Supplementary Figures



Figure S1. CLL-plasma derived EVs by Opti-CUC (A) Protein yield per mL starting plasma volume (µg/mL) detected in Opti-CUC EV isolate and the pelleted supernatant for Opti-CUC EV isolates shown in Figure. 1B (n=22, p=0.029, paired t-test), horizontal line represents mean (B) Representative electron microscopy images comparing Opti-CUC EV isolate and the pelleted supernatant. Scale bar 100 nm.



Figure S2. Protein yield (μ g) per mL starting plasma volume (A) and Particle/ μ g value (B) for the 13 plasma pool samples that are averaged in Figure 2B. The plasma pools were isolated by DUC, Opti-CUC or Opti-CUC-Tre.



Figure S3. Isolated particles subjected to nanoparticle tracking analysis (NTA) measurement for concentration and size distribution, a representative plot presented is the average of three 30-second videos for 6 of the plasma pool sets discussed in Figure 2. The plasma pools were isolated by DUC, Opti-CUC or Opti-CUC-Tre.

Bin centre (nm)



Figure S4. Bead-based flow cytometry analysis of EV isolates from the individual plasma pool sets that are averaged in Figure 2D . Delta median fluorescence of samples calculated by subtracting median fluorescence intensity (MFI) of each sample from its isotype control.



А

в



Figure S5. EV isolation from MEC1 and OSU-CLL in standard flasks cultured in AIM V or RPMI (A) Isolated particles subjected to nanoparticle tracking analysis (NTA) measurement for concentration and size distribution, a representative plot presented is the average of three 30-second videos for MEC1 (upper panel) and OSU-CLL (lower panel) for the different media, n=6. Further data for these samples in Figure 3. (B) Viability and total cell count at harvest for MEC1 or OSU-CLL cultured in 100 mL RPMI or AIM V, n=6, paired t-test, data are represented as mean ± SD. RPMI= EV depleted complete RPMI.



Β



Figure S6. Representative Transmission Electron Microscopy (TEM) images with size annotations of MEC1 EVs, scale bar 200 nm. (A) MEC1 cultured in standard flask in RPMI isolated by Opti-CUC. (B and C) CeLLine Flask EVs isolated by Opti-CUC in absence (B) or presence (C) of initial trehalose. RPMI= EV depleted complete RPMI.





Figure S7. Size comparisons of extracellular vesicles (EVs) measured with nanoparticle tracking analysis (NTA) and analyzed by bins of size classes, (n=6) for cells cultured in standard flasks (SF) versus CLF (CeLLine Flask) no trehalose condition. For both flasks, cells were cultured in EV-depleted RPMI and EVs were isolated by Opti-CUC. (A) Segmented Bar Graph showing mean percentage per size bin (B) 2 tailed t-test statistical comparisons between the SF-EVs and the CLF-EVs for each bin, $p \le 0.05$ in bold.

В

Bin nm range		1-100	101-200	201-300	301-400	401-500	501 -
MEC1	SF-EV	1.13	68.65	24.38	4.67	0.98	0.2
	CLF-EV	7.31	79.62	11.16	1.68	0.18	0.04
	t-test	<.0001	0.0194	0.0014	0.0208	0.0211	0.0837
OSU-CLL	SF-EV	2.32	58.06	28.62	7.52	2.67	0.81
	CLF-EV	5.5	74.2	16.56	2.92	0.75	0.07
	t-test	0.0516	0.0028	0.0019	0.0089	0.1093	0.0466

 $p \leqslant \! 0.05$ in bold









Buffer

DUC

Opti-CUC

Opti-CUC-Tre

Figure S9. CLL-plasma pool- derived EVs promote cell proliferation in vitro. A) Comparison of percent proliferation change of HS-5-GFP stromal cells based on green fluorescence readout after incubation with increasing concentrations of EVs derived from CLL-plasma pools at 24hr, 48hr and 72hr. The 72hr data is repeated from figure 8 for comparison. Data reported as percent change normalized to control (PBS/Tre buffer), n=4, data are represented as mean ± SEM. The p-value (mixed effect model) trend analysis are indicated on the graph. B) Percent proliferation change of HS-5 stromal cells after 96 hours of incubation with increasing concentrations of CLL-plasma pool-derived EVs using MTS assay. Data reported as percent proliferation change normalized to control (PBS/Tre buffer), n=6, data are represented as mean ± SEM.

В

А

Percent proliferation to buffer with 5µg/mL EV dose

150

125

100

75

50

25

p=0.006

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p=0.5

T

p=0.0024

4. .

Full unedited gel for Figure 2-C











Full unedited gel for Figure 6-A, Set 1&2





 $I^{AO} = I = I = AIM AME = + -1 = IDECC$ TYCLE TYCLE IS SO100-1 = AIM AME = + -1 = IDECCTYCLE TYCLE IS SO12-13-135

Albumin



TSG101

CD81

CD63

Calnexin

Full unedited gel for Figure 6-A, Set 3&4







