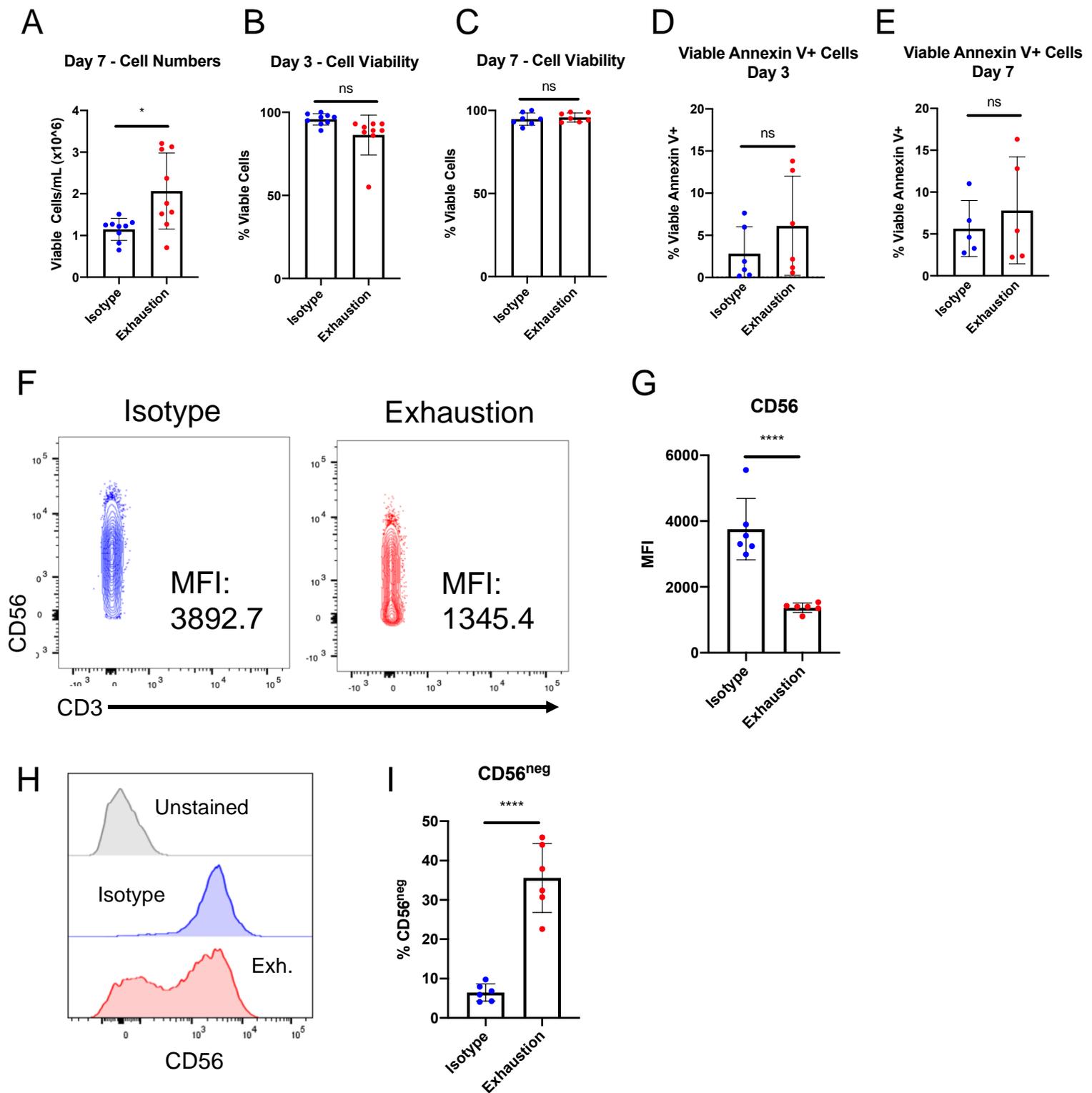
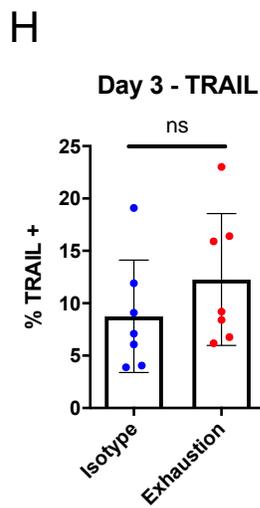
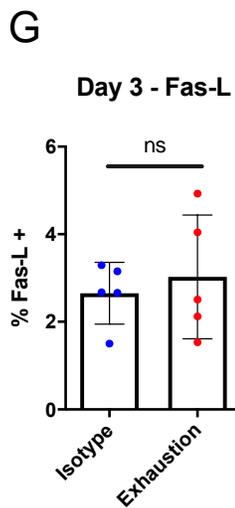
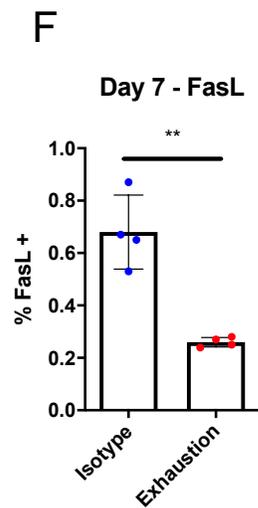
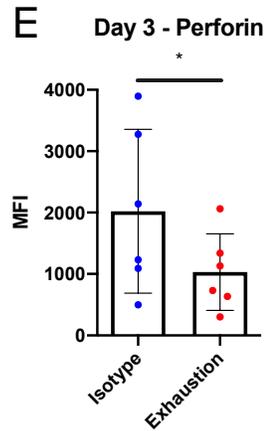
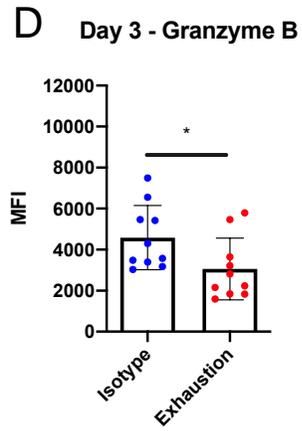
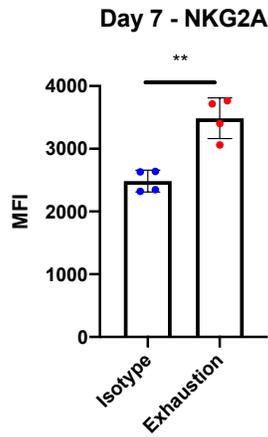
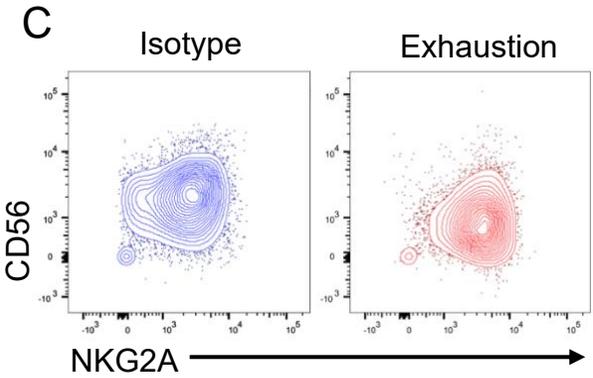
