (%)				
AS-1	43.6	44.8	39.7	40.7
AS-3	50.0	47.1	52.0	50.1
Other	6.4	8.1	8.4	9.2
Leukoreduced (%)	85.0	88.1	90.9	86.2
Washed (%)	0.4	0.5	0.8	1.2
Volume reduction (%)	2.0	1.4	2.0	2.1
Transfusion Recipient Characteristics				
Male sex (%)	53.7	54.0	52.6	49.6
Female sex (%)	42.3	46.0	47.4	50.4
Age, mean (SD)	62 (16)	62.0 (16.3)	60.0 (15.5)	61.2(15.0)
Body mass index, mean (SD)	28.4 (7.3)	28.5 (7.2)	27.9 (7.0)	27.5 (7.0)
(+) Rh status (%)	85.7	88.3	85.6	85.2
ABO status (%)				
A	35.3	37.3	37.2	36.1
B	13.6	15.0	14.9	13.5
AB	5.0	4.9	4.8	4.7
O	43.3	40.1	40.6	43.7

Missing	0.9	2.7	2.4	2.0
Race (%)				
White	72.6	73.3	74.7	75.4
Black/African American	15.7	16.2	13.1	13.7
Asian	2.1	1.9	2.3	1.7
Other	1.0	0.8	0.8	0.7
Missing Race	8.6	7.8	9.1	8.5
Ethnicity (%)				
Hispanic/Latino	6.6	5.9	7.7	6.3
Concomitant plasma transfusion (%)	4.0	4.0	6.5	4.3
Concomitant platelet transfusion (%)	5.9	5.5	13.0	7.0
Issue location (%)				
Medical/surgical floor	54.9	54.9	63.6	54.9
Emergency department	4.5	4.5	4.8	4.2
Intensive care	29.9	29.9	20.8	26.5
Operating room	3.4	3.4	4.2	4.1
Outpatient	3.8	3.8	3.5	6.5
Other/missing	3.5	1.2	3.1	3.8
Count of prior RBC transfusions, median (IQR)	4 (2-9)	4 (2-10)	5 (2-11)	4 (2-10)
Pre-transfusion laboratory measure, mean (SD)	7.35 (0.84)	7.33 (0.83)	0.67 (0.34)	0.86 (0.29)
Time Pre-TX to TX in hours, median (IQR)	4 (2-9)	5 (2-8)	8 (4-15)	6 (3-10)
Time TX to Post-TX in hours, median (IQR))	21 (18-25)	21 (18-25)	15 (8-19)	67 (61-72)

RBC=red blood cell; Hb=hemoglobin; SD=standard deviation; IQR=interquartile range; TX=transfusion

Table 2: Changes in outcomes following RBC transfusion [95% CI] - Multivariate models^A

	Hemoglobin	Hemoglobin (RBC-Omics)	Bilirubin
N (transfusion episodes)	102,043	6,168	19,205
Blood Donor Characteristics			
Sex, male (ref: female)	0.05 (.04, .06)	0.04 (-.01, .10)	0.02 (.01, .04)
(+) Rh status	0.02 (.01, .04)	NS	NS
Donor hemoglobin	0.03 (.02, .03)	0.05 (0.03, 0.07)	0.01 (.00, .01)
History of tobacco use	0.03 (.01, .05)	NS	0.03 (.01, .05)
Blood Component Characteristics			
Gamma irradiated	NS	NS	NS
RBC storage age (by week)	-0.01 (-.02, -.01)	-0.01 (-.03, .01)	0.04 (.03, .05)
Gamma irradiated * RBC storage age	-0.01 (-.02, -.00)	-0.04 (-.07, -.01)	NS
Apheresis-derived	-0.10 (-.12, -.08)	-0.18 (-.28, -.09)	-0.03 (-.04, -.02)
Storage solution AS-3 (ref: AS-1)	NS	NS	NS
Leukoreduced	-0.08 (-.10, -.05)	-0.08 (-.17, -.01)	NS
Transfusion Recipient Characteristics			
Sex, Male	-0.26 (-.27, -.24)	-0.27 (-.33, -.24)	-0.02 (-.03, -.01)
Age (per decade)	0.06 (-.05, -.06)	0.05 (-.04, .07)	0.01 (.01, .02)
Body mass index by unit	-0.02 (-.02, -.01)	-0.02 (-.02, -.01)	-0.003 (-.003, -.002)
Race (ref: White)			
Black/African American	-0.07 (-.09, -.05)	-0.03 (-.09, -.04)	-0.02 (-.04, -.01)
Asian	0.09 (.03, -.14)	NS	0.06 (.02, .10)
Other	NS	NS	NS
Concomitant plasma transfusion	-0.09 (-.12, -.07)	NS	0.18 (.14, .21)
Concomitant platelet transfusion	-0.10 (-.13, -.07)	-0.12 (-.23, -.01)	0.03 (.01, .04)
Issue location (ref: medical/surgical floor)			
Emergency department	-0.13 (-.16, -.10)	NS	0.04, (.01, .06)
Intensive care	-0.03 (-.05, -.02)	0.08 (-.14, -.02)	NS
Operating room	-0.06 (-.10, -.02)	0.08 (-.14, -.02)	-0.02 (-.03, -.01)
Outpatient	-0.29 (-.32, -.26)	-0.25 (-.36, -.14)	0.17 (.14, .21)
Pre-TX laboratory measure	-0.58 (-.59, -.57)	-0.51 (-.54, -.49)	-0.03 (-.04, -.01)
Time Pre-TX to TX	0.006 (.004, .007)	0.009 (.003, .015)	-0.007 (-.008, -.006)
Time TX to Post-TX	-0.005 (-.006, -.004)	-0.007 (-.01, -.002)	-0.001 (-.002, -.001)
Count of prior RBC Tx	-0.002 (-.001, -.002)	-0.002 (-.001, -.002)	-0.002 (-.001, -.002)

