

Supplementary Figure legends:

Figure S1. Representative images of adipose tissue of WT or *Brd4*-CKO mice fed ND or HFD for 20 weeks.

Figure S2. Weights of indicated organs from WT or *Brd4*-CKO mice fed ND or HFD for 20 weeks. Data are means and standard deviation and are determined by an unpaired two-tailed Student's t test (left) or one-way ANOVA (right). n=5 mice. *** $P \leq 0.001$; ns, statistically not significant.

Figure S3. Metabolic studies of WT or *Brd4*-CKO mice fed HFD measured over 48 hr by CLAMS. (A) Physical activity measurement of ambulatory activity on the X-axis (XAMB), total beam breaks on the X-axis (XTOT), and total beam breaks on the Z-axis (ZTOT). (B&C) Food intake (B) and CO₂ release (C) of WT or *Brd4*-CKO mice fed HFD. Data are means and standard deviation and are determined by an unpaired two-tailed Student's t test. n=4-5 mice. * $P \leq 0.05$, ** $P \leq 0.01$, *** $P \leq 0.001$; ns, statistically not significant.

Figure S4. Plasma levels glucose (A) and insulin (B) of fasting WT or *Brd4*-CKO mice upon ND or HFD. Data are means and standard deviation and are determined by one-way ANOVA. n=5 mice. * $P \leq 0.05$, *** $P \leq 0.001$; ns, statistically not significant.

Figure S5. FACS analysis of stromal vascular cells (SVCs) of eWAT of WT or *Brd4*-CKO mice fed HFD for 20 weeks. (A) The representative plot represents adipose tissue macrophages (ATMs: CD45⁺ F4/80⁺ CD11b⁺). (B) The number of ATMs in eWAT of WT or *Brd4*-CKO mice fed HFD (left). The ratio of ATMs to SVCs (right). Data are means and standard deviation and are determined by an unpaired two-tailed Student's t

test. n=3 mice. **P≤0.01.

Figure S6. Heat map of the relative expression levels of genes from ATMs of HFD-fed WT or *Brd4*-CKO mice (scaled Z-score) based on log2-normalized expression levels of differently expressed genes (DEGs).

Figure S7. WT or *Brd4*-deficient BMDMs were treated with or without 1 nM Ins or 50 μM Rsg for 12 hr, *Gdf3* mRNA levels were analyzed by real-time PCR. Data are means and standard deviation and are determined by one-way ANOVA. n=3 culture dishes from 3 independent experiments. ns, statistically not significant.

Figure S8. Schematic representation of *Gdf3* promoter region with the positions of PPARγ binding site. TSS: transcription start site.

Figure S9. *Cdf3* gene tracks of ChIP-seq peaks for Brd4, PPARγ and histone modification H3K27Ac on chromosome 6, mm10. Y-axis indicated normalized ChIP-seq signals. Seven putative *Gdf3* enhancers are highlighted by boxes. The diagram was obtained by the modification of GEO Datasets (GSE109131 for Brd4, GSE21314 for PPARγ, and GSE106701 for H2K27Ac) of the National Center for Biotechnology Information.

Figure S10. The mRNA and protein levels of C/EBPα and PPARγ were measured by real-time PCR (A) or immunoblotting (B) in eWATs of WT or *Brd4*-CKO mice fed ND or HFD for 20 weeks. Each lane represents one mouse. Data are means and standard deviation and are determined by one-way ANOVA. n=4-5 mice. *P≤0.05, ns, statistically not significant.

Figure S11. Protein levels of Brd4 were detected by flow cytometry in adipose tissue

macrophages (ATMs) of WT mice fed ND or HFD for 20 weeks. Data are means with standard deviation and are determined by an unpaired two-tailed Student's t test. n=3 mice. ns, statistically not significant.