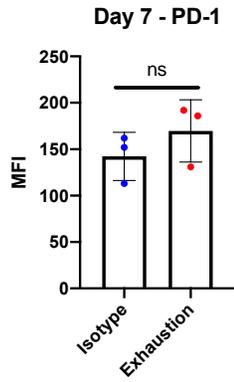
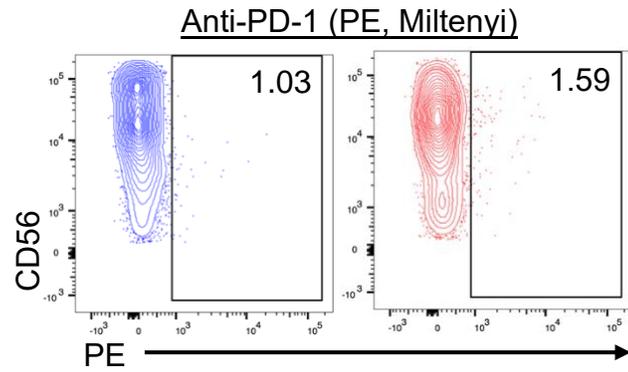
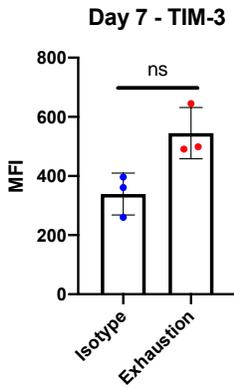
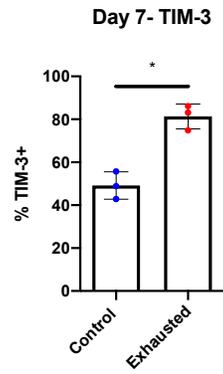
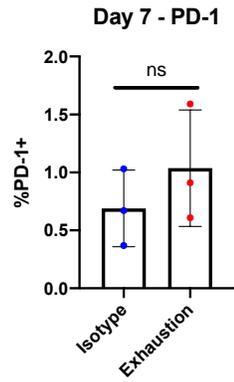
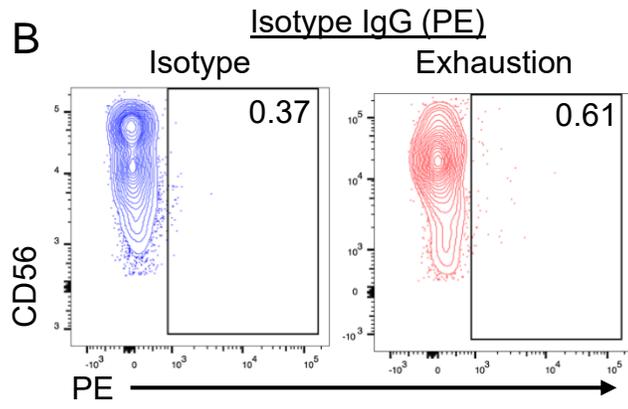
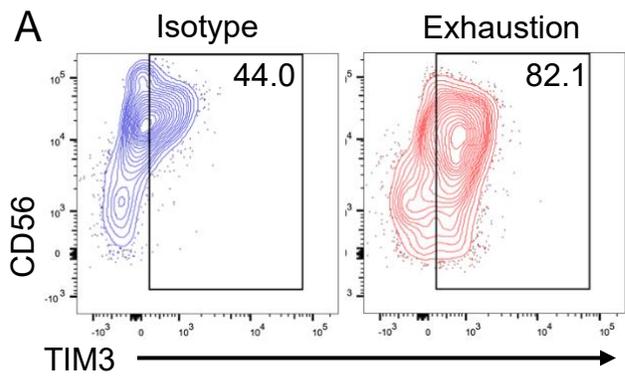


