T-cell receptor sequencing identifies prior SARS-CoV-2 infection and correlates with neutralizing antibodies and disease severity

Rebecca Elyanow,^{1*} Thomas M. Snyder,^{1*} Sudeb C. Dalai,^{1,2**} Rachel M. Gittelman,¹ Jim Boonyaratanakornkit,^{3,4} Anna Wald,^{3,4,5,6} Stacy Selke,⁶ Mark H. Wener,^{3,6} Chihiro Morishima,⁶ Alexander L. Greninger,⁶ Michael Gale Jr.,^{7,8,9} Tien-Ying Hsiang,⁷ Lichen Jing,³ Michael R. Holbrook,¹⁰ Ian M. Kaplan,¹ H. Jabran Zahid,¹¹ Damon H. May,¹ Jonathan M. Carlson,¹¹ Lance Baldo,¹ Thomas Manley,^{1**} Harlan S. Robins,¹ David M. Koelle^{3,4,6,9,12}

*These authors share the first-author position and are listed in alphabetical order. **Author was an Adaptive employee at time of research.

¹Adaptive Biotechnologies, Seattle, Washington, USA
²Stanford University School of Medicine, Stanford, California, USA
³Department of Medicine, University of Washington, Seattle, Washington, USA
⁴Fred Hutchinson Cancer Research Center, Seattle, Washington, USA
⁵Department of Epidemiology, University of Washington, Seattle, Washington, USA
⁶Department of Laboratory Medicine and Pathology, University of Washington, USA

⁷Department of Immunology, University of Washington, Seattle, Washington, USA ⁸Department of Microbiology, University of Washington, Seattle, Washington, USA ⁹Department of Global Health, University of Washington, Seattle, Washington, USA ¹⁰National Institute of Allergy and Infectious Diseases Integrated Research Facility, Frederick, Maryland, USA ¹¹Microsoft Research, Redmond, Washington, USA

¹²Benaroya Research Institute, Seattle, Washington, USA

Corresponding author:

David M. Koelle, MD

750 Republican Street

Room E651, Mail Stop 358061

Seattle, WA 98109 USA

Tel (206) 616-1940

DKoelle@medicine.washington.edu

Supplemental Materials

Supplemental Tables

Supplemental Table 1. Longitudinal sensitivity of the T-Cell, EUROIMMUN, and

Abbott ARCHITECT tests by time from symptom onset.

Days From	T-Cell Test		EUROIMMUN		Abbott ARCHITECT	
Symptom			(anti-S1 IgG)		(anti-NP IgG)	
Onset	Sensitivity	95% CI	Sensitivity	95% CI	Sensitivity	95% CI
All	317/357	85 1_91 9	297/357	79 0-86 8	300/357	79 8-87 7
samples ^A	(88.8%)	00.1 01.0	(83.2%)	10.0 00.0	(84.0%)	10.0 01.1
29–50	60/70	77.1–92.9	57/70	71.4–90.0	59/70	75.7–91.4
	(85.7%)		(81.4%)		(84.3%)	
51_100	154/170	85 9_94 7	145/170	80 0-90 6	146/170	83 5-93 5
	(90.6%)		(85.3%)		(88.8%)	
101–150	84/97	79.4–92.8	81/97	76.3–90.7	80/97	75.3–89.7
	(86.6%)		(83.5%)		(82.5%)	
150–190 ^в	19/20	85 0-100	14/20	50.0–90.0	10/20	30.0–70.0
	(95%)		(70%)		(50%)	

^A P=0.01, T-cell test vs EUROIMMUN or Abbott ARCHITECT by mid-p McNemar's test.
 ^B P<0.03, T-cell test vs EUROIMMUN or Abbott ARCHITECT by mid-p McNemar's test.
 NP, nucleoprotein; S1, spike protein subunit 1.