Figure S12. mRNA levels of indicated FAO related genes in eWAT (A), liver (B) and muscle (C) were measured by RT-PCR from WT or *Brd4*-CKO mice fed ND or HFD for 20 weeks. (D) Plasma levels of β -hydroxybutyrate in WT or *Brd4*-CKO mice fed ND or HFD for 20 weeks. All data are means and standard deviation and are determined by one-way ANOVA. n=4 mice. *P \leq 0.05, **P \leq 0.01, ***P \leq 0.001, ns, statistically not significant.

Figure S13. Plasma levels of FFA (A) and TG (B) in the WT or *Brd4*-CKO mice fed ND or HFD for 20 weeks. Data are means and standard deviation and are determined by one-way ANOVA. n=5 mice. *P \leq 0.05; ns, statistically not significant.

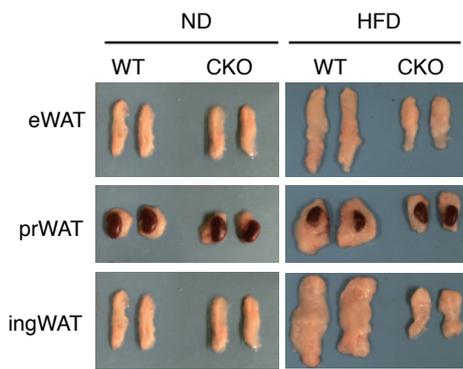
Figure S14. WT or *Brd4*-deficient BMDMs were treated with or without 0.1, 1, 10 nM Insulin (Ins) and 50 μ M rosiglitazone (Rsg) for 16 hr. PPAR γ protein levels were analyzed by immunoblotting.

Figure S15. WT or *Brd4* KO BMDMs were treated with or without RSG (50 μ M) for 24 h. The mRNA levels of indicated PPAR γ -targeted genes were analyzed by RT-PCR. Data are means and standard deviation and are determined by one-way ANOVA. n=3 culture dishes from 3 independent experiments. *P \leq 0.05, **P \leq 0.01, ns, statistically not significant.

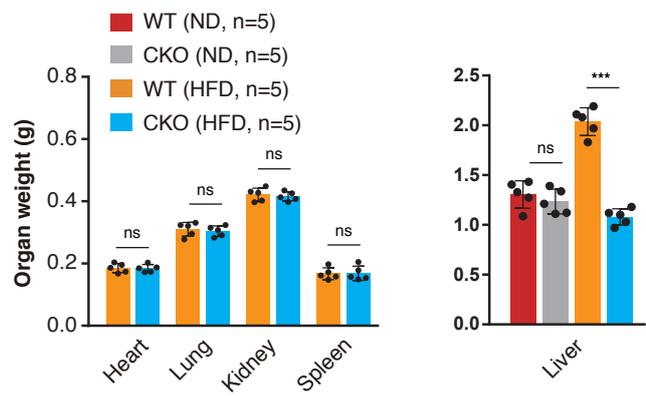
Figure S16. mRNA levels of *Il6* and *Tnfa* in CD11b⁺ ATMs isolated from WT or *Brd4*-CKO mice fed HFD for 20 weeks. Data are means and standard deviation and are determined by an unpaired two-tailed Student's t test. n=4 mice. ***P \leq 0.001.

Figure S17. WT BMDMs were pretreated with Brd4 inhibitor JQ1 (1 μ M or 5 μ M) for 1 hr, followed by the stimulation with Ins+Rsg for 12 hr. mRNA (A) and protein (B) levels of Gdf3 were analyzed by RT-PCR or immunoblotting, respectively. Data are means and standard deviation and are determined by one-way ANOVA. n=3 culture dishes from 3 independent experiments. ** $P \leq 0.01$; *** $P \leq 0.001$; ns, statistically not significant.