SUPPLEMENTARY FIGURE 1. (A-C) NK cells harvested from isotype-coated and exhaustion plates (day 3) were incubated with K562 targets for four hours (E/T: 2:1). Production of IFN γ (A) ($n=8$) and TNF α (B) ($n=8$) and degranulation (CD107a [C]) ($n=8$) were measured via flow cytometry. (D-F) NK cells were stimulated through NKP46 and NKG2D independently and simultaneously for 7 days. Production of IFN γ (D) ($n=3$) and TNF α (E) ($n=3$) and degranulation (CD107a [F]) ($n=3$) were measured via flow cytometry. (G-M) On day 3 of stimulation, flow cytometry was used to stain for several receptors. Percent of parent and MFI are included where indicated. CD96 (G-H), TIGIT (I-J), CD16 (K-L), NKG2D (M); ($n=7$); * $P \leq 0.05$; ** $P < 0.01$; *** $P < 0.001$; **** $P < 0.0001$ ns, not significant.

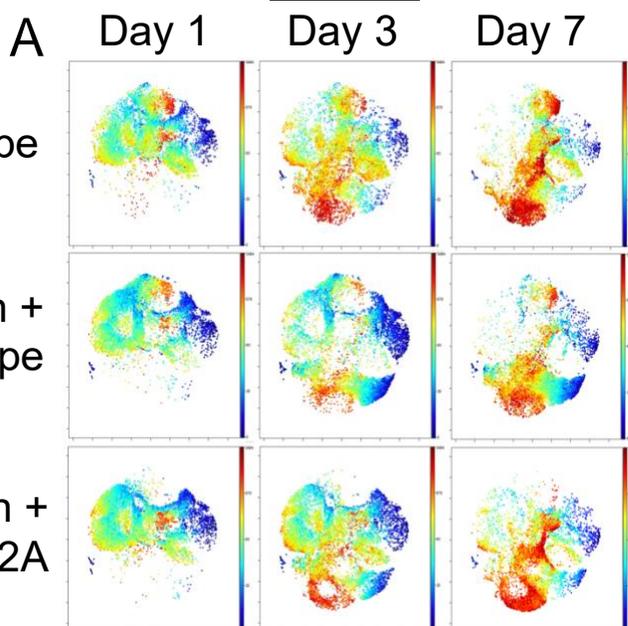
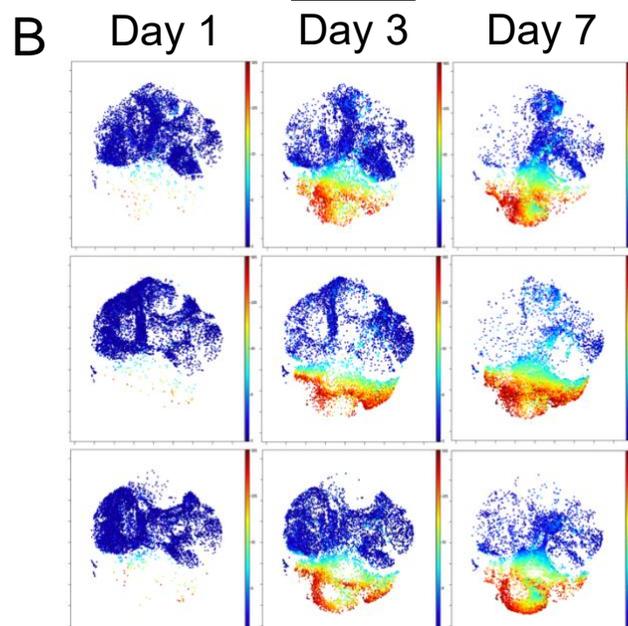
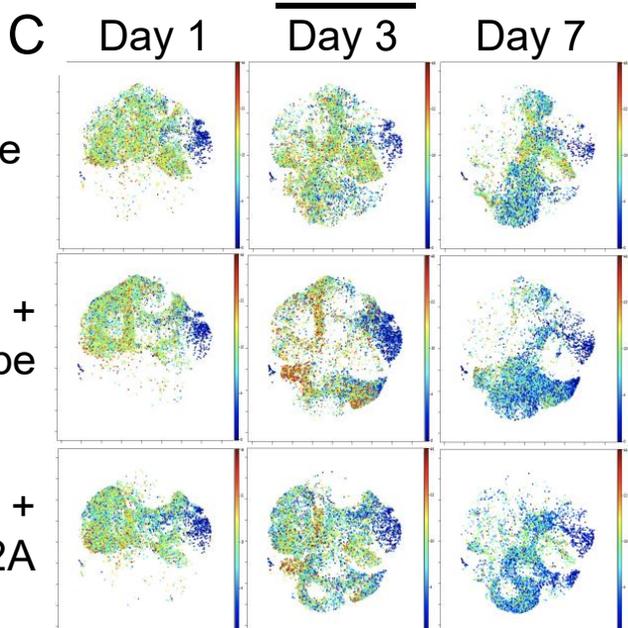
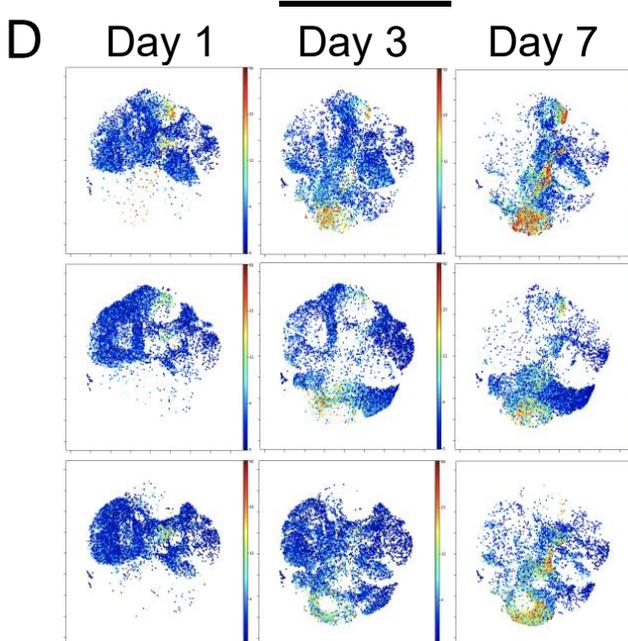
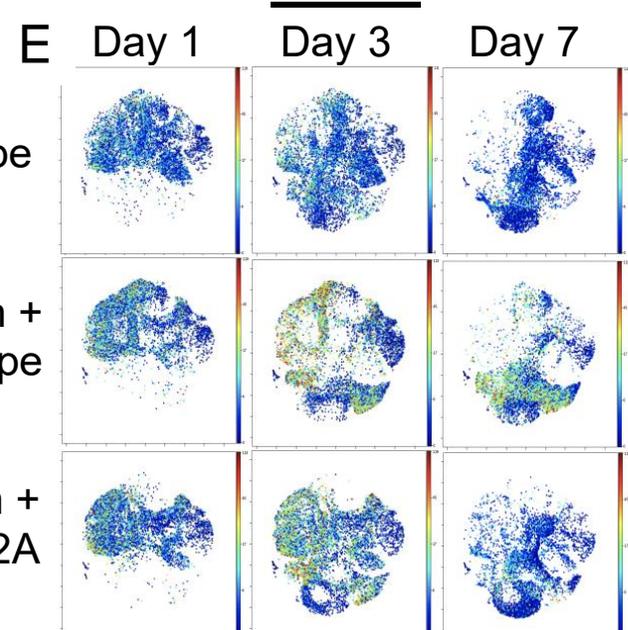
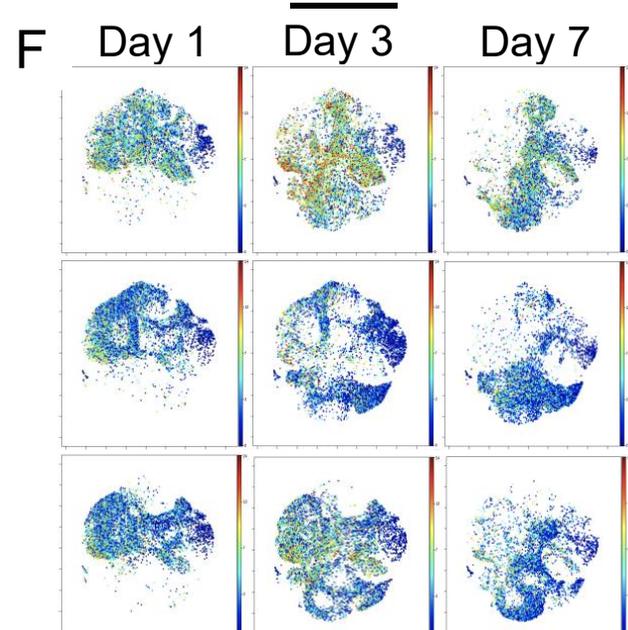