RBC=red blood cell; CI=confidence interval; Hb=Hemoglobin; TX = transfusion

^ARegression estimates of the effect size for hemoglobin and bilirubin increments in g/dL and mg/dL, respectively

All presented outcomes were statistically significant (p<0.05) except where indicated (NS)

Conditional coefficients of determination for generalized mixed-effect models for hemoglobin, hemoglobin for RBC-Omics, and bilirubin outcomes were 0.54, 0.47, and 0.22, respectively.

Table 3. Hemoglobin increments after single unit RBC transfusion by SNP allele – g/dL [95% CI]

Hemolysis	Gene	Single nucleotide polymorphism	Homozygous dominant	Heterozygous dominant	Homozygous recessive ^A
Oxidative	<i>G6PD</i>	rs1050828	Male: 1.01 [0.98-1.05] (n=3458) Female: 0.91 [0.87-0.94] (n=2584)	Male: - Female: 0.77 [0.48-0.90] (n=46)	Male: 0.75 [0.65-0.84] (n=43) Female: -
Oxidative	<i>SEC14L4</i>	rs9606739	0.96 [0.93-0.99] (n=3837)	0.95 [0.91-0.99] (n=2016)	0.83 [0.72-0.96] (n=247)
Osmotic	<i>MYO9B</i>	rs35365035	0.97 [0.93-1.01] (n=2118)	0.95 [0.92-0.98] (n=2879)	0.91 [0.86-0.97] (n=1113)
Osmotic	<i>HBA2</i>	Chr 16:226229	0.95 [0.93-0.97] (n=5868)	0.82 [0.66-0.99] (n=134)	0.52 [0.10-1.14] (n=10)

RBC=red blood cell; SNP=single nucleotide polymorphism; CI=confidence interval

^ADifferences of hemoglobin increments in the receipt of homozygous recessive vs. dominant RBC units were statistically significant (p<0.05)

