

Supplemental Figure 1. Design and characterization of PR8 HA-ferritin

nanoparticles. (A) Schematic of PR8 HA gene fused to ferritin from Helicobacter pylori.
(B) Size exclusion chromatography (greyed area signifies retained fractions) of HA-ferritin. (C) SDS PAGE showing HA-ferritin with expected molecular weight of 85 kDa.
(D) Negatively stained TEM images of purified HA-ferritin and ferritin nanoparticles confirming expected particle size of 37nm and 14nm, respectively. Scale bar 30nm. (E) Antigenicity of HA-ferritin was confirmed by binding of mAbs specific to the head (TN1F11, 446D1 and TN1B09) and stem (C179, CR9114) of influenza PR8 HA in an ELISA.

Α HA-ferritin 3.8 μ g



HA-ferritin 0.38 μ g



С Soluble HA 3.8 μ g



Soluble HA 0.38 μ g



Е



PBS



Supplemental Figure 2. Confocal microscopy evaluation of germinal centers in lymph nodes. C57Bl/6 (n = 5 mice per group) mice were immunized with (A) high dose HA-ferritin (5 μ g), (B) low dose HA-ferritin (0.5 μ g), (C) molar equivalent of high dose soluble HA (3.8 μ g), (D) low dose soluble HA (0.38 μ g), (E) 1.2 μ g ferritin, all adjuvanted with AddaVax or (F) PBS alone. Draining inguinal lymph nodes were excised, sectioned and stained with antibodies to identify B cells (B220; magenta), germinal centers (GL7; green), follicular dendritic cells (CD35; blue), and class switching (IgD; white).



Supplemental Figure 3. Identification of HA-specific germinal center B cells in mouse lymph nodes. Germinal center cells in lymph node suspensions were gated as singlets, lymphocytes, CD45⁺B220⁺, live, IgD⁻, GL7⁺ using flow cytometry. PR8 HA-Biotin labelled with Streptavidin-PE was used to probe for HA-specific B cells. Representative examples of probe staining in lymph node cells from mice vaccinated with ferritin, soluble HA and HA-ferritin are shown.



Supplemental Figure 4. Frequencies of HA-specific germinal center B cells at early timepoints following vaccination. C57Bl/6 (n = 5 mice per group) mice were immunized with HA-ferritin (5 or 0.5 μ g) or a molar equivalent of soluble HA (3.8 or 0.38 μ g), adjuvanted with AddaVax. At 14 days post vaccination, the absolute count of HA-specific germinal center B cells in draining iliac (left) and inguinal (right) lymph nodes at 7 (A) and 14 (B) days post vaccination was determined using a probe of biotinylated PR8 HA labelled with streptavidin-PE. Data represent mean ± SD and are representative of one experiment. *p <0.05, **p<0.01, determined by a Mann-Whitney test.



Supplemental Figure 5. Identification of T follicular helper (T_{FH}) cells in mouse lymph nodes ex vivo. T_{FH} cells in lymph node suspensions were gated as lymphocytes, singlets, live F4/80⁻, CD3⁺B220⁻, CD4⁺, CXCR5⁺PD-1⁺ using flow cytometry.



Supplemental Figure 6. Absolute counts of T follicular helper (T_{FH}) cells in lymph nodes after vaccination. C57Bl/6 (n = 10 mice per group) mice were immunized with HA-ferritin (5 or 0.5 μ g) or a molar equivalent of soluble HA (3.8 or 0.38 μ g), or 1.2 μ g ferritin alone, adjuvanted with AddaVax. 14 days following vaccination, draining iliac (left) and inguinal (right) lymph nodes were harvested. The count of T_{FH} cells (identified as CD4+CXCR5+PD-1+) per lymph node pair was determined by flow cytometry. Data shown are combined from two independent experiments. Data represent mean ± SD. Significance determined by a Mann-Whitney test.



Supplemental Figure 7. Identification of activated T follicular helper (T_{FH}) cells following in vitro stimulation. (A) Following 18 h stimulation of mouse lymph node suspensions with HA or ferritin peptide pools or DMSO control, T_{FH} cells were gated on as lymphocytes, singlets, live F4/80⁻, CD3⁺B220⁻, CD4⁺, CXCR5⁺PD-1⁺ using flow cytometry. (B) Representative examples of TFH activation following DMSO, ferritin or HA peptide stimulation is shown with gates indicating CD154⁺ or CD25⁺⁺OX40⁺⁺ cells.



Supplemental Figure 8. Stem HA-specific antibody and HAI titers in vaccinated macaques. Pigtail macaques (n = 5) were vaccinated once intramuscularly with HA-ferritin (15 μ g) or a molar equivalent of soluble HA (11.25 μ g), both adjuvanted with AddaVax. A control group received AddaVax alone (n = 2). Sera was collected at 0, 7 and 21 days post vaccination. (A) Serum IgG titers against HA stem were measured by capture ELISA. (B) Serum HAI titers against PR8 virus were measured at 21 days post vaccination. The dashed lines indicate detection cutoff (1:100 dilution for ELISA, 1:20 for HAI assay). Data represent mean ± SD. Significance was determined by a Mann-Whitney test.



Supplemental Figure 9. Identification of activated T follicular helper (T_{FH}) cells in macaque lymph nodes. Activated T_{FH} cells in lymph node suspensions were gated as lymphocytes, singlets, live, CD3+CD4+,CXCR5+PD-1+, Bcl6+Ki67+ using flow cytometry. Representative examples of probe staining from macaques vaccinated with HA-ferritin, soluble HA and AddaVax alone are shown.



Supplemental Figure 10. Identification of germinal center B cells in macaque lymph nodes. Germinal center B cells in lymph node suspensions were gated as lymphocytes, singlets, live, CD3⁻CD4⁻,CD20⁺, IgM⁻IgD⁻, IgG⁺, BcI6⁺Ki67⁺ using flow cytometry. PR8 HA-biotin labelled with streptavidin-PE and streptavidin-BV421 was used to probe for HA-specific B cells. Representative examples of probe staining from macaques vaccinated with HA-ferritin, soluble HA and AddaVax alone are shown.

Supplemental Videos. 3D antigen localization of antigen in the draining lymph nodes. C57Bl/6 mice (n = 2 per group) were immunized with AF647-labelled HA-ferritin (5 μ g) (Video 1) or a molar equivalent of AF647-labelled soluble HA (3.8 μ g) (Video 2), both adjuvanted with AddaVax. 14 days later, draining inguinal lymph nodes were fixed, stained (IgD, yellow; CD35, blue), cleared and imaged by lightsheet microscopy. Co-localization of antigen with CD35⁺ follicular dendritic cells within germinal centers is observed following vaccination with HA-ferritin but not soluble HA.