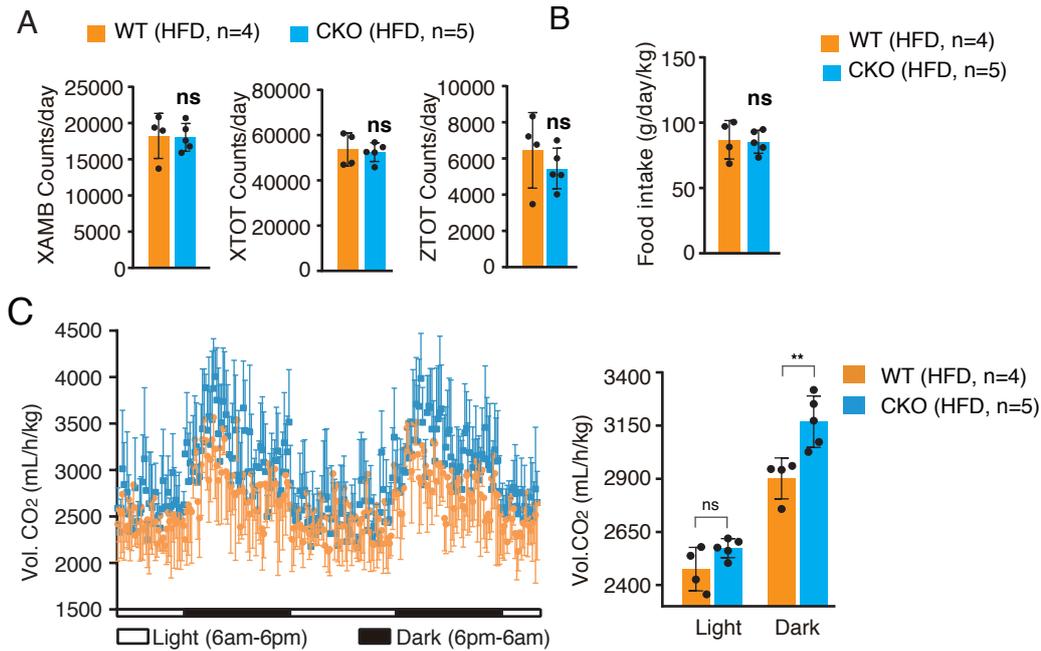
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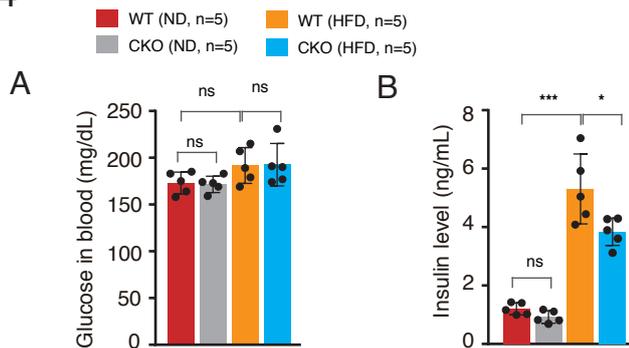
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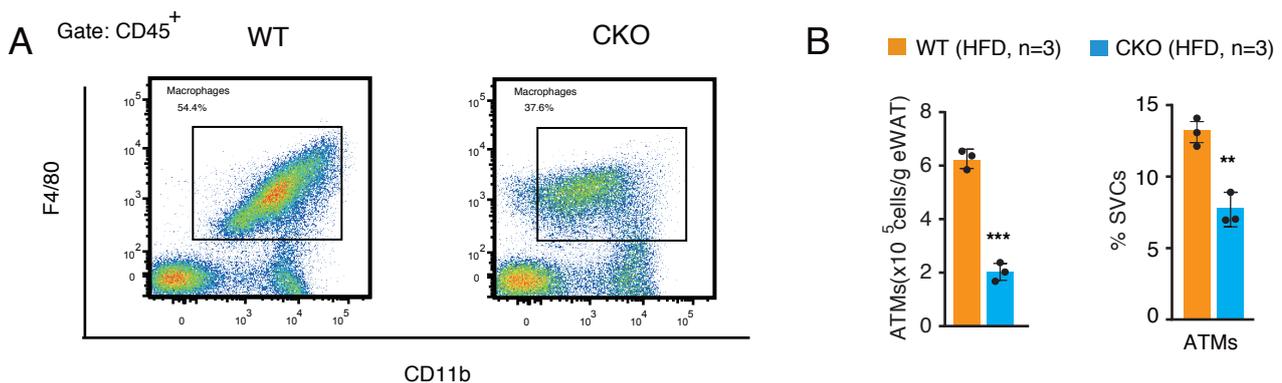
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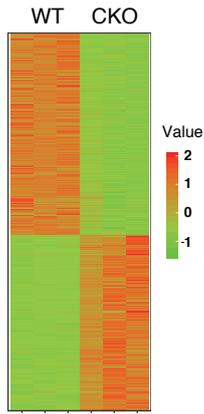
