

1

SUPPLEMENTAL INFORMATION

2 Supplemental Figure 1

3

A >DOT1L; human; range -1000bp to 100bp.

TACCTCAGCCCCCGCAGTAGCTGGGATTACAGGCGCCCGCCACCAGCGCCAGCTAATTTTTGTATTTTAA
 ATAGAGACAGGGTTTCACCATGTTGGCCAGGCTGATCTCGAACTCCTGACCTCAGGTGATCTTCCCGCCT
 CGGCCTCACAAAGTGCTGGGATTACAGGCATGAGCCAGGGCTCCTGGCCGTGAGAGGGGGTTTCTCCATG
 TTGGTCAGGCTGGTCTCGAACTCCTGACCTCAGGTGATCTGCCCCTTGGCCTCCCAAAGTGCTGGGAT
 TACAGGGGTGAATCACCGCGCCTGGCCTTTTTTTTTTTTTTTTGTACCCAATAAACAGCATTTGTTGTGAA
 TGAATACTGGACTTGGAGCTACAGCAAGTCTGGACCGTGACTCTTATGGGGGGAAATTCGGATTTTTGG
 TTTTACTAAGCCGTGTGTGGGGAGGTGTCCGGCGTCTCCTCCTGGGACGGGATTCGAAACCCGCTATCCG
 ACGGGCCCGCCACAGGGTCTCCCGGGTCCCCGCTTCGGGCCGGCAGTGGGGGAAAGGGTTCGGCCGAG
 GGCAACCGAGG**ACGTGCGTG**CGTACGTTCTG**GCGTGCCTG**GATTTCGGGCGGGCGGGCAGTCCACGGGGC
 GGGGCGCCGAGGGGTGGCGCGCGGGTTCGGCCGCTGGGCGGGGGCACGCGCCGGCGTCTTCGCTCC
 GGGTCCCCTAGCGCGCGGGGGGAGTGGTTCGGCCCGGCCCGGCTCATTGTGCTCGCTTCACGCCGGC
 CCAAGATGGCGGAGGCGCTGGAGGCCCGGGCCTGTGACTACAAAGAGGGAGTCGGGGGGCCGGGCCGGAC
 CGGAGCGCGGGCGGGCGGGCGGGCGGGCCGAGGCCAGGCCAGGCCCTCCCTCAGCCTCCCGCCCC
 TCCCTCCCGCCCCCTCCTCCGCCACCAGCGGCCCGGCCCTCCCCCAACCGCCCGCTAGCATGGTG
 CGGCGGCCGCGCGCGGACATGGGGGAGAAGCTGGAGCTGAGACTGAAGTCGCCGTGGGGGGTGGAGCC
 CGCCGTCTACCCGTGGCCGTGCCGGTCT**ACGTG**AGTGCCGCCCTCCACCG

B >DOT1L; mouse; range -1000bp to 100bp.