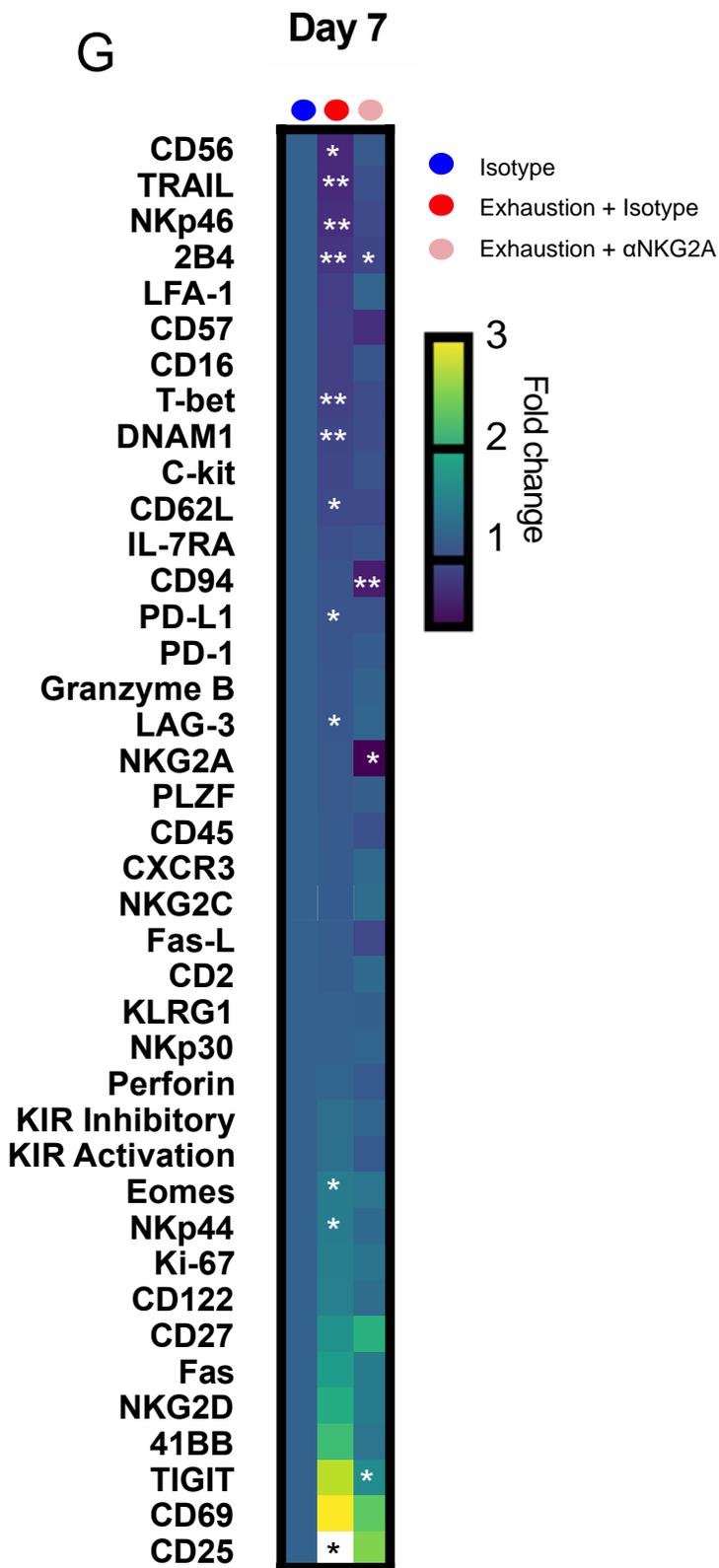
SUPPLEMENTARY FIGURE 2. (A) Viable cells were quantified at day 7 using the Beckman Coulter ViCell counter. (Starting cell density = 1×10^6 cells/mL, $n=9$.) Paired *t* tests were used for comparisons. $*P \leq 0.05$. (B-C) Viability of NK cells harvested from isotype-coated and exhaustion plates (harvested at days 3 [B] [$n=9$] and 7 [C] [$n=7$]) was assessed via Trypan blue uptake. (D-E) NK cells harvested from isotype-coated

