

1 **Supplementary Materials:**

2 **Methods**

3 *Animal studies*

4 The *Pkd1*^{RC/RC} (*PKD1* p.R3277C) mice, a murine model of ADPKD, were provided by Dr.
5 Peter C. Harris. Wild-type (WT) C57BL/6J mice were purchased from Jackson Labs. PAPP-
6 A deficient mice were generated by crossing PAPP-A deficient mice, *Pappa*^{tm1Cac} (1) and
7 *Pkd1*^{RC/RC} mice. *Pkd1*^{RC/RC} mice were crossed with *Prkar1a*^{f/f}; *Pkhd1*-Cre mice to generate
8 ADPKD mice with kidney-specific over-activation of PKA (2). Five animals per cage were
9 housed in standard cages in a room maintained at constant temperature and humidity, and 12
10 h light/dark cycles.

11 The experimental protocol was reviewed and approved by the Institutional Animal Care and
12 Use Committee at Mayo Clinic (Protocol no. A47715 and A00003864), and studies were
13 conducted in adherence to the NIH Guide for the Care and Use of Laboratory Animals. For
14 IGF-1 antibody experiment, the animals were injected intraperitoneally with 0.2 mg/kg IGF-1
15 antibody or IgG control (R&D Systems) per week for 6 weeks. 5 mg/kg FSK or control (5%
16 DMSO) was administered intraperitoneally for FSK experiment.

17 At the end of the study mice were sacrificed, blood was taken for biochemical analysis, and
18 organs were weighed. Portions of tissue were placed in formalin and processed for
19 histological studies or snap frozen in liquid nitrogen and stored at -80 °C for gene expression
20 analysis.

21 *Real time Polymerase chain reaction*

22 Total RNA was isolated using the RNeasy Plus Mini Kit (Qiagen) and cDNA was prepared
23 using the QuantiTect Reverse Transcription Kit (Qiagen). Commercially available TaqMan
24 gene expression probes were obtained from Applied Biosystems and quantitative real-time

25 PCR was performed in duplicates as described previously (3). The relative mRNA expression
26 of target genes was calculated using the $2^{-\Delta\Delta Cq}$ method with *Gapdh* as internal reference gene.

27 *Serum IGF-1 levels*

28 Serum was collected from C57BL6 WT and *Pkd1*^{RC/RC} mice and stored at -80° C. The
29 amounts of IGF-1 in the serum were measured using the Ultra-Sensitive Mouse IGF-1 ELISA
30 kit from Crystal Chem, Inc., according to the manufacturer instructions.

31 *cAMP ELISA*

32 cAMP levels were measured in renal tissues of WT and *Pkd1*^{RC/RC} -*Pappa* mutant mice using
33 Direct cAMP ELISA kit from Enzo Life Sciences, Inc. kidneys were homogenized in 0.1M
34 HCl and supernatant was used for assay. Protein was measured using Bradford assay.

35 *Cell culture*

36 RCTE and 9-12 cells were cultured in DMEM:F12 with 10% FBS and 1%
37 penicillin/streptomycin. Cells were treated with 10 μ M FSK in presence or absence of IL1 β
38 (5 ng/ml), IL-2 (2 ng/ml), EGF (10 ng/ml, TGF β (1 ng/ml), 0.5 μ M ionomycin and 0.2 μ M
39 verapamil for 16 hour and RNA was isolated. Cells were treated with 10 μ M FSK in presence
40 or absence of budesonide (150 μ M) or decitabine (0.5 μ M) for 16 hours followed by RNA
41 isolation. Cells were pre-treated with Budesonide (150 μ M) or decitabine for 24 hours before
42 adding FSK. The cells were treated with FSK for 16 hours in presence or absence of
43 compound C (10 μ M) or ERK inhibitor, SCH772984 (2 μ M) which were added to the media
44 after 1 hour of FSK treatment and RNA was isolated. The recombinant PAPP-A (4) was used
45 in pathway studies.

46 Madin-Darby canine kidney (MDCK) cells were maintained in DMEM/F12 (Life
47 Technologies) supplemented with 10% fetal bovine serum (FBS) and penicillin and
48 streptomycin (Invitrogen). Cystogenesis studies with MDCK cells were performed essentially
49 as previously described (5). Briefly, cells were suspended in collagen I solution, seeded in

50 24-well plates at 4,000 cells per well, and incubated at 37 °C for 30-45 min. for collagen to
51 polymerize. Cells were then given medium (DMEM/F12, 1% FBS) containing the cAMP
52 agonist forskolin (FSK, 10 µM) and IGF-1 (50 ng/ml). Cells were cultured for 14 days, with
53 media change every other day. At the end of the study, the number of cysts in each well was
54 counted.

55 *PKA kinase activity assay*

56 RCTE and 9-12 cells were treated with or without FSK for 24 hours and protein was
57 extracted with 1 ml lysis buffer (20 mM MOPS, 50 mM β-glycerolphosphate, 50 mM sodium
58 fluoride, 1 mM sodium orthovanadate, 5 mM EGTA, 2 mM EDTA, 1% NP40, 1 mM
59 dithiothreitol (DTT) and protease inhibitor cocktail). The PKA kinase activity was measured
60 using 2 µg of extracted protein using PKA kinase assay kit following manufacturer's
61 instructions (Enzo Life Sciences, Inc.)

