

Supplemental Figure 1. Myomaker expression in myofibers of dystrophic muscle. X-gal stains of cross-sections of tibialis anterior (TA) and diaphragm muscle from 6-8 month old $mdx Mymk^{LacZ/+}$ mice showing expression of LacZ in dystrophic myofibers, both in concentrated clusters of myofibers as well as scattered in a punctate pattern (inset). Scale bars: 50 µm.



Supplemental Figure 2. The satellite cell population is maintained in *mdx*

Mymk^{scKO} mice. (A) Pax7 immunofluorescence shows persistence of SCs following deletion of Myomaker. Quantification of Pax7⁺/DAPI⁺ cells shows comparable tissue concentration to controls. (B) Myogenin immunofluorescence reveals similar levels of activated myoblasts in *mdx Mymk*^{scKO} mice and controls. Data are represented as mean \pm SD. Scale bars: 20 µm. *n* = 3-6.



Supplemental Figure 3. Satellite cell fusion is required for regeneration in older *mdx* mice. (A) *Mdx* mice at various ages (2 months, 6 months, and 1 year) were injected with BrdU daily for 14 days prior to sacrifice in order to assess fusion rates. Quantification indicates a steep drop in fusion levels between 2 and 6 months (n = 4). (B) Schematic for ablation of Myomaker in SCs of 12 month old *mdx* mice. (C) Gross appearance of *mdx Mymk*^{scKO} mice showed smaller size, frail body habitus, and kyphosis of the spine. (D) Body weights of *mdx Mymk*^{scKO} mice were significantly reduced by 5 months following initial Myomaker deletion. (E) Individual dry muscle weights of tibialis anterior (TA), quadriceps, and gastrocnemius/plantaris/soleus (GPS) show loss of muscle mass. (F) Grip strength was measured on a forelimb meter and showed impaired function in dystrophic mice with Myomaker-deficient SCs. Statistical analyses and data presentation: (A) one-way ANOVA with Tukey post-hoc comparison, **p<0.01. (D-F) unpaired t-test; *p<0.05, **p<0.01, ***p<0.001, ****p<0.0001. Data are represented as mean ± SD. Scale bars: (A) 50 µm; (C) 1 cm. n = 10-13 except where noted.



Supplemental Figure 4. Equivalence of mdx Mymk^{scKO} and mdx SC^{DTA}

phenotypes. (A) Juxtaposition of dry quadriceps weights from *mdx Mymk*^{scKO} and *mdx* SC^{DTA} mice and respective controls (n = 5-11). (B) Comparison of forelimb grip strength measurements from both models (n = 8-11). (C) Comparison of quantification of fibrotic area from Picrosirius Red staining of tibialis anterior (TA) cross-sections (n = 5-6). Statistical analyses and data presentation: (A), (B), (C) one-way ANOVA with Tukey post-hoc comparison, ****p<0.0001. Data are represented as mean ± SD.



Supplemental Figure 5. *Mymk* transcription is downregulated in isolated muscle fibers of *mdx Mymk*^{fiberKO} mice. (A) Schematic showing tamoxifen administration of mice prior to sacrifice and myofiber isolation from the extensor digitorum longus muscle. (B) qPCR for *Mymk* mRNA from isolated muscle fibers (n = 7). (C) qPCR for *Mymk* in isolated mononuclear cells shows persistent expression in *mdx Mymk*^{fiberKO} mice but efficient deletion in *mdx Mymk*^{scKO} mice. Expression is presented relative to control for each model (n = 5-8). Statistical analyses and data presentation: (B) unpaired t-test, **p<0.01. (C) Mann-Whitney test, **p<0.01. Data are represented as mean ± SD.



Supplemental Figure 6. No reduction in muscle damage is observed in vehicletreated *mdx Mymk*^{fiberKO} mice or due to activation of *Acta1*^{CreER} alone. (A) Schematic of treatment with vehicle-only injections. (B) No reduction in IgM⁺ myofibers was seen in cross-sections of vehicle-treated *mdx Mymk*^{fiberKO} quadriceps (rectus femoris) muscle. (C) Schematic of experiment to test the effect of tamoxifen-induced *Acta1*^{CreERT2} activation in *mdx* mice in the absence of floxed Myomaker alleles. (D) *Acta1*^{CreERT2} activation alone does not reduce IgM⁺ myofibers in dystrophic rectus femoris muscle with quantification. Data are represented as mean ± SD. Scale bars: (B), (D) 50 µm.