TTCTAGCTCCGGATCCTTCTTTTTGGGGGCTGGCAAGCGCTAGGATGAGGGGTGGGAGCCATCTCCCTAG
 ATTACATTCACCTCCCATGTAGTCAGGAAGAAGAAGCAGCTATCCAGAAGCGGCTGGTGAGCTGGTGACAG
 CTCACCTCAAGGTAACCTATTCTCAGAAGTTTTGGGGGGAGGAACTTAAGTATTAGAAGGATTGGAAT
 GGCCTAAGAAGGAAATAAGTTTGGCTCTGACTCTGTAAAGTTCTGGGCAGGTGAGAGAATAACATTGCT
 TTGAACTCTTAATTGAGAGCCTCCGACCTATAGGGTCCAATTTGACTGATGATTTATTAATTAACCTATT
 GCTCACATCTTTAATCCAGCTCTCAGGAGGCAGAGGCAGGTGGATTTTGGTAAATTCGAGGTCAGACTG
 GTCAACAGAATGAGTTTCTGGACAGCCAGGCACCCCTGTCTCAAAAAAAAAAAAAAAAAAAGCATG
 TAAATAAACAATAAGCAGATAGGCTAGAAAAGTGAAGAAATAGAAAAGCTAAAGAAGGGGAAAAATGAA
 AAATGAAAAAAAAAAGAAAGAAAGAAAAAGCCAGAAAAACAGAAAAAGAAAATTGACGGGAAGGGAAGAG
 AAAGAAAAAGAACAGGAAATGGAAAGAAAAAGCAAGCAAGCGAAGGGACGCGAAGCAAGCCT
 GTGCAACTGGGATTTGAACTTGACCCCGCTGGTCTCTCTCGCTTTCTCCGAGGGTTTCCCGGGTCC
 CCGCTTCCGGCTGGCGGCGACGTCGGAGGGCAACCGAGG**ACGTGCGTG**CGCCGCT**GCGTG**CGTAAAGT**GCG**
TGCGTCGGTCGGAGGAGGGCGAGTCCAAGGGG**GCGTGCGTG**CGAGGGGTGGCGCGCGGGCGGGCGGG
 CGGAGGCTCTGGGCGCGCACCGCCTCTGGCTCCCGCCTCCCGCGCGCGCTCGCCGAGTGGTTCGCG
 CCGACCCCGGCTCATTGTGCCTTCTCCTCACGCCGGCCAAAGATGGCGGGCGGCTCTAGACGCCCGGG
 TCTGTGACTCCTACAAAGAGGGGAGCTGGGGCCACACGGGAGCGGGCGGG

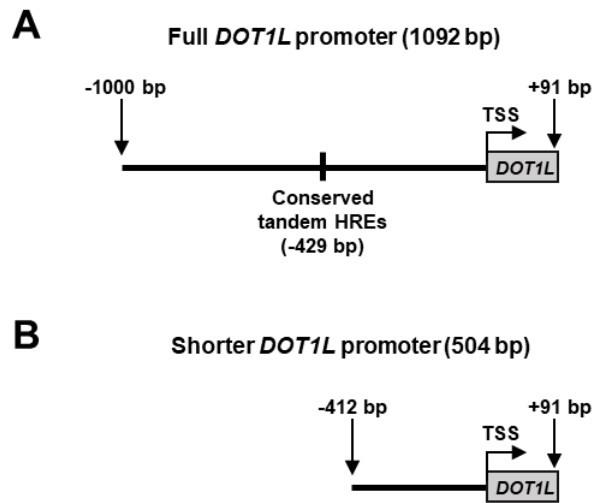
C >DOT1L; rat; range -1000bp to 100bp.

ATGTATGTTTATGCACCAGATTGCCCTGGTGCATGGAGAGGCCAGAAGAGGGCGTTGGAACCTGGGAAT
 TCAGTCAGAGATGGTTGTGAGCCGAGGGAGGGCTGGGAATGGAATCTAAGTTCTCTAAAAGAGCAGCC
 TGTAGTCTTAGCTACTGAACCATCTCTTAACCCATTTGATGATTTTGTTTTTTTCAGTTTGTCTTGCTG
 TTAGGAATGGTTTTCACTCTGTAGCTCAAGTGGGCTTTGAGTCGTGATCCCTTGCCCTCAGCTTCCCAAGAC
 CTTGGATTATGGGTCATAGATTCACGGTTTTGTTAGTGTGAACTGCATAGAAAGTCGAAAAATATCTTC
 AAAAGAATGAAAAGAAGCTAG**GCGTG**GTGGTGCACACCTTTAATCCAGCACTAGGGAGGGAGGCAGAG
 GCGGGGAATCTTGGTGAATTTGAGGTAAGACTGGTCAACAGAGTAGACGGAGTTCCAGGACAGCCAGGGC
 TACCGTGTCTCAAAAAAAAAAAAAAAAAACCAATAAGTAAATAACAATAAATAAGTGATAAGATAAAAA
 AAAGTGAAAAGAGAAACAGAAAAAGAAAAAGCCGGAAAAACAGAAAAAGAAATAGGGACGAGAAGAGAA
 AGAAAAGGAAAAGGAAAAGAAAAGAAAAAGAAAAGAAAAAGAAAAAGCAAGCAAGCGAGCCT
 GCGCAACTGGGACTTGAACCTGTCCACCTGGTCTCTCTCGCTTGCCTCAGCAGGGTTTCCCGGGTCC
 CCGCTTCCGGCTGGCGGTGGCGTCGGAGGGCAACCGAGG**ACGTGCGTG**CGACCCT**GCGTG**CGA**ACGTGCG**
TGCGTCGGCGGGAGGTGGCGAGTCCAAGGGG**GCGTG**CGTACGAGGGGTGGTGCGCGGCGCGGGCGGG
 CGGAGGCTTTGGGCGCGCACCGCCTCTGGCTCCCGCCTCCCGCGCGCGCGCCAGTGGTTCAGC
 CCGACCCCGGCTCATTGTGCCTTCTCCTCACGCCGGCCAAAGATGGCGGGCGGCTCTAGACGCCCGGG
 TCTGTGACTCCTACAAAGAGGGGAGCTGGGGCCACACGGGAGCGGTGGCCG