and exhaustion plates (days 3 [D] [$n=6$] and 7 [E] [$n=5$]) were stained with Annexin V to assess apoptosis. (Analysis was performed via flow cytometry.) (F) Representative contour plots of CD56 expression for isotype (blue) and exhausted (red) cells. Analysis was performed via flow cytometry: NK cells were live, CD3- CD56+. (G) Quantification of pooled results ($n=7$). (F) Histogram of CD56 expression. (I) Quantification of pooled results ($n=6$) *** $P < 0.001$; **** $P < 0.0001$ ns, not significant.

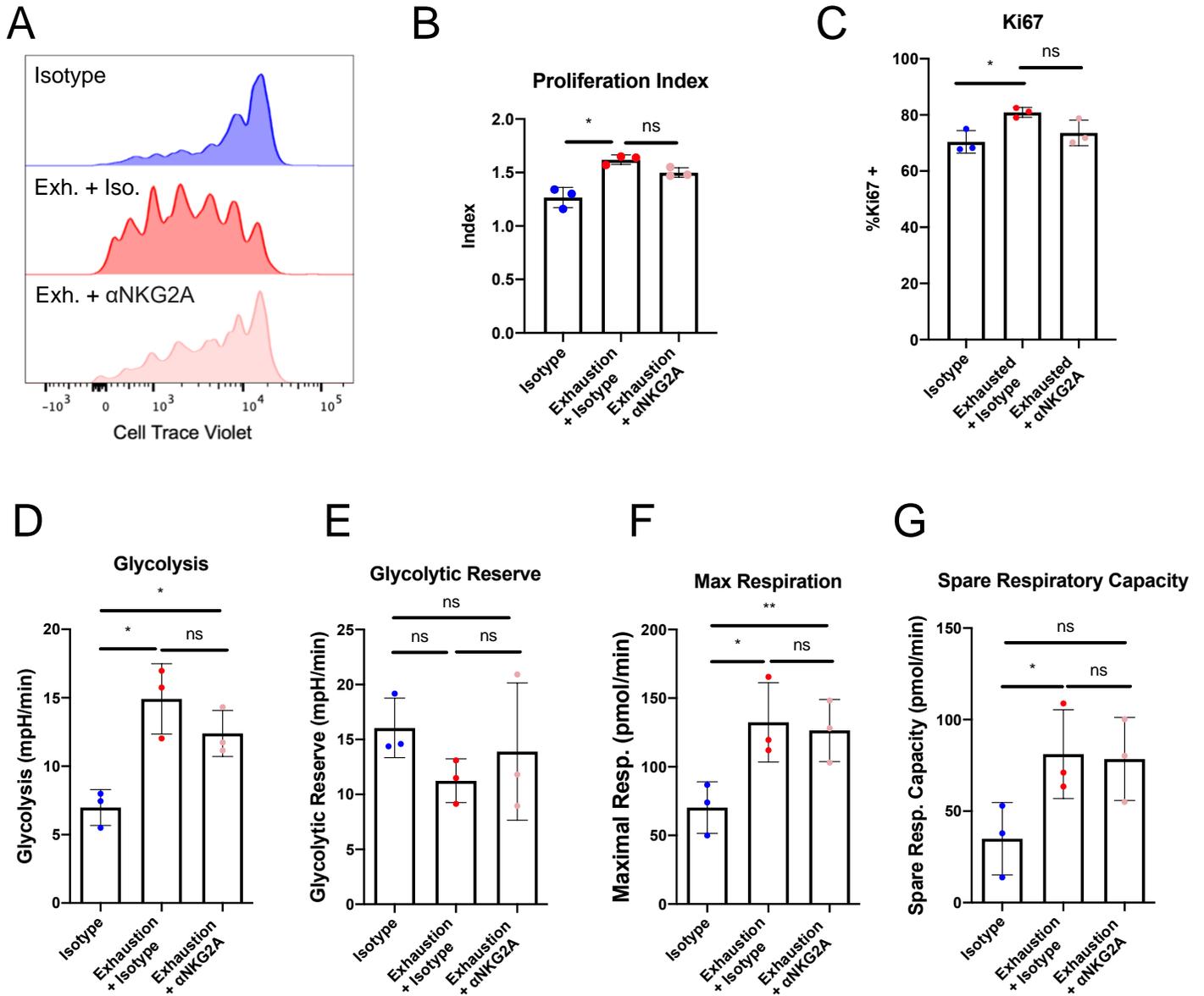


SUPPLEMENTARY FIGURE 3. (A-H) NK cells were harvested from isotype-coated and exhaustion plates and the expression of various receptors was analyzed via flow cytometry: TIM3, Day 7 (A) ($n=3$) representative dot plots, MFI data, and % of parent included; PD-1, Day 7 (B) ($n=3$) PE-conjugated isotype control, representative dot plots, MFI data, and % of parent included; NKG2A (C) ($n=4$) representative dot plots and MFI data included; Granzyme B, Day 3 (D) ($n=10$); Perforin, Day 3 (E) ($n=6$); Fas-L, Day 7 and 3 (F-G) ($n=4$, $n=5$); TRAIL, Day 3 (H) ($n=8$). * $P \leq 0.05$; ** $P < 0.01$; ns, not significant.