62 *Metanephros organ culture*

63 Metanephros organ culture was performed as previously described(6) (7) except the addition
64 of insulin to the media. Embryos were aseptically isolated from C57BL/6J or *Pkd1*^{RC/RC} mice
65 at embryonic day 12 – 13 (E12 - 13). Metanephroi from embryos were collected and placed
66 onto a transparent Falcon 0.4 µm cell culture insert floating on a defined serum-free medium.
67 The metanephroi were incubated for 0-14 days in 12-well Falcon tissue culture plates (Fisher
68 Scientific, CA) at 37 °C in a humidified incubator (5% CO₂). Defined medium was
69 DMEM/F12 supplemented with 5 µ /ml transferrin, 2.8 nM selenium, 25 ng/ml prostaglandin
70 E1, 6 ng/ml triiodothyronine (T3) and 1% penicillin and streptomycin. IGF-1 was added to
71 the medium for C57BL/6J. Growth medium was changed every other day. Cysts were
72 generated by treating with forskolin (FSK) and the development of cysts was observed daily
73 under microscope.

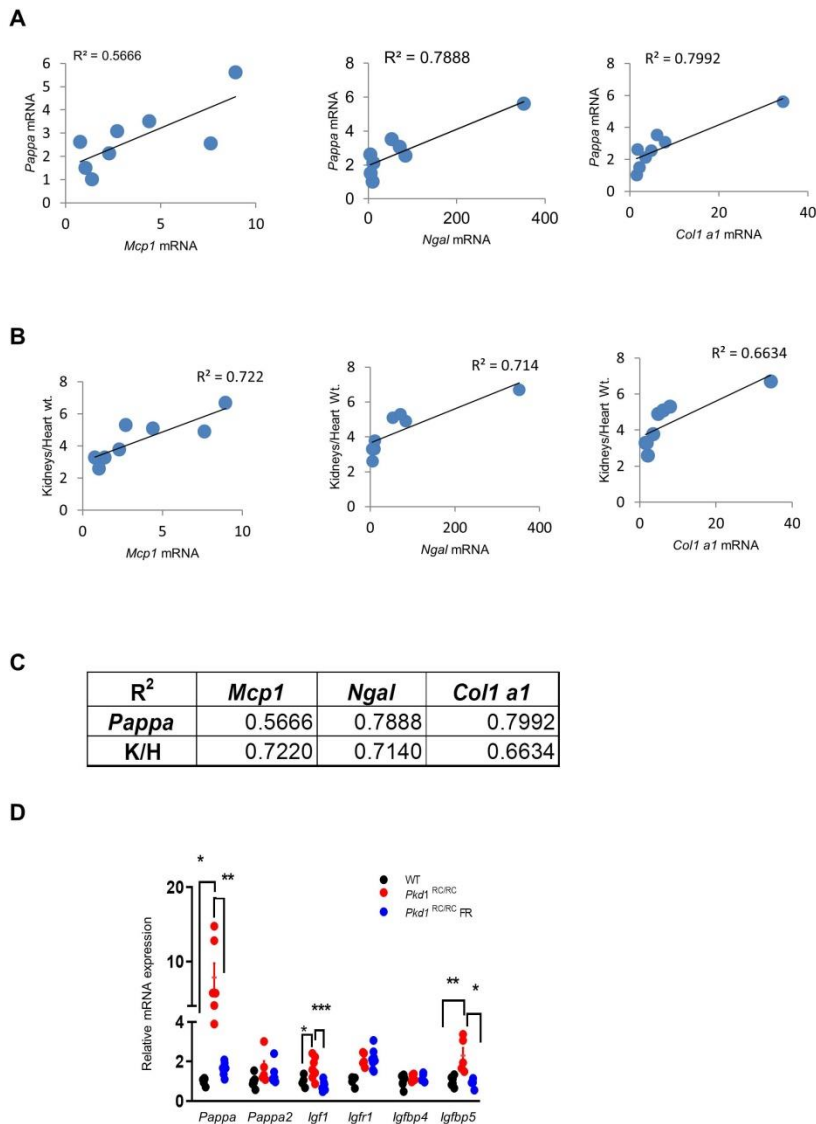
74 *Western blotting*

75 Western blot analysis on kidney tissues and cultured cells was performed as described earlier
76 (3). Antibodies against LC3 (4108), p62 (#5114) cleaved caspase 3 (#9664), pERK(#4370),
77 ERK(#4695), pAkt (#4060), Akt (#4691), pS6 (#4858) and S6 (#2217) were used and
78 purchased from Cell signaling. Membranes were stripped and probed with tubulin or GAPDH
79 antibody to control for equal gel loading and transfer. Films were scanned and densitometry
80 was performed using ImageJ.

81 *Statistical analysis*

82 Data are expressed as means \pm SEM. Comparisons were made by unpaired Student's *t-test*
83 and analysis of variance (ANOVA). Nonparametric tests were used wherever required by
84 data distribution. Significance was set at $P < 0.05$.

85



88 **Fig. S1. Food restriction ameliorated the IGF-1 pathway components in ADPKD. (A-B)**

89 Graphs showing correlation from *Pkd1*^{RC/RC} mice (n=8, 3-7.5 months old) between (A)

90 *Pappa* and *Mcp1*, *Ngal* and *Colla1* mRNA expression levels, (B) kidneys to heart ratio and

91 *Mcp1*, *Ngal* and *Colla1* mRNA expression levels. (C) Table showing the R^2 values for