Supplemental Table 2. Longitudinal sensitivity of the T-cell, EUROIMMUN, and

Days	T-Cell Test		EUROIMMUN		Abbott ARCHITECT	
From			(anti-S1 IgG)		(anti-NP IgG)	
Symptom	Lloopitalized	Non-	Lloopitalized	Non-	Lloopitalized	Non-
Onset	nospitalized	Hospitalized	nospitalized	Hospitalized	nospitalized	Hospitalized
All	41/44	276/313	44/44	254/313	42/45	257/313
samples ^A	(93.2%)	(88.2%)	(97.7%)	(81.2%)	(95.5%)	(82.4%)
29–50	3/4	57/66	4/4	53/66	4/4	55/66
	(75%)	(86.4%)	(100%)	(80.3%)	(100%)	(83.3%)
51–100	16/17	138/153	16/17	129/153	16/17	135/153
	(94.1%)	(90.2%)	(94.1%)	(84.3%)	(94.1%)	(88.2%)
101–150	18/19	66/78	19/19	62/78	18/19	62/78
	(94.7%)	(84.6%)	(100%)	(79.5%)	(94.7%)	(79.5%)
150–190	4/4	15/16	4/4	10/16	4/4	6/16
	(100%)	(93.8%)	(100%)	(62.5%)	(100%)	(37.5%)

Abbott ARCHITECT tests for hospitalized and non-hospitalized patients.

^A *P*=0.15 for hospitalized individuals and *P*=0.01 for non-hospitalized individuals comparing T-cell test vs EUROIMMUN or Abbott ARCHITECT by mid-p McNemar's test.

NP, nucleoprotein; S1, spike protein subunit 1.

Supplemental Figures



Supplemental Figure 1. Quantification of SARS-CoV-2–specific enhanced sequences and MIRA TCRs in ex vivo AIM-sorted CD4+ T cells. (A) PBMCs from 3 convalescent donors at 2 timepoints were subjected to AIM-based enrichment of SARS-CoV-2–specific T-cells using whole cell-associated viral antigen. Clonal breadth of SARS-CoV-2–specific enhanced sequences was compared between AIM-sorted CD4+ T cells and matched convalescent PBMCs. Significance was evaluated by Mann-Whitney U test. Data are expressed as median ± interquartile ranges. (B) Intersection of SARS-CoV-2–specific TCRs present in ex vivo AIM-sorted CD4+ T-cells and MIRA TCR set, annotated by protein specificity.



Supplemental Figure 2. Association of T-cell clonal depth with clinical variables. Correlation of clonal depth with (**A**) hospitalization, (**B**) fever, (**C**) difficulty breathing, (**D**) sex, and (**E**) age was evaluated by univariate Mann-Whitney U test (p) and multivariate linear regression with age, sex, hospitalization, fever, difficulty breathing, and TCR rearrangements as variables (\hat{p}). Data are expressed as median ± interquartile ranges (*n*=302). Population means and 95% confidence interval (CI) values for panels A–E were as follows: (**A**) Non-hospitalized, 88.3 (97.0-97.8), hospitalized, 168.3 (126.2-210.5); (**B**) no fever, 63.6 (50.3-76.9, fever, 108.2 (96.3-120.1); (**C**) no difficulty breathing, 85.6 (73.4-97.8), difficulty breathing, 105.4 (90.5-120.2); (**D**) female, 85.0 (72.4-97.6), male, 107.6 (93.1–122.1).



Supplemental Figure 3. Neutralizing antibody titer as a function of days from symptom onset to sample collection. Blue and red dots indicate hospitalized and non-hospitalized individuals, respectively. Trend lines connect visit 1 and visit 2 samples from the same individual. Blue and red bold trend lines indicate smoothed means [locally estimated scatterplot smoothing, LOESS (1)] for hospitalized and non-hospitalized individuals (*n*=302), respectively.

Supplemental Reference

 Cleveland WS. LOWESS: a program for smoothing scatterplots by robust locally weighted regression. *Am Stat.* 1981;35(1):54.