

Supplementary Figure 1. Expression of different markers in human scleroderma samples. Chromatin

accessibility changes in human scleroderma samples.

Representative images of human scleroderma skin sections stained against different markers.



Supplementary Figure 2. Jun expands FSP1+ fibroblasts throughout the skin and the underlying tissue. (A) Genetic modifications of the JUN-inducible mouse. rrTA = reverse tetracycline transactivator, TetOP = Tetracycline/doxycycline-responsive operator

(B) Representative Trichrome-stained whole skin sections without (-JUN) and with JUN induction (+JUN). D = Dermis, scF = subcutaneous fat, M = subcutaneous muscle, sc CT = subcutaneous connective tissue, dF = dermal fat, Hf = Hairfollicle. Scale bar = 500 μ m.

(C) Whole skin section after Jun induction (corresponding to the section B). Green = FSP1+, blue = DAPI. D = Dermis, M = subcutaneous muscle, sc CT = subcutaneous connective tissue, dF = dermal fat, Hf = Hairfollicle (D) Quantification of FSP1+ fibroblasts in the upper dermis, the lower dermis and the subcutaneous connective tissue. Indicated are the percentages of FSP1+ cells among all spindle-shaped fibroblasts. Turkey's multiple comparisons test. ** p < 0.01 *** p < 0.001. Scale = 500 µm. n=3. Bar graphs represent means with standard deviations.

(E) Representative stains against FSP1 in the upper and lower dermis and the subcutaneous connective tissue after Jun induction.



Supplementary Figure 3. Skin immune filtration under JUN induction.

(A) Representative IHC stain against pJUN. Scale bar = $200 \,\mu m$.

(B) Representative IHC stain against pJUN. Scale bar = $200 \mu m$.

(C) Quantification of CD45+ cells under JUN induction over up to 5 days. Turkey's multiple comparisons test. *

p < 0.05. n=4-5. Bar graphs represent means with standard deviations.

(D) Quantification of myeloid CD11b+ cells under JUN induction over up to 5 days. Fisher's multiple

comparisons test. n=4-5. Bar graphs represent means with standard deviations.

(E) Quantification of dendritic CD11b+CD11c+ cells under JUN induction over up to 5 days. Turkey's multiple

comparions test. * p < 0.05. n=4-5. Bar graphs represent means with standard deviations.

(F) Quantification of CD3+ T cells under JUN induction over up to 5 days. Turkey's multiple comparisons test.

* p < 0.05. n=4-5. Graph bars represent means with standard deviations.

(G) Quantification of hematopoietic Cx3cr1+ cells under JUN induction over up to 5 days. Fisher's multiple

comparisons test. n=4-5. Graph bars represent means with standard deviations.

(H) In vitro collagen 1 secretion of fibroblasts and macrophages +/- JUN induction. One-way ANVOA * p <

0.05 *** p < 0.001 (n=4). Bar graphs represent means with standard deviations.



Supplementary Figure 4. Fibroblast gating strategy.

After identifying cells and then single cells, hematopoietic (CD45+) and dead cells (PI) are excluded. In a next step, macrophages (F4/80+) and endothelial (CD31+) cells are excluded. After removing epithelial (CD326+) cells, fibroblasts are divided, based on their expression of CD26 and Sca1.



Supplementary Figure 5. JUN initially expands CD26+ fibroblasts.

(A) Quantification of CD26+ fibroblasts, over five days of JUN induction. Turkey's multiple comparisons test. * p < 0.05 ** p < 0.01. n=4-5. Bar graphs represent means with standard deviations.

(B) Quantification of DPF over five days of JUN induction. Turkey's multiple comparisons test. *** p < 0.001.

n=5. Bar graphs represent means with standard deviations.

(C) Quantification of Sca1+ fibroblasts over five days of JUN induction. Turkey's multiple comparisons test. **

p < 0.01*** p < 0.001. n=5. Bar graphs represent means with standard deviations.

(**D**) Quantification of CD26+ fibroblasts and DP fibroblasts (DPF) over up to 10 days of JUN induction. Fisher's multiple comparisons test. * p < 0.05 ** p < 0.01 *** p < 0.001. n=4-6. Bar graphs represent means with standard deviations.

(E) Representative immunofluorescence stains against Caspase3 and CD26 two and seven days after JUN induction. Scale bar = $100 \mu m$. (F)

Representative immunofluorescence stain against CD26 and corresponding EdU visualization two and seven days after JUN induction.



Supplementary Figure 6. CD26+ fibroblasts activate hedgehog signaling.

Normalized qPCR data from facs purified fibroblast populations from non JUN-induced JUN mice. The values for each gene are compared to the mean value of the DN fibroblasts.

(A) Gli1. Turkey's multiple comparisons test. *** p < 0.001. n = 4. Bar graphs represent means with standard deviations.

(B) Gli2. Turkey's multiple comparisons test. n=3-5. Bar graphs represent means with standard deviations.