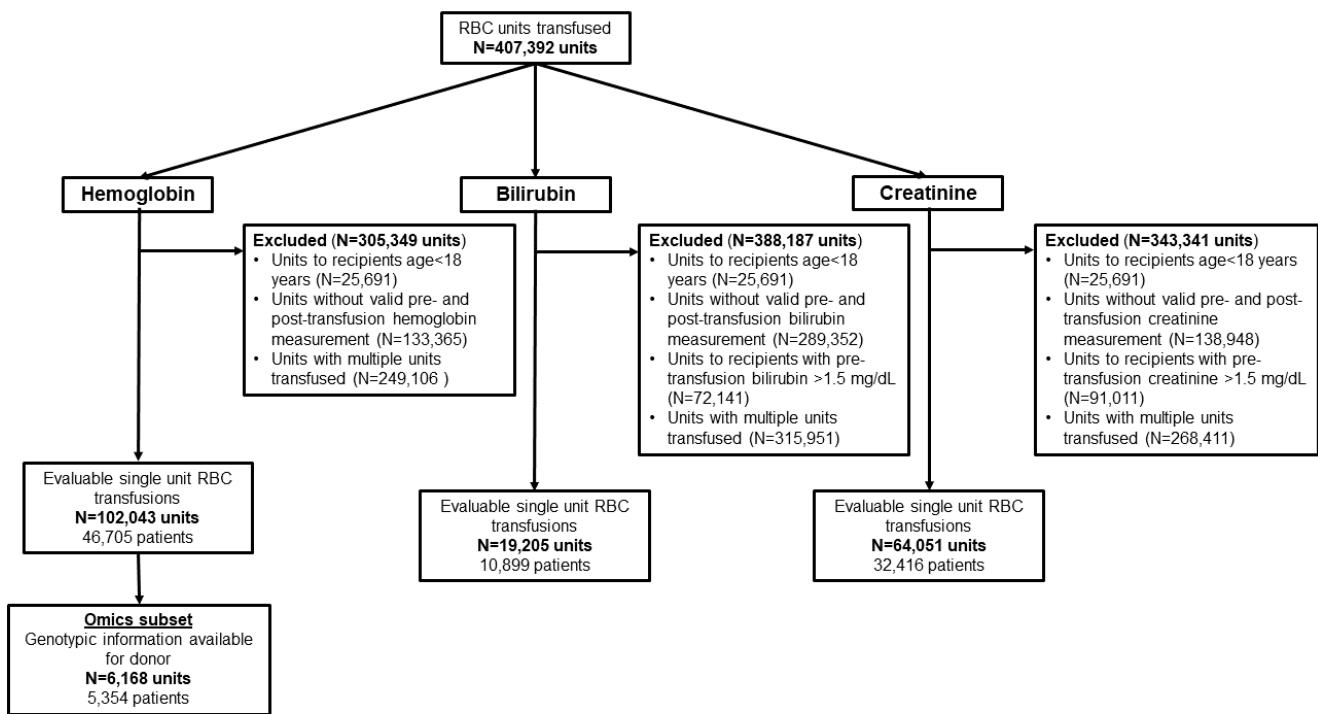


Figure 1. Study flow diagram. RBC = red blood cell.

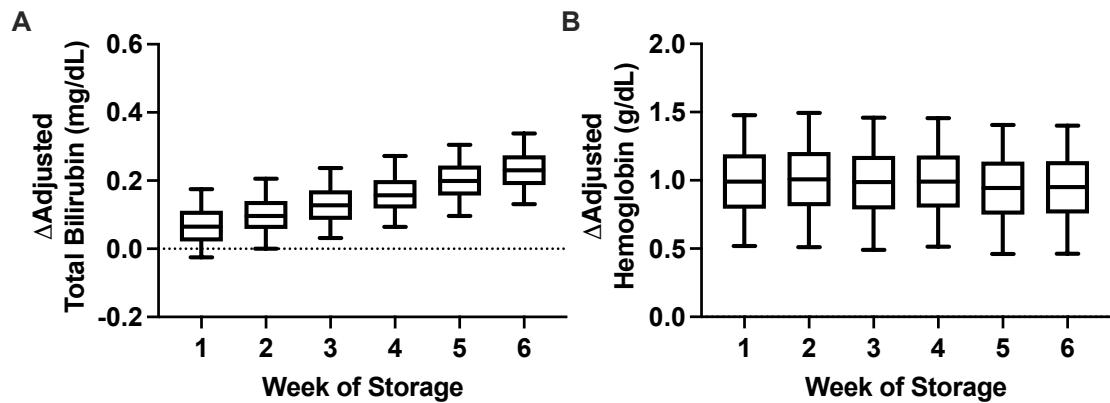


Figure 2: Effect of storage age and irradiation on hemoglobin and bilirubin increments. (A) The adjusted change in total bilirubin (N=19,205) and (B) hemoglobin (N=102,043) level between pre- and post-transfusion laboratory measures is shown stratified by week of storage, as labeled. Hemoglobin and bilirubin increments were adjusted for concomitant donor, component, and recipient factors presented in Table 2 and plotted as derivatives of the regression equation. $P<0.0001$ for linear association between storage duration and change in laboratory measure for both figure panels.

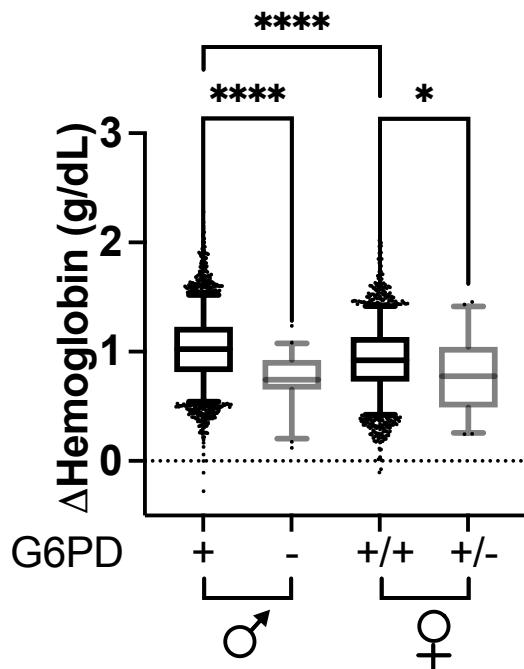


Figure 3: Adjusted hemoglobin increments stratified by gender and glucose-6-phosphate dehydrogenase (G6PD) deficiency. Adjusted change in hemoglobin level between pre- and post-transfusion measures is shown stratified by male (N=3458) and female (N=2584) gender and G6PD genotype, as labeled. As G6PD is X-linked, males labeled (-) are G6PD-deficient (N=43) and females labeled (+/-) are heterozygous (N=46). Hemoglobin is adjusted for concomitant donor, component, and recipient factors. ***P<0.0001, *P<0.05 for one-way ANOVA comparison as indicated. Line represents median, boxes represent 25th to 75th percentile, and whiskers represent the 5th to 95th percentile.