4 **Supplemental Figure 1** Predicted hypoxia response elements (HREs) in human, mouse and rat
5 *DOTIL* gene promoters. (A-B-C) Sequences of the human (A), mouse (B) and rat (C) *DOTIL*
6 gene promoters from 1000 base pairs (bp) upstream of the transcription start site (TSS) to 100
7 bp downstream of the TSS as determined by the Eukaryotic Promoter Database. HREs with
8 core consensus sequence 5'-(A/G)CGTG-3', potential binding sites for HIF heterodimers, are
9 marked with a yellow box. The bp downstream of the TSS are highlighted in grey.

13 **Supplemental Figure 2** Alignment of the human, mouse and rat *DOTIL* gene promoters.
14 Alignment of the human, mouse and rat sequences of the *DOTIL* gene promoters from 1000
15 base pairs (bp) upstream of the transcription start site (TSS) to 100 bp downstream of the TSS
16 as determined by the Eukaryotic Promoter Database. This alignment was performed using
17 MUSCLE. Hypoxia response elements (HREs) are marked with a yellow box.

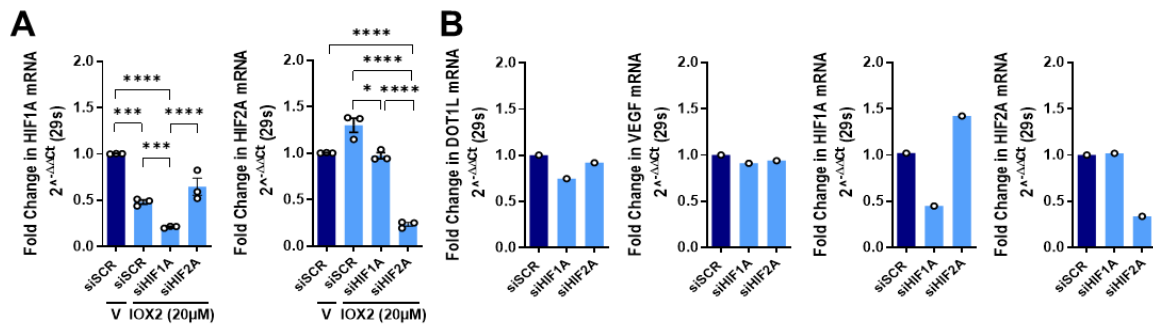
18 **Supplemental Figure 3**



19

20 **Supplemental Figure 3** Promoter constructs for the luciferase reporter assay. (A) The full
21 human *DOT1L* promoter from 1000 base pairs (bp) upstream to 91 bp downstream relative to
22 the transcription start site (TSS) as defined by the Eukaryotic Promoter Database was cloned
23 into the pGL3 basic luciferase reporter vector. This 1092 bp fragment contains the conserved
24 overlapping tandem Hypoxia response elements (HREs) located at -429 bp relative to the TSS.
25 (B) The shorter *DOT1L* promoter from 412 bp upstream to 91 bp downstream relative to the
26 TSS was cloned into the pGL3 basic luciferase reporter vector. This 504 bp fragment does not
27 contain the conserved overlapping tandem HREs.

