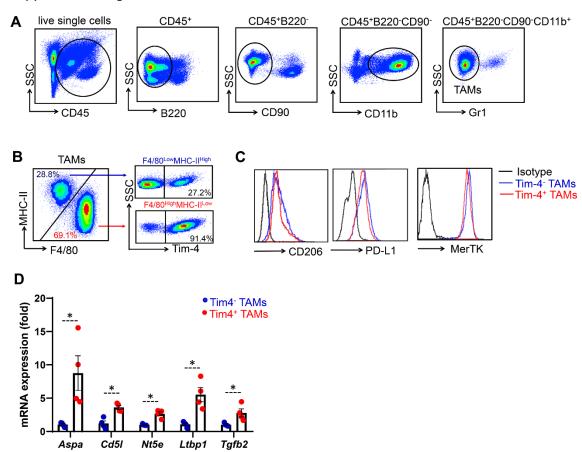
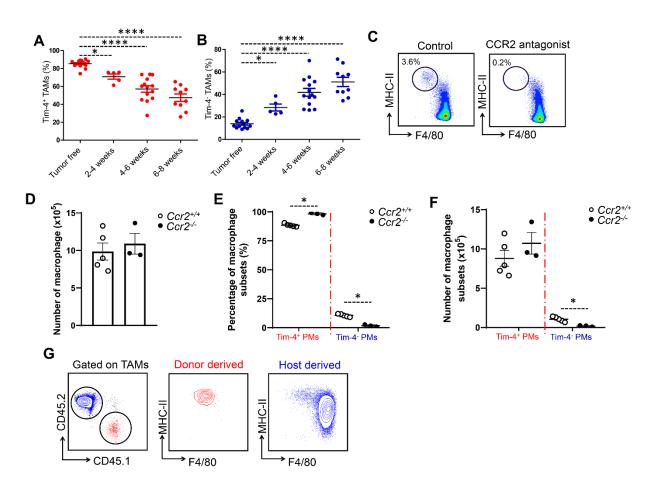
Supplemental data

Supplemental Figure 1.



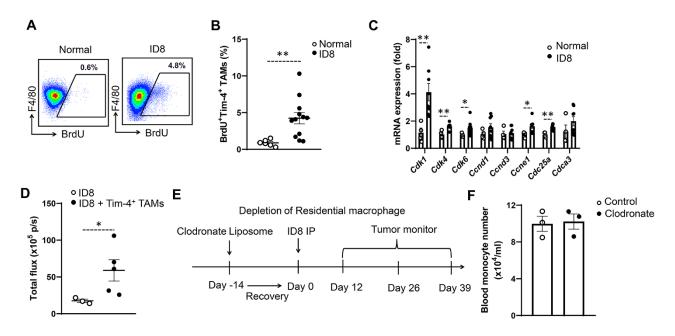
Supplemental Figure 1. Tim-4 defines two distinct peritoneal macrophage subsets in ovarian cancer. (A) Representative flow cytometry plots showing the gating strategy to define peritoneal TAMs in mice bearing peritoneal ID8 ovarian cancer. (B) Relationship between Tim-4 $^+$ TAMs and F4/80 and MHC-II expressing TAMs. One of 10 is shown. (C) Expression of CD206, PD-L1, and MerTK on Tim-4 $^+$ and Tim-4 $^-$ TAMs. One representative of 3 is shown. (D) mRNA expression of GATA6 down-stream genes in Tim-4 $^+$ and Tim-4 $^-$ TAMs (n=4 mice/group, mean \pm SEM) * P < 0.05 (Mann-Whitney U tests).

Supplemental Figure 2.