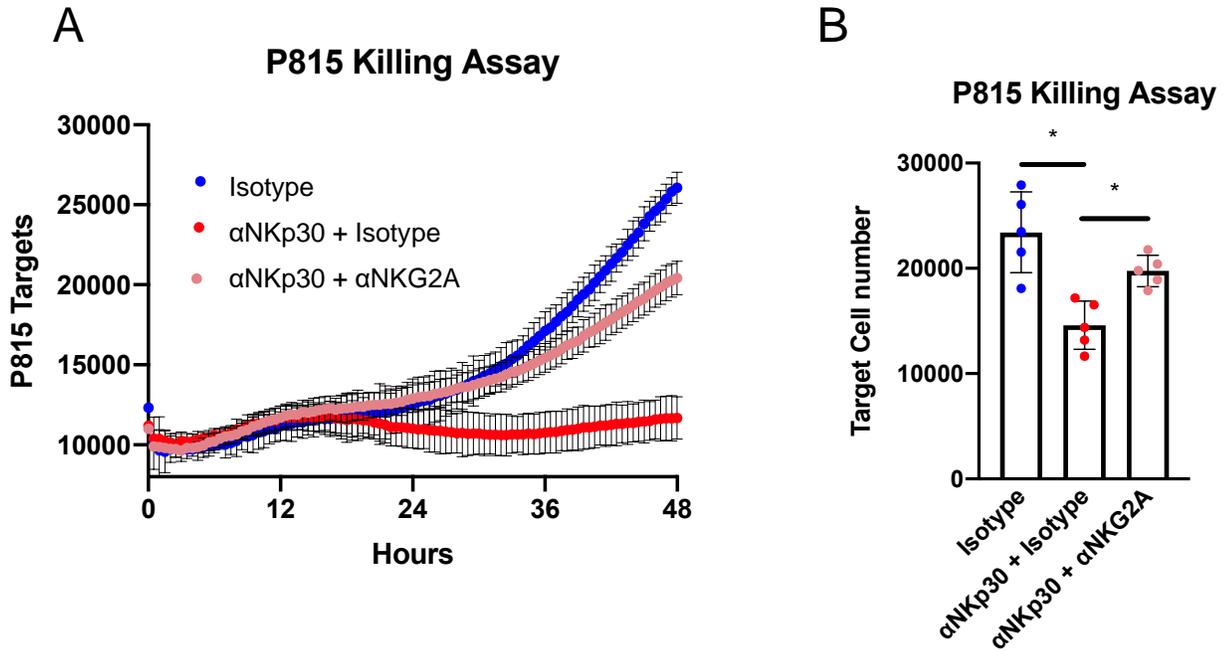
CD56**Ki67****T-bet****TRAIL****TIGIT****2B4**



SUPPLEMENTAL FIGURE 4. (A-G) At day 7 of stimulation, cells from three donors were analyzed via mass cytometry (CyTOF). viSNE analysis was performed for several markers: CD56 (A), Ki67 (B), T-bet (C), TRAIL (D), TIGIT (E), and 2B4 (F). Heatmap (G) indicates fold change relative to isotype mean metal intensity (MMI). * $P \leq 0.05$; ** $P < 0.01$; data that is not significant was left unlabeled ($n=3$).



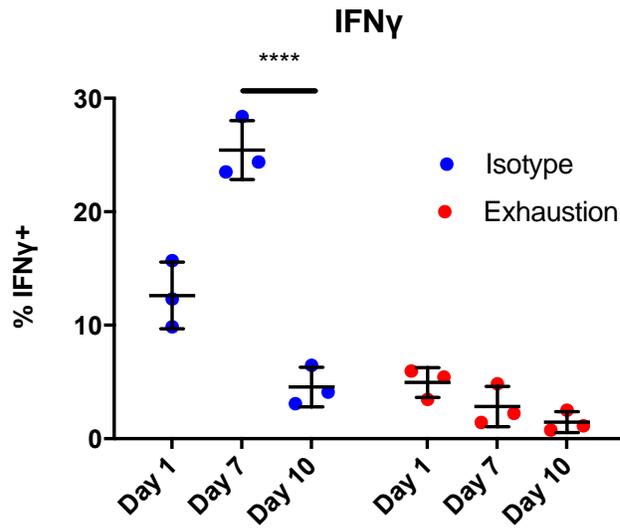
SUPPLEMENTAL FIGURE 5. (A) NK cells were stained with Cell Trace Violet and cultured on stimulatory plates for 7 days, as previously described. Cells were analyzed via flow cytometry 7 days later. (B) Proliferative indices were calculated using FlowJo V10 (C) NK cells were stained with Ki67 and analyzed via flow cytometry ($n=3$). (D-G) NK cells were harvested at day 7 and stained for Ki67 ($n=3$). (D-G) NK cells were immobilized on tissue culture plates and subjected to live-cell metabolic assays using Agilent's Seahorse XFe24 Analyzer. Extracellular acidification rates (ECAR) were measured following injection of glucose (G), oligomycin (O), FCCP (F), and Antimycin A + Rotenone (A+R). Oxygen consumption rates were measured following injection of glucose (G), oligomycin (O), FCCP (F), and Antimycin A + Rotenone (A+R). Graphical representations of glycolysis (D), glycolytic reserve (E), maximal respiration (F), and spare respiratory capacity (G) are pictured ($n=3$) ONE way ANOVA was used for comparisons. * $P \leq 0.05$; ** $P < 0.01$; ns, not significant.