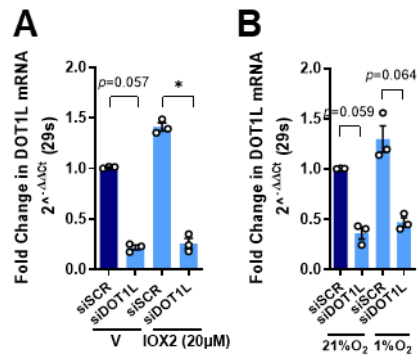
28 **Supplemental Figure 4**



29

30 **Supplemental Figure 4** Efficiency of siRNA-mediated gene silencing in C28/I2 cells. (A)
 31 Real-time PCR analysis of *HIF1A* and *HIF2A* in human articular chondrocyte C28/I2 cells after
 32 treatment with hypoxia mimetic IOX2 (20 µM) and siRNA-mediated silencing of *HIF1A*
 33 (siHIF1A), *HIF2A* (siHIF2A) or scrambled control (siSCR) (n=3, **p*<0.05, ****p* <0.001,
 34 *****p*<0.0001 Sidak-corrected for 6 tests in one-way ANOVA). (B) Real-time PCR analysis
 35 of *DOT1L*, *VEGF*, *HIF1A* and *HIF2A* in C28/I2 cells after siRNA-mediated silencing of
 36 *HIF1A*, *HIF2A* or scrambled control (siSCR) (n=1). Bar graphs are mean±sem.

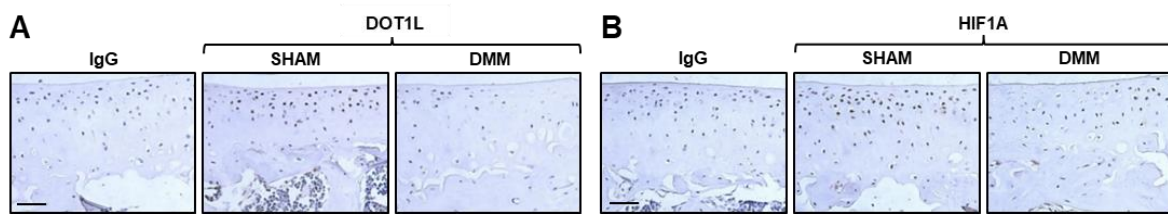
37 **Supplemental Figure 5**



38

39 **Supplemental Figure 5** Efficiency of siRNA-mediated gene silencing in hACs. (A-B) Real-
40 time PCR analysis of *DOT1L* in primary human articular chondrocytes (hACs) after treatment
41 with hypoxia mimetic IOX2 (20 μM) or vehicle (V) (A) (n=3, * $p < 0.05$, Sidak-corrected for 6
42 tests in two-way ANOVA) or culturing in hypoxic conditions (1% O₂) (B) and siRNA-mediated
43 silencing of *DOT1L* or scrambled control (siSCR) (n=3, Sidak-corrected for 6 tests in two-way
44 ANOVA). Bar graphs are mean ± sem.

45 **Supplemental Figure 6**



47 **Supplemental Figure 6** DOT1L and HIF-1A protein levels in osteoarthritis murine articular
48 cartilage. (A-B) Immunohistochemical detection of DOT1L (A) and HIF1A (B) in the articular
49 cartilage of wild-type mice with OA triggered by destabilisation of the medial meniscus (DMM)
50 surgery compared to sham operated mice. The images are representative of three different
51 animals. Scale bar, 50 μ m.