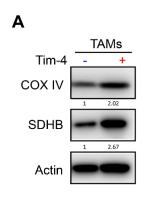
Supplemental Figure 2. Tim-4⁻ TAMs migrate from peripheral monocytes without affecting tumor growth. (A and B) Dynamic changes of Tim-4⁺ (A) and Tim-4⁻ peritoneal TAMs (B) during ovarian cancer progression. TAMs were analyzed by FACS at different time points. (n = 5-14 mice/group, mean \pm SEM). * P < 0.05, **** P < 0.0001(one-way ANOVA with Dunnett's multiple comparisons test) between normal mice and mice bearing ID8 tumor at Week 2-4, 4-6, and 6-8. (C) Effect of CCR2 antagonist on peritoneal macrophages in tumor free mice (n = 5). (D) Number of peritoneal macrophage in wild type and $Ccr2^{-/-}$ tumor free mice (n = 3-5 mice/group, mean \pm SEM). P > 0.05. (E and F) Percentage (E) and number (F) of Tim-4⁺ and Tim-4⁻ peritoneal macrophages (PMs) in wild type and $Ccr2^{-/-}$ tumor free mice (n = 3-5 mice/group, mean \pm SEM). * P < 0.05 (Mann-Whitney U tests). (G) Representative flow cytometry plots of the monocyte derived TAMs. CD45.1⁺ monocytes were transferred into ID8 tumor-bearing mice. CD45.1⁺ monocyte derived TAMs (red) were analyzed with F4/80 and MHC-II expression. One representative of 4 is shown.

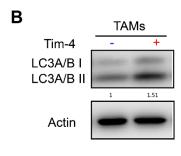
Supplemental Figure 3.



Supplemental Figure 3. Tim-4* TAMs are embryonically derived proliferative cells with pro-tumor function. (A and B) Representative flow cytometry showing the percentages of BrdU+ (A) and BrdU+Tim-4+ (B) peritoneal residential macrophages. Peritoneal macrophages were from tumor-free and ID8 tumor bearing mice (n = 6-12 mice/group, mean \pm SEM). ** P < 0.01 (Mann-Whitney U tests). (C) mRNA expression of cell cycle related genes in Tim-4+ TAMs normalized to Tim-4+ peritoneal residential macrophage (n = 4-8 mice/group, mean \pm SEM). ** P < 0.01 (Mann-Whitney U tests) on the expression of Cdk1, Cdk4, Cdc25a. * P < 0.05 (Mann-Whitney U tests) on the expression of Cdk6, Ccne1. (D) Effect of Tim-4+ TAMs on ID8 growth. Wild type mice were inoculated (I.P) with ID8 cells or ID8 cells plus Tim-4+ TAMs. Tumor volume was shown at day 27 (n = 3-5 mice/group, mean \pm SEM). * P < 0.05 (Mann-Whitney U tests). (E) Scheme of experimental procedure, including peritoneal residential macrophage depletion and tumor inoculation. (F) Number of blood monocytes in tumor free mice treated with control liposome or clodronate liposome, P > 0.05 (n = 3 mice/group, mean \pm SEM).

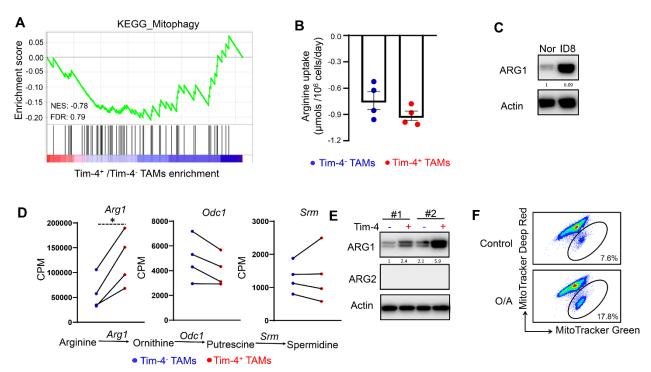
Supplemental Figure 4.





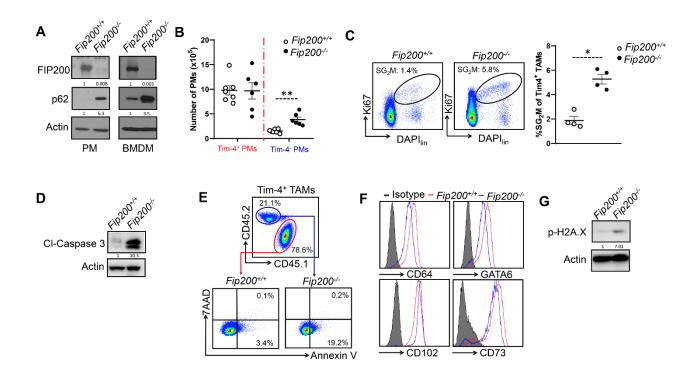
Supplemental Figure 4. Tim-4⁺ TAMs exhibit and maintain high mitochondria activity and autophagy function. (A) Expression of mitochondria related proteins in Tim-4⁺ and Tim-4⁻ TAMs. One representative of 3 is shown. (B) LC-3 in Tim-4⁺ and Tim-4⁻ TAMs. One representative of 3 is shown.