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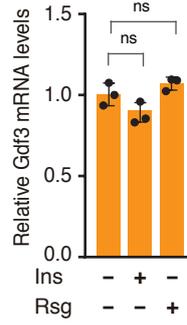
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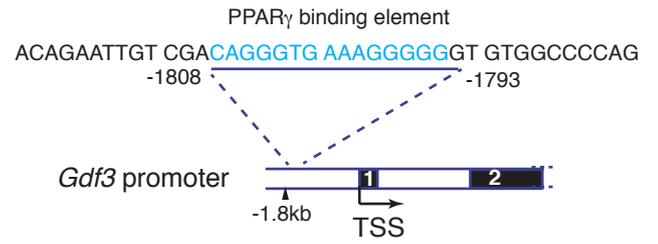
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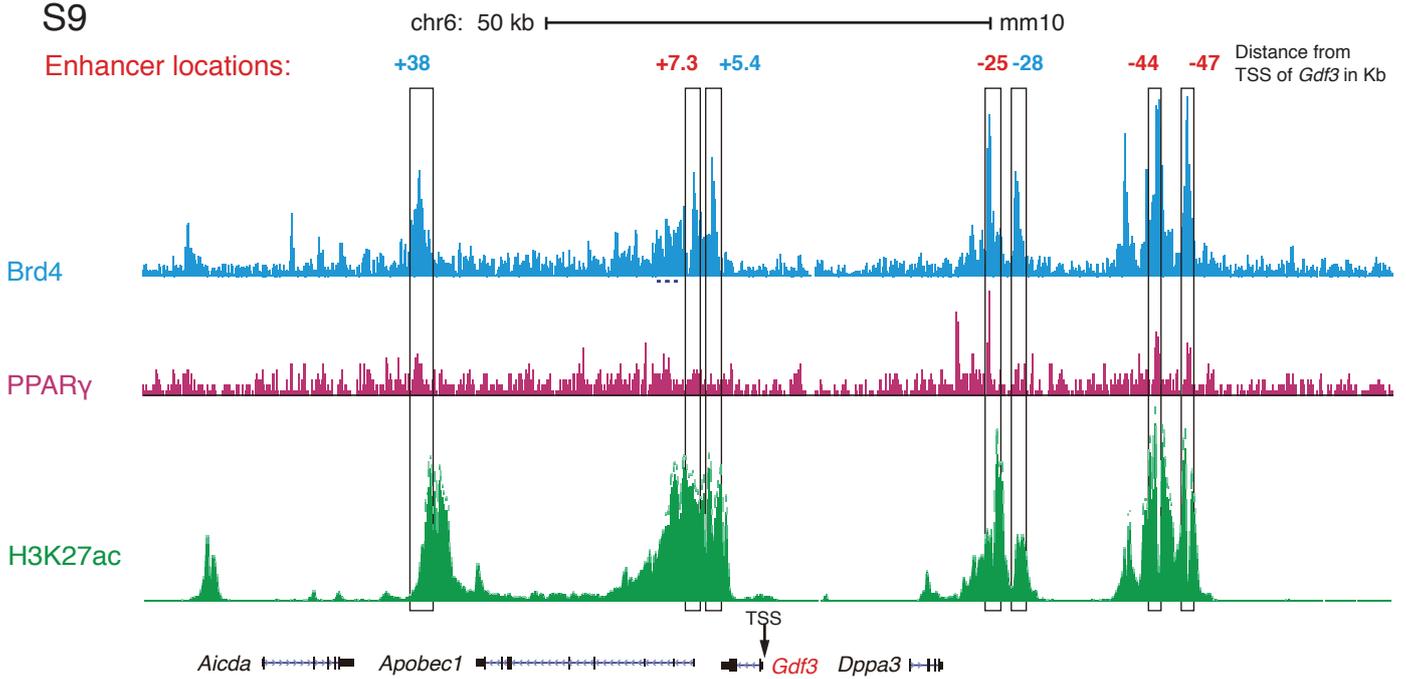
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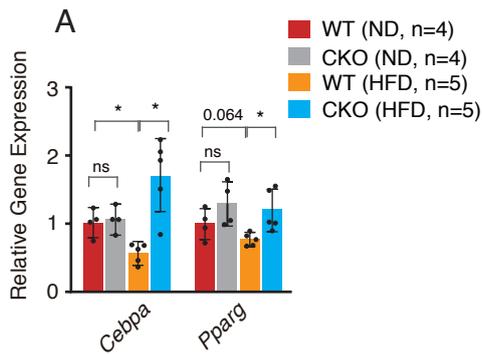