52 **Supplemental Table 1**53 **Human primers used for qPCR**

Primer name	Sequence
hACAN_fw	ATCCGAGACACCAACGAGAC
hACAN_rv	CACTCATTGGCTGCTTCCTG
hCOL2A1_fw	TGGCAGAGATGGAGAACCTG
hCOL2A1_rv	CATCAAATCCTCCAGCCATC
hDOT1L_fw	GGATCTCAAGCTCGCTATGG
hDOT1L_rv	GTCGATGGCACGGTTGTACT
hHIF1A_fw	TCATCCATGTGACCATGAGG
hHIF1A_rv	TTCCTCGGCTAGTTAGGGTACA
hHIF2A_fw	CTGCGACCATGAGGAGATTC
hHIF2A_rv	GTACGGCCTCTGTTGGTGAC
hS29_fw	GGGTCACCAGCAGCTGTACT
hS29_rv	AAACACTGGCGGCACATATT
hTCF1_fw	CCCCCAACTCTCTCTCTACGA
hTCF1_rv	TGCCTGAGGTCAGGGAGTAG
hVEGF_fw	TGCAGATTATGCGGATCAAACC
hVEGF_rv	TGCATTCACATTTGTTGTGCTGTAG

54

55 **Supplemental Table 2**56 **Human primers used for ChIP-qPCR**

Primer name	Sequence
hDOT1Lprom_fw	CCAATAAACAGCATTGTTGTCG
hDOT1Lprom_rv	CCCACACACGGCTTAGTAAAA
hVEGFprom_fw	TCACTTTCCTGCTCCCTCCT
hVEGFprom_rv	GCAATGAAGGGGAAGCTCGA

57

58 **Supplemental Table 3**59 **Primers used for the PCR amplification for luciferase assay**

Construct	Primer	Sequence
Full DOT1L promoter (1092 bp)	Forward	GGTACCTACCTCAGCCCCCGCAGTA
	Reverse	ACGCTCGAGGCGGCACTCACGTAGACC
Shorter DOT1L promoter (504 bp)	Forward	GGTACCCGTGCGTGCGTGGATTCG
	Reverse	ACGCTCGAGGCGGCACTCACGTAGACC

60

61

62 **Supplemental Table 4**

63

Statistical Analysis Details

FIGURE 3A	Generalized least square model with constant plus power variance function structure						
<i>DOT1L</i>	beta=0.006, t=6.209 $p<0.0001$						
<i>VEGF</i>	beta=0.018, t=10.65 $p<0.0001$						
FIGURE 3B	Generalized least square model with constant plus power variance function structure						
<i>DOT1L</i>	beta=0.015, t=11.14 $p<0.0001$						
<i>VEGF</i>	beta=0.005, t=24.62 $p<0.0001$						
FIGURE 3C	Welch-correct unpaired t-test						
<i>DOT1L</i>	$t_2=4.940$ $p=0.0385$						
<i>VEGF</i>	$t_2=6.231$ $p=0.0248$						
FIGURE 3F	Generalized least square model						
<i>Immuno-fluorescent</i>	beta=0.007, t=17.32 $p<0.0001$						
FIGURE 4A	Two-way ANOVA	V:Empty vs. V:DOT1L	V:Empty vs. IOX2:Empty	V:Empty vs. IOX2:DOT1L	V:DOT1L vs. IOX2:Empty	V:DOT1L vs. IOX2:DOT1L	IOX2:Empty vs. IOX2:DOT1L
Luciferase activity	$F_{1,8} = 21.71$ $p=0.0016$ for IOX2 treatment $F_{1,8} = 21.86$ $p=0.0016$ for reporter $F_{1,8} = 1.38$ $p=0.755$ for interaction	$t_8 = 3.078$ $p=0.088$	$t_8 = 3.067$ $p=0.089$	$t_8 = 6.601$ $p=0.001$	$t_8 = 0.0115$ $p=0.99$	$t_8 = 3.522$ $p=0.046$	$t_8 = 3.534$ $p=0.045$
FIGURE 4B	One-way ANOVA	siSCRv vs. siSCR	siSCRv vs. siHIF1A	siSCRv vs. siHIF2A	siSCR vs. siHIF1A	siSCR vs. siHIF2A	siHIF1A vs. siHIF2A
<i>DOT1L</i>	$F_{3,8} = 36.28$	$t_8 = 9.53$ $p=0.0001$	$t_8 = 4.426$ $p=0.0132$	$t_8 = 8.10$ $p=0.0002$	$t_8 = 5.10$ $p=0.0055$	$t_8 = 1.427$ $p=0.7207$	$t_8 = 3.678$ $p=0.0369$
<i>VEGF</i>	$F_{3,8} = 113.0$	$t_8 = 17.25$ $p=0.0001$	$t_8 = 10.68$ $p<0.0001$	$t_8 = 14.2$ $p<0.0001$	$t_8 = 6.57$ $p=0.0011$	$t_8 = 3.042$ $p=0.0923$	$t_8 = 3.526$ $p=0.0458$
FIGURE 5A	Linear mixed model						
<i>DOT1L</i>	beta=0.01, t=2.73 $p=0.046$						
<i>VEGF</i>	beta=0.023, t=15.71, $p<0.0001$						