SUPPLEMENTAL FIGURE 6. (A) P815 targets were labelled with Cell trace Far Red and coated with either anti-NKp30 and anti-NKG2A or anti-NKp30 and isotype IgG. P815 cells coated in isotype alone served as a negative control. NK cells were incubated with P815 target cells (E/T 2:1) and target cell growth was tracked using the Incucyte live cell imaging platform. (B) The number of target cells remaining after 48 hours was quantified for every condition and E/T ratio ($n=5$). $*P \leq 0.05$.

SUPPLEMENTAL FIGURE 7. (A) Schematic representing experimental design: sublethally irradiated NSG mice were injected (i.v.) with 1×10^6 NK cells. NK cells had been incubated for 7 days on plates with either isotype IgG (control) or anti-NKp46 and MICA/B as previously described. (B) 14 days post NK cell injection in the non-tumor model, blood was drawn, and NK cells were restimulated with K562 leukemia cells for 4 hours as previously described. Ki67, IFN γ , TNF α , and CD107a expression was assessed via flow cytometry. NK cells were CD45h+ CD56+ CD3-. Paired *t* tests were used for comparisons (*n*=8). ***P* < 0.01; ns, not significant. (C) Tumor burden at several time points was measured via BLI. One way ANOVA was used for comparisons. **P* < 0.05; ****P* < 0.001; ns, not significant (*n*=8). (D) 14 days post NK cell injection in the HL60 tumor model, blood was drawn, and NK cells were restimulated with K562 leukemia cells for 4 hours as previously described. TNF α and CD107a expression was assessed via flow cytometry. NK cells were CD45h+ CD56+ CD3-. Paired *t* tests were used for comparisons (*n*=8). ****P* < 0.001; ns, not significant.

A



SUPPLEMENTARY FIGURE 8. (A) NK cells harvested from isotype-coated and exhaustion plates (days 1, 7, and 10) were incubated with K562 targets for four hours (E/T: 2:1). IFN γ production was measured via flow cytometry. Paired *t* tests were used for comparisons ($n=3$). **** $P < 0.0001$.

Marker	Metal	Catalog no.	Clone
CD3	141Pr	3141019B	UCHT1
KIR Activating Pool	142Nd	201142A	KIR2DL1(143211)
C-Kit	143Nd	3143001B	104D2
CD69	144Nd	3144018B	FN50
KIR Inhibitory Pool	145Nd	201145A	KIR2DS1 (1127B)
CD8a	146Nd	3146001B	RPA-T8
Fas Ligand	147Sm	201147A	NOK-1
CD274 (PD-L1)	148Nd	3148017B	29E.2A3
CD25 (IL-2R)	149Sm	3149010B	2A3
PLZF	150Nd	201150A	6318100
CD2	151Eu	3151003B	TS1/8
CD95/Fas	152Sm	3152017B	DX2
CD62L (L-selectin)	153Eu	3153004B	DREG-56
CD16	154Sm	3154016B	MBSA43
CD27	155Gd	3155001B	L128
CXCr3	156Gd	3156004B	G025H7
CD137/4-1BB	158Gd	3158013B	4B4-1
CD337 (NKp30)	159Tb	3159017B	Z25
NKG2C	160Gd	201160A	2098A
Eomes	161Dy	201161A	644730
CD335 (NKp46)	162Dy	3162021B	BAB281
CD56 (NCAM)	163Dy	3163007B	NCAM 16.2
TRAIL	164Dy	201164A	RIK2
CD223/LAG-3	165Ho	3165037B	11C3C65
CD314 (NKG2D)	166Er	3166016B	ON72
NKp44	167Er	201167A	P-448
Ki-67	168Er	3168007B	B56
CD159a (NKG2A)	169Tm	3169013B	Z199
CD122	170Er	3170004B	Tu27
CD226 DNAM-1	171Yb	3171013B	DX11
CD57	172Yb	3172009B	HCD57
Granzyme B	173Yb	3173006B	GB11
CD279 (PD-1)	174Yb	3174020B	EH12.2H7

Perforin	175Lu	3175004B	B-D48
CD127 (IL-7Ra)	176Yb	3176004B	A019D5
TIGIT	209Bi	3209002B	MBSA43
CD45	89Y	3089003B	HI30
CD94	111CD	201111A	DX22
LFA-1	112CD	201112A	M24
KLRG-1	113CD	201113A	14C2A07
2B4	114CD	201114A	C1.7
T-bet	116CD	201116A	4B10

SUPPLEMENTARY TABLE 1. Antibodies used for Mass Cytometry by time-of-flight (CyTOF). Marker, metal tag, catalog number, and clone ID for each antibody are listed.