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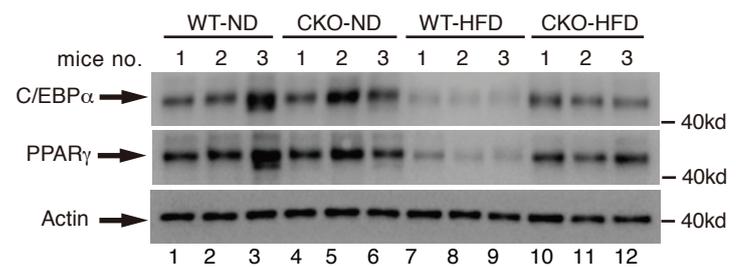
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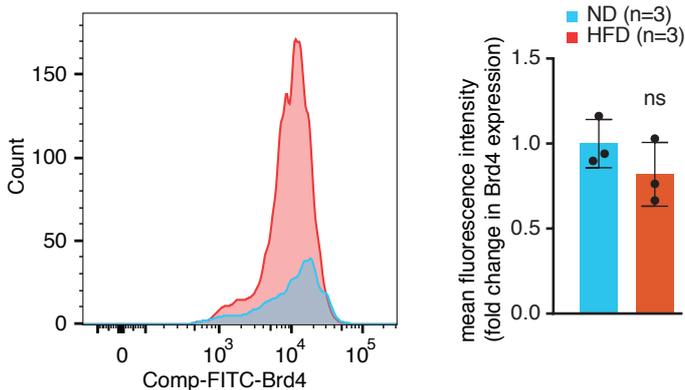
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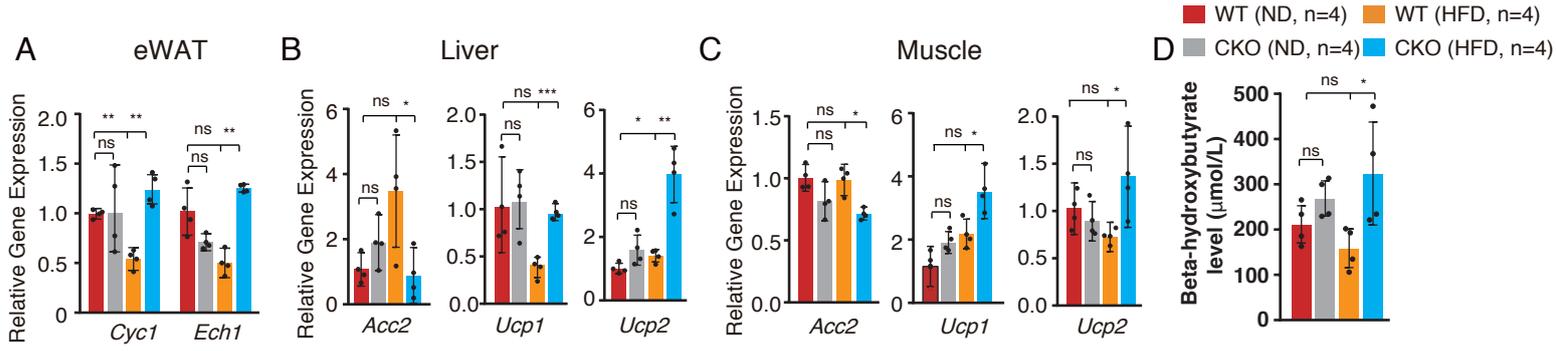
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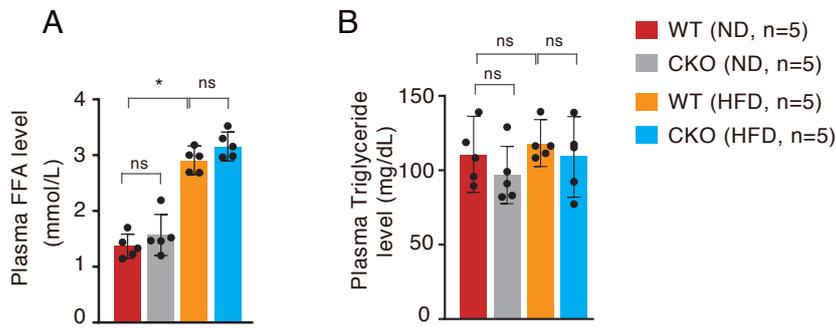