FIGURE 5B	Paired t-test						
<i>DOT1L</i>	$t_2=3.280$ $p=0.0817$						
<i>VEGF</i>	$t_2=10.27$ $p=0.0093$						
FIGURE 5C	Linear mixed model						
<i>COL2</i>	beta=0.02, $t=4.27$, $p=0.0009$						
<i>ACAN</i>	beta=0.024, $t=4.69$, $p=0.0007$						
FIGURE 5D	Paired t-test						
<i>COL2</i>	$t_2=14.51$ $p=0.0047$						
<i>ACAN</i>	$t_2=16.84$ $p=0.0035$						
FIGURE 5E	Two-way ANOVA	Vehicle-siSCR vs vehicle-siDOT1L	Vehicle-siSCR vs IOX2-siSCR	Vehicle-siSCR vs IOX2-siDOT1L	Vehicle-siDOT1L v IOX2-siSCR	Vehicle-siDOT1L vs IOX2-siDOT1L	IOX2-siSCR vs IOX2- siDOT1L
<i>COL2</i>	$F_{1,2}= 3.378$ $p=0.2075$ for IOX2 treatment $F_{1,2}= 9.659$ $p=0.089$ for DOT1L silencing $F_{1,2}= 3.33$ $p=0.2084$ for interaction	$t_2=2.254$ $p=0.6306$	$t_2=6.480$ $p=0.1303$	$t_2=1.644$ $p=0.8103$	$t_2=8.734$ $p=0.0747$	$t_2=3.898$ $p=0.310$	$t_2=4.836$ $p=0.218$
<i>ACAN</i>	$F_{1,2}= 0.768$ $p=0.4732$ for IOX2 treatment $F_{1,2}= 25.93$ $p=0.0365$ for DOT1L silencing $F_{1,2}= 1.14$ $p=0.3977$ for interaction	$t_2=2.792$ $p=0.496$	$t_2=1.343$ $p=0.8933$	$t_2=2.958$ $p=0.461$	$t_2=4.136$ $p=0.282$	$t_2=0.1656$ $p=0.9$	$t_2=4.301$ $p=0.265$
<i>TCF1</i>	$F_{1,2}= 31.30$ $p=0.0305$ for IOX2 treatment $F_{1,2}= 80.53$ $p=0.0122$ for DOT1L silencing $F_{1,2}= 8.81$ $p=0.097$ for interaction	$t_2=4.880$ $p=0.215$	$t_2=3.384$ $p=0.383$	$t_2=5.694$ $p=0.164$	$t_2=8.265$ $p=0.083$	$t_2=0.814$ $p=0.985$	$t_2=9.079$ $p=0.069$
FIGURE 5F	Two-way ANOVA	Normoxia-siSCR vs normoxia-siDOT1L	Normoxia-siSCR vs hypoxia-siSCR	Normoxia-siSCR vs hypoxia-siDOT1L	Normoxia- siDOT1 vs hypoxia-siSCR	Normoxia-siDOT1L vs hypoxia-siDOT1L	Hypoxia-siSCR vs hypoxia-siDOT1L
<i>COL2</i>	$F_{1,2}= 226.3$ $p=0.0044$ for oxygen exposure	$t_2=3.435$ $p=0.375$	$t_2=12.68$ $p=0.0364$	$t_2=9.61$ $p=0.0623$	$t_2=16.1$ $p=0.0227$	$t_2=13.04$ $p=0.0344$	$t_2=3.07$ $p=0.4383$