(C) Gli3. Turkey's multiple comparisons test. n=3-5. Bar graphs represent means with standard deviations.

(D) Ptch1. Turkey's multiple comparisons test. ** p < 0.01 *** p < 0.001. n=3-5. Bar graphs represent means with standard deviations.

(E) Kif7. Turkey's multiple comparisons test. * p < 0.05. n=3-4. Bar graphs represent means with standard deviations.

(F) Smo. Turkey's multiple comparisons test. n=4-6. Bar graphs represent means with standard deviations.



Supplementary Figure 7. PDL1 inhibition eliminates ectopic fibroblasts.

(A) Immunofluorescence stains against PDL1 after local JUN induction in skin after 14 days

(B) PDL1 expression in different subsets of fibroblasts with and without JUN induction. Turkey's multiple

comparisons test. ** p < 0.01 *** p < 0.001. n=3. Bar graphs represent means with standard deviations.

(C) Representative optical images of ectopically transplanted JUN inducible fibroblasts +/- PDL1 inhibition.

n=4. (D) Corresponding quantification of photon emissions. Turkey's multiple comparisons test. * p < 0.05. n=4.

Bar graphs represent means with standard deviations.

(E) Immunofluorescence stains against PD1 and CD11b on macrophages harvested from the peritoneum.

(F) FACS plots of PD1 expression in CD45+CD11b+ blood cells. n=2. Graph bars represent means with standard deviations.



Supplementary Figure 8. CD47 inhibition increases phagocytosis of dermal fibroblasts in vitro. Images are taken with a confocal microscope. RFP+ target cells are detected in the PE channel. Macrophages who have fully digested target cells are marked by small isolated DNA pieces. Boxes in the DAPI represent macrophages with additional DNA pieces as signs of advanced phagocytosis.



Supplementary Figure 9. Combining CD47 and IL6 inhibition prevents loss in subcutaneous fat tissue.

(A) Experimental outline

(B) Representative H&E and Trichrome skin stains of untreated mice, mice under CD47/PDL1 inhibition and vismodegib, and mice under CD47/IL6 inhibition. Scale = $500 \mu m$. Bar graphs represent means with standard deviations.

(C) Thickness of the dermal fibrotic/connective tissue in μ m. Fisher's multiple comparisons test. n=4-7. Bar graphs represent means with standard deviations.

(**D**) Area of the fibrotic tissue in untreated and treated samples, values indicate $\mu m^2/\mu m$ skin width. Fisher's multiple comparisons test. n=4-7. Bar graphs represent means with standard deviations.

(E) Percentage of dermal fat, compared to the overall dermal area, in treated and untreated samples. Turkey's multiple comparisons test. * p < 0.05. n=4-7. Bar graphs represent means with standard deviations.

(F) Area of dermal fat tissue in untreated and treated samples, values indicate $\mu m^2/\mu m$ skin width. Turkey's multiple comparisons test. * p < 0.05. n=4-7. Bar graphs represent means with standard deviations.

(G) Representative stains against Ki67 and CD3. Counterstains with DAPI.

(H) Quantification of CD3+ and Ki67+ stains. Indicated are the number of positive cells/high power view (63x). Turkey's multiple comparisons test. * p < 0.05 ** p < 0.01 *** p < 0.001. n=8-20. Bar graphs represent means with standard deviations.

(I) Representative pictures of CD11b+ cells in skin fibrosis +/- CD47/IL6 inhibition.

(J) Quantification of macrophage agglomerates determined by more than 20 macrophages/High power view in each sections. Two-sided t-test * p < 0.05. n=8. Bar graphs represent means with standard deviations...



Supplementary Figure 10. Immune infiltrate in the therapeutic study.

(A) Representative immunofluorescence stains against CD3 and CD11b in the three groups (Untreated, IL6 inhibition only and CD47/IL6 inhibition)

(B) Quantification through flow cytometry for CD45+ cells. Turkey's multiple comparisons test. n=4. Bar graphs represent means with standard deviations.

(C) Quantification through flow cytometry for myeloid CD11b+ cells. Turkey's multiple comparisons test. n=4. Bar graphs represent means with standard deviations.

(D) Quantification for CD3+ T cells. Turkey's multiple comparisons test. n=4. Bar graphs represent means with standard deviations.

(E) Corresponding organ sections from the untreated and the CD47/IL6 inhibition group. Flow cytometry numbers represent number of cells/100,000 live cells.