Supplemental Figure 5.



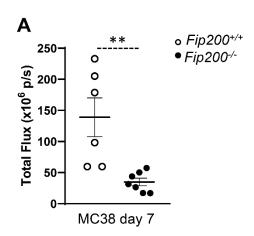
Supplemental Figure 5. Arginase-1 impacts mitochondria fitness and mitophagy via mTORC1 in Tim-4⁺ TAMs. (A) Enrichment of mitophagy gene set in Tim-4⁺ TAMs compared with Tim-4⁻ TAMs. Normalized enrichment score (NES), false discovery rate (FDR). FDR < 0.25 is considered significant. (B) Measurement of arginine uptake in Tim-4⁻ TAMs and Tim-4⁺ TAMs. P > 0.05 (n = 4 mice/group, mean \pm SEM). (C) Western blot showing arginase-1 expression in Tim-4⁺ PMs and Tim-4⁺ TAMs. One representative of 3 is shown. (D) Expression of Arg1, Odc1, and Srm in Tim-4⁻ TAMs and Tim-4⁺ TAMs based on RNA sequencing data. The paired Tim-4⁺ and Tim-4⁻ TAMs were from same mice. (n = 4 mice/group). * P < 0.05 (Mann-Whitney U tests). (E) Western blot showing arginase 1 and arginase 2 expression in Tim-4⁻ TAMs and Tim-4⁺ TAMs. One representative of 3 is shown. (F) Representative flow cytometry plots showing damaged mitochondria in Tim-4⁺ TAMs treated with or without oligomycin (10 µM) plus antimycin A (4 µM) for 24 hours. One of 4 is shown.

Supplemental Figure 6.



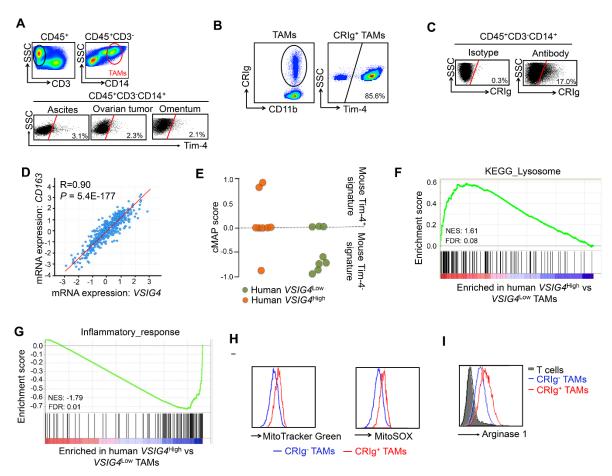
Supplemental Figure 6. Autophagy deficiency results in loss of Tim-4⁺ TAMs in ovarian cancer. (A) FIP200 and p62 protein expression in Tim-4⁺ peritoneal macrophage (PM) and bone marrow derived macrophage (BMDM) from *Fip200*⁺⁺ and *Fip200*⁺⁺ tumor-free mice. One of 3 is shown. (B) Effect of FIP200 deficiency on mouse peritoneal macrophage (PM) subsets. Tim-4 expression was determined by FACS (*n* = 6 to 7 mice/group, mean ± SEM). ** *P* <0.01 (Mann-Whitney *U* tests) for Tim-4⁻ PMs between *Fip200*⁺⁺ and *Fip200*⁺⁻ tumor-free mice. (C) Percentage of Tim-4⁺ TAMs with self-renewal in *Fip200*⁺⁻ and *Fip200*⁻⁻ and *Fip200*⁻ and *Fip200* and

Supplemental Figure 7.



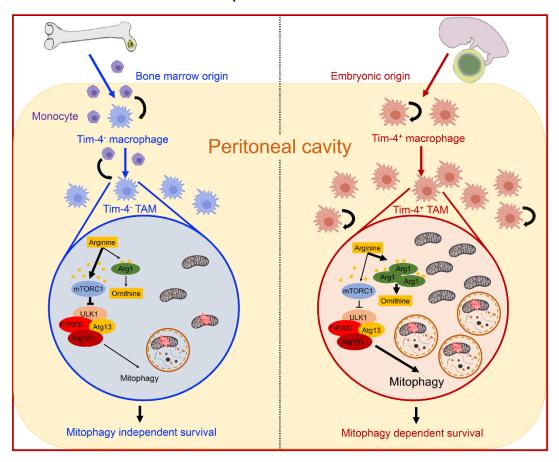
Supplemental Figure 7. Autophagy deficiency in macrophages supports T cell-mediated anti-tumor immunity. (A) Effect of macrophage FIP200 deficiency on peritoneal colon cancer growth. Wild type ($Fip200^{+/+}$) and FIP200 deficient ($Fip200^{-/-}$) mice were inoculated (I.P) with luciferase expressing MC38 colon tumor cells. Tumor volume was monitored. (n = 6 to 7 mice/group, mean \pm SEM) ** P < 0.01 (Mann-Whitney U tests).

Supplemental Figure 8.



Supplemental Figure 8. CRIg⁺ TAMs are the murine counterparts of Tim-4⁺ TAMs in human ovarian cancer. (A) Tim-4 expression on human TAMs. The surface expression of Tim-4 was analyzed by FACS on CD45⁺CD3⁻CD14⁺ TAMs from ascites, ovarian cancer tissues, and ovarian cancer associated omentum. One representative of 3 is shown. (B) Relationship between Tim-4 and CRIg expression on mouse TAMs. Expression of Tim-4 and CRIg was analyzed by FACS on peritoneal TAMs in ID8 tumor bearing mice. One representative of 4 is shown. (C) CRIg expression on human TAMs. The surface staining of CRIg on human TAMs from ovarian cancer associated omentum in patients with ovarian cancer. One representative of 3 is shown. (D) Scatterplot showing Pearson's correlation between *CD163* and *VSIG4* expression in human high grade serous ovarian serous carcinoma. Red line indicates regression fit. (E) cMap analysis of human ovarian cancer related *VSIG4*^{High} and *VSIG4*^{Low} TAMs showing their enrichment for mouse Tim-4⁺ and Tim-4⁻ TAMs gene sets established in (Figure 1E). (F) Enrichment of lysosome gene set in *VSIG4*^{High} TAMs compared with *VSIG4*^{Low} TAMs. Normalized enrichment score (NES), false discovery rate (FDR). FDR < 0.25 is considered significant. (G) Enrichment of inflammatory response gene set in *VSIG4*^{Low} TAMs compared with *VSIG4*^{High} TAMs. Normalized enrichment score (NES), false discovery rate (FDR). FDR < 0.25 is considered significant. (H) Mitochondrial mass and mitochondria related ROS in human ovarian CRIg⁺ and CRIg⁻ TAMs. One representative of 3 is shown. (I) Arginase-1 expression in human ovarian CRIg⁺ and CRIg⁻ TAMs. One representative of 3 is shown.

Graphical Abstract



Graphical abstract

Tim-4⁺ and Tim4⁻ TAMs in ovarian cancer-associated peritoneal metastasis. Tim-4⁺ TAMs were embryonically originated and locally sustained, and promoted tumor progression; while Tim-4⁻ TAMs replenished from circulating monocytes. Relative to Tim-4⁻ TAMs, Tim-4⁺ TAMs expressed high level of mitochondria activity and ROS production, which forced them adapt to mitophagy to alleviate oxidative stress. High arginase-1 in Tim-4⁺ TAMs supported mitophagy formation via weakened mTORC1 activation due to low arginine resultant from arginase-1-mediated arginine metabolism. Tim-4⁺ TAMs, but not Tim-4⁻ TAMs, depended on mitophagy to survive in the tumor microenvironment. Autophagic adaptation to oxidative stress alters peritoneal residential macrophage phenotype and ovarian cancer progression

Supplementary Table 1. Analysis of RNA-Sequencing data related to mouse TAM subsets

Supplementary Table 2. Lists of gene signatures, flow antibodies and primers