	F _{1,2} = 12.95 <i>p</i> =0.069 for DOT1L silencing F _{1,2} = 0.07 <i>p</i> =0.812 for interaction						
ACAN	F _{1,2} = 158.4 <i>p</i> =0.0063 for oxygen exposure F _{1,2} = 65.75 <i>p</i> =0.0149 for DOT1L silencing F _{1,2} = 0.012 <i>p</i> =0.9225 for interaction	t ₂ =11.0 <i>p</i> =0.0476	t ₂ =15.58 <i>p</i> =0.0243	t ₂ =4.39 <i>p</i> =0.2555	t ₂ =26.6 <i>p</i> =0.0083	t ₂ =15.43 <i>p</i> =0.0248	t ₂ =11.18 <i>p</i> =0.0456
TCF1	F _{1,2} = 2.609 <i>p</i> =0.2476 for oxygen exposure F _{1,2} = 130.6 <i>p</i> =0.0076 for DOT1L silencing F _{1,2} = 9.23 <i>p</i> =0.093 for interaction	t ₂ =5.945 <i>p</i> =0.152	t ₂ =3.553 <i>p</i> =0.357	t ₂ =6.691 <i>p</i> =0.123	t ₂ =9.50 <i>p</i> =0.0637	t ₂ =0.745 <i>p</i> =0.989	t ₂ =10.24 <i>p</i> =0.055
FIGURE 5G	Two-way ANOVA	Normoxia V vs. normoxia EPZ	Normoxia V vs hypoxia V	Normoxia-V vs hypoxia EPZ	Normoxia UPZ vs hypoxia V	Normoxia EPZ vs hypoxia EPZ	Hypoxia V vs hypoxia EPZ
	F _{1,2} = 77.07 <i>p</i> =0.0127 for oxygen exposure F _{1,2} = 45.62 <i>p</i> =0.0212 for EPZ treatment F _{1,2} = 16.08 <i>p</i> =0.057 for interaction	t ₂ =11.7 <i>p</i> =0.0426	t ₂ =14.55 <i>p</i> =0.0278	t ₂ =8.528 <i>p</i> =0.078	t ₂ =26.2 <i>p</i> =0.0087	t ₂ =20.22 <i>p</i> =0.0145	t ₂ =6.022 <i>p</i> =0.1487
FIGURE 6B	Kruskal -Wallis	SHAM V vs DMM		DMM V vs DMM IOX2		SHAM V vs DMM IOX2	
OARSI	χ ² =14.297 <i>p</i> =0.0008	Z=3.09 <i>p</i> =0.002		Z=2.31 <i>p</i> =0.021		Z=2.41 <i>p</i> =0.016	
FIGURE 6C	Kruskal -Wallis	SHAM V vs DMM V		DMM V vs DMM IOX2		SHAM V vs DMM IOX2	
osteophyte	χ ² =15.36 <i>p</i> =0.0004	Z=2.71 <i>p</i> =0.0068		Z=2.22 <i>p</i> =0.0267		Z=2.71 <i>p</i> =0.0068	
FIGURE 6D	Kruskal -Wallis	SHAM V vs DMM		DMM V vs DMM IOX2		SHAM V vs DMM IOX2	
synovitis	χ ² =10.93 <i>p</i> =0.0064	Z=2.57 <i>p</i> =0.01		Z=1.75 <i>p</i> =0.08		Z=1.13 <i>p</i> =0.26	
FIGURE 6E	One-way ANOVA	SHAM V vs DMM		DMM V vs DMM IOX2		SHAM V vs DMM IOX2	
HIF1A	F _{2,12} =45.45 <i>p</i> <0.0001	t ₁₂ =9.533 <i>p</i> <0.0001		t ₁₂ =4.909 <i>p</i> =0.001		t ₁₂ =4.624 <i>p</i> =0.0018	
DOT1L	F _{2,12} =15.07 <i>p</i> =0.0005	t ₁₂ =4.574 <i>p</i> =0.0019		t ₁₂ =4.917 <i>p</i> =0.001		t ₁₂ =0.3435 <i>p</i> =0.98	
H3K79	F _{2,12} =45.12 <i>p</i> <0.0001	t ₁₂ =7.792 <i>p</i> <0.0001		t ₁₂ =8.60 <i>p</i> <0.0001		t ₁₂ =0.808 <i>p</i> =0.82	