FACS Antibodies

| Antigen | Manufacturer | Catalog # | Clone | Conjugatio n |
|---------------------------|--------------|-----------|----------|-----------------|
| CD3 | Biolegend | 100209 | 17A2 | APC |
| CD4 | Biolegend | 100422 | GK1.5 | PE-Cy7 |
| CD11b | BD | 553311 | M1/70 | PE |
| CD11c | Biolegend | 117324 | N418 | APC-Cy7 |
| CD25 | Biolegend | 102035 | PC61 | BV605 |
| CD26 | Biolegend | 137805 | H194-112 | FITC |
| CD31 | BD | 553373 | MEC 13.3 | PE |
| CD45 | Biolegend | 103110 | 30-F11 | Pe-Cy5 |
| CD47 | Biolegend | 127527 | Miap301 | BV421 |
| CD326 | Biolegend | 118218 | G8.8 | APC-Cy7 |
| F4/80 | Biolegend | 123116 | BM8 | APC |
| PDL1 | Biolegend | 124312 | 10F.9G2 | APC |
| Phospho c- Jun (Ser73) | CST | 32705 | D47G9 | Rabbit |
| Sca1 | Biolegend | 108114 | D7 | Pe-Cy7 |
| PI | Biolegend | 421301 | | |

Immunofluorescence/Immunohistochemistry antibodies

| Antigen | Manufacturer | Catalog # | Clone | Dilution |
|---------------------------|----------------|------------|--------|----------|
| Phospho c- Jun (Ser73) | CST | 32705 | D47G9 | 1:100 |
| Collagen 1 | Abcam | ab34710 | | 1:100 |
| CD47 | R&D | AF1866 | | 1:40 |
| CD47 | ThermoFisher | 14-0479-82 | B6H12 | 1:50 |
| PD1 | Cell marque | 315M-96 | NAT105 | 1:100 |
| PD1 | R&D | AF1021 | | 1:40 |
| CD68 | Agilent | GA60961-2 | KP1 | 1:200 |
| CD31 | Dako | m0823 | | 1:150 |
| Adiponectin | Abcam | ab22554 | | 1:100 |
| CD26 | Abcam | ab28340 | | 1:100 |
| CD26 | R&D | AF954 | | 1:40 |
| PDL1 | R&D | AF1019 | | 1:40 |
| FSP1 | MilliporeSigma | 07-2274 | | 1:200 |

Western Blot antibodies

| Antigen | Manufacturer | Catalog # | Clone | Dilution |
|----------------------------|----------------|-----------|---------|----------|
| Phospho c- Jun (Ser73) | CST | 32705 | D47G9 | 1:1000 |
| c-Jun | CST | 91655 | 10F.9G2 | 1:1000 |
| Phospho- Stat3 (Tyr105) | CST | 91315 | | 1:1000 |
| GAPDH | GeneTex | GTX627408 | GT239 | 1:50,000 |
| FSP1 | MilliporeSigma | 07-2274 | | 1:1000 |

Primer sequences for qPCR

| Gene | Species | | Primer sequence |
|------------|---------------|---------------------------|-------------------|
| | | | TGGGATGAAGAAGCAG |
| Gli1 | Mouse | mGli1 Forward | TTGGG |
| | | | TTGAACATGGCGTCTCA |
| | | mGli1 Reverse | GGG |
| | | | GCCTCTGAGATGGAGA |
| Gli2 | Mouse | mGli2 Forward | CTTCTG |
| | | | TCATGTCAATCGGCAAA |
| | | mGli2 Reverse | GGC |
| | | | GGAGGGTGTTCTCCTCT |
| Gli3 | Mouse | mGli3 Forward | GAC |
| | | | AGGCCATCACATCCCAA |
| | | mGli3 Reverse | СТС |
| | | | CTCTGGCTCCTAGCACC |
| Actb | Mouse | mActb Forward | ATGAAGA |
| | | | GTAAAACGCAGCTCAGT |
| | | mActb Reverse | AACAGTCCG |
| Smo | Mouso | m ^c mo Forward | GGTTTTAATGGTGGGA |
| 51110 | wouse | IIISIIIO FOIWaru | GAGGGA |
| | | m ^c mo Povorco | GATCGAAGCTGTCTTCA |
| | | IIISIIIO Reverse | ACCC |
| Dtah 1 | Mouro | mDtch1 Forward | ACAAAGCCGACTACATG |
| PICHI | wouse | IIIPICIII FOIWaru | CCA |
| | | mDtch1 Boyorco | TCGTAGGCCGTTGAGGT |
| | | IIIPICITI Reverse | AGA |
| Kif7 Mouse | mKif7 Forward | ATGCCACCGTCTTTGCC | |
| | | | TAT |
| | | mkif7 Povorso | GTAGGATACATGCACCA |
| | | IIIKII/ Nevelse | GGCA |

Primer for Genotyping

| Gene | Primer# | Sequence |
|--------|---------|-----------------------|
| Rosa26 | Rosa A | AAAGTCGCTCTGAGTTGTTAT |
| | Rosa B | GCGAAGAGTTTGTCCTCAACC |
| | Rosac | GGAGCGGGAGAAATGGATATG |
| | | |

| Jun | ColA | GCACAGCATTGCGGACATGC |
|-----|------|----------------------|
| | ColB | CCCTCCATGTGTGACCAAGG |
| | ColC | GCAGAAGCGCGGCCGTCTGG |

Supplementary Table 1. Primary antibodies and Primers.



GAPDH