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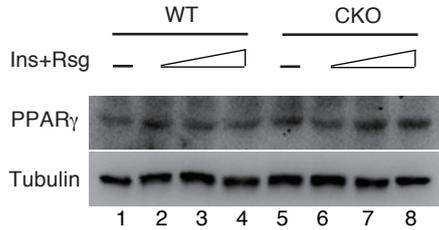
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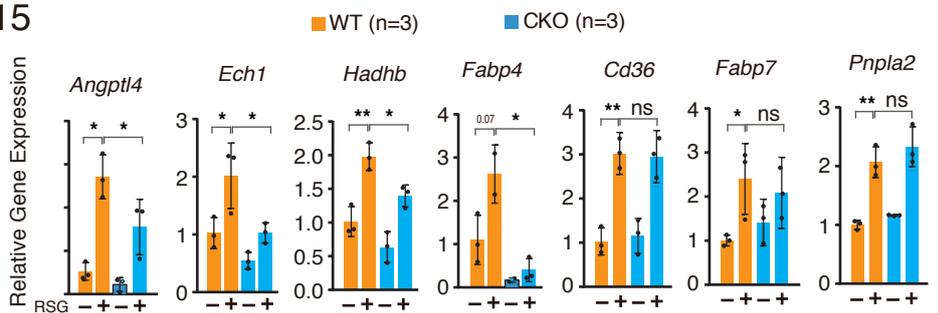
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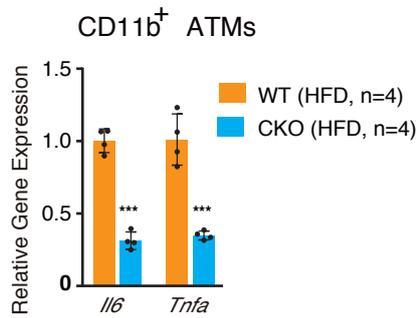
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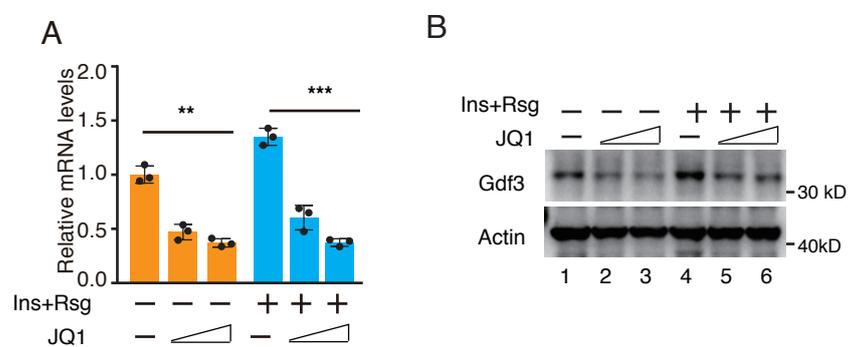


Table 1. Primers used for qRT-PCR

qRT-PCR	Forward (5'-3')	Reverse (5'-3')
<i>Actin</i>	CGGTTCCGATGCCCTGAGGCTCTT	CGTCACACTTCATGATGGAATTGA
<i>Ilf6</i>	TAGTCCTTCCTACCCCAATTTCC	TTGGTCCTTAGCCACTCCTTC
<i>Tnfa</i>	CCCTCACACTCAGATCATCTTCT	GCTACGACGTGGGCTACAG
<i>Mcp1</i>	CCACTCACCTGCTGCTACTCAT	TGGTGATCCTCTTGTAGCTCTCC
<i>Gdf3</i>	CGAGTTTCAAGACTCTGACC	CACGTAGCATAAGTCCTGCG
<i>Atgl/Pnpla2</i>	TGACCATCTGCCTTCCAGA	TGTAGGTGGCGCAAGACA
<i>Hsl</i>	GCGCTGGAGGAGTGTTTTT	CCGCTCTCCAGTTGAACC
<i>Mgll</i>	CGGACTTCCAAGTTTTTGTGAGA	GCAGCCACTAGGATGGAGATG
<i>Cebpa</i>	CAAGAACAGCAACGAGTACCG	GTCACTGGTCAACTCCAGCAC
<i>Pparg</i>	CCAGAGTCTGCTGATCTGCG	GCCACCTCTTTGCTCTGATC
<i>Cyc1</i>	CAGCTTCCATTGCGGACAC	GGCACTCACGGCAGAATGAA
<i>Ech1</i>	GCTACCGCGATGACAGTTTC	TCAGAGATCGAAGGCTGATGTT
<i>Acc2</i>	CGCTCACCAACAGTAAGGTGG	GCTTGGCAGGGAGTTCCTC
<i>Ucp1</i>	AGGCTTCCAGTACCATTAGGT	CTGAGTGAGGCAAAGCTGATTT
<i>Ucp2</i>	ATGTTTGGTTTTCAAGGCCACA	CGGTATCCAGAGGGAAAGTGAT
<i>Angptl4</i>	CATCCTGGGACGAGATGAACT	TGACAAGCGTTACCACAGGC
<i>Hadhb</i>	ACTACATCAAATGGGCTCTCAG	AGCAGAAATGGAATGCGGACC
<i>Fabp4</i>	AAGGTGAAGAGCATCATAACCT	TCACGCCCTTTCATAACACATTCC
<i>CD36</i>	ATGGGCTGTGATCGGAACTG	GTCTTCCAATAAGCATGTCTCC
<i>Fabp7</i>	GGACACAATGCACATTCAAGAAC	CCGAACCACAGACTTACAGTTT

Table 2. Primers used for CHIP-qPCR

CHIP	Forward (5'-3')	Reverse (5'-3')
<i>Gdf3</i> (Promoter)	TCTCAACTAAAGCTAGAACC	GACAATTCTGTGCTTTTCCAC
<i>Gdf3</i> (-25k Enhancer)	GCTCTGTAGATCTCCCAAGGTTG	CTGTGGTGAGGCAGCAAGTCATG
<i>Gdf3</i> (+5.4k Enhancer)	GTGCACGCCTTTAATCCCAGCAC	GTGTTCTGCAAACACCTCTCAC