FIGURE S4A	One-way ANOVA	Vehicle-siSCR vs IOX2-siSCR	Vehicle-siSCR vs IOX2-siHIF1a	Vehicle-siSCR vs IOX2-siHIF2a	IOX2-siSCR vs IOX2-siHIF1a	IOX2-siSCR vs IOX2-siHIF2a	IOX2-siHIF1a vs IOX2-siHIF2a
<i>HIF1A</i>	$F_{3,8} = 77.26$	$t_8 = 7.013 p = 0.0007$	$t_8 = 14.80$ $p < 0.0001$	$t_8 = 4.374 p = 0.0141$	$t_8 = 7.786 p = 0.0003$	$t_8 = 2.639 p = 0.1659$	$t_8 = 10.42 p < 0.0001$
<i>HIF2A</i>	$F_{3,8} = 191.4$	$t_8 = 3.208 p = 0.0725$	$t_8 = 0.4094$ $p = 0.9992$	$t_8 = 18.36 p < 0.0001$	$t_8 = 3.617 p = 0.0402$	$t_8 = 21.57 p < 0.0001$	$t_8 = 17.96 p < 0.0001$
FIGURE S5A	Two-way ANOVA	Vehicle-siSCR vs vehicle-siDOT1L	Vehicle-siSCR vs IOX2-siSCR	Vehicle-siSCR vs IOX2-siDOT1L	Vehicle-siDOT1L vs IOX2-siSCR	Vehicle-siDOT1L vs IOX2-siDOT1L	IOX2-siSCR vs IOX2-siDOT1L
<i>DOT1L</i>	$F_{1,2} = 7.730 p = 0.1087$ for IOX2 treatment $F_{1,2} = 140.9 p = 0.0070$ for DOT1L silencing $F_{1,2} = 0.7453 p = 0.4789$ for interaction	$t_2 = 9.986 p = 0.0578$	$t_2 = 2.150$ $p = 0.6601$	$t_2 = 9.057 p = 0.0697$	$t_2 = 12.14 p = 0.0397$	$t_2 = 0.9286$ $p = 0.9727$	$t_2 = 11.21 p = 0.0463$
FIGURE S5B	Two-way ANOVA	Normoxia-siSCR vs normoxia-siDOT1L	Normoxia-siSCR vs hypoxia-siSCR	Normoxia-siSCR vs hypoxia-siDOT1L	Normoxia-siDOT1L vs hypoxia-siSCR	Normoxia-siDOT1L vs hypoxia-siDOT1L	Hypoxia-siSCR vs hypoxia-siDOT1L
<i>DOT1L</i>	$F_{1,2} = 12.32 p = 0.0725$ for oxygen exposure $F_{1,2} = 91.27 p = 0.0108$ for DOT1L silencing $F_{1,2} = 0.06641$ $p = 0.8207$ for interaction	$t_2 = 9.809 p = 0.0599$	$t_2 = 2.292$ $p = 0.6201$	$t_2 = 7.153 p = 0.1087$	$t_2 = 12.10$ $p = 0.0399$	$t_2 = 2.656 p = 0.5270$	$t_2 = 9.445 p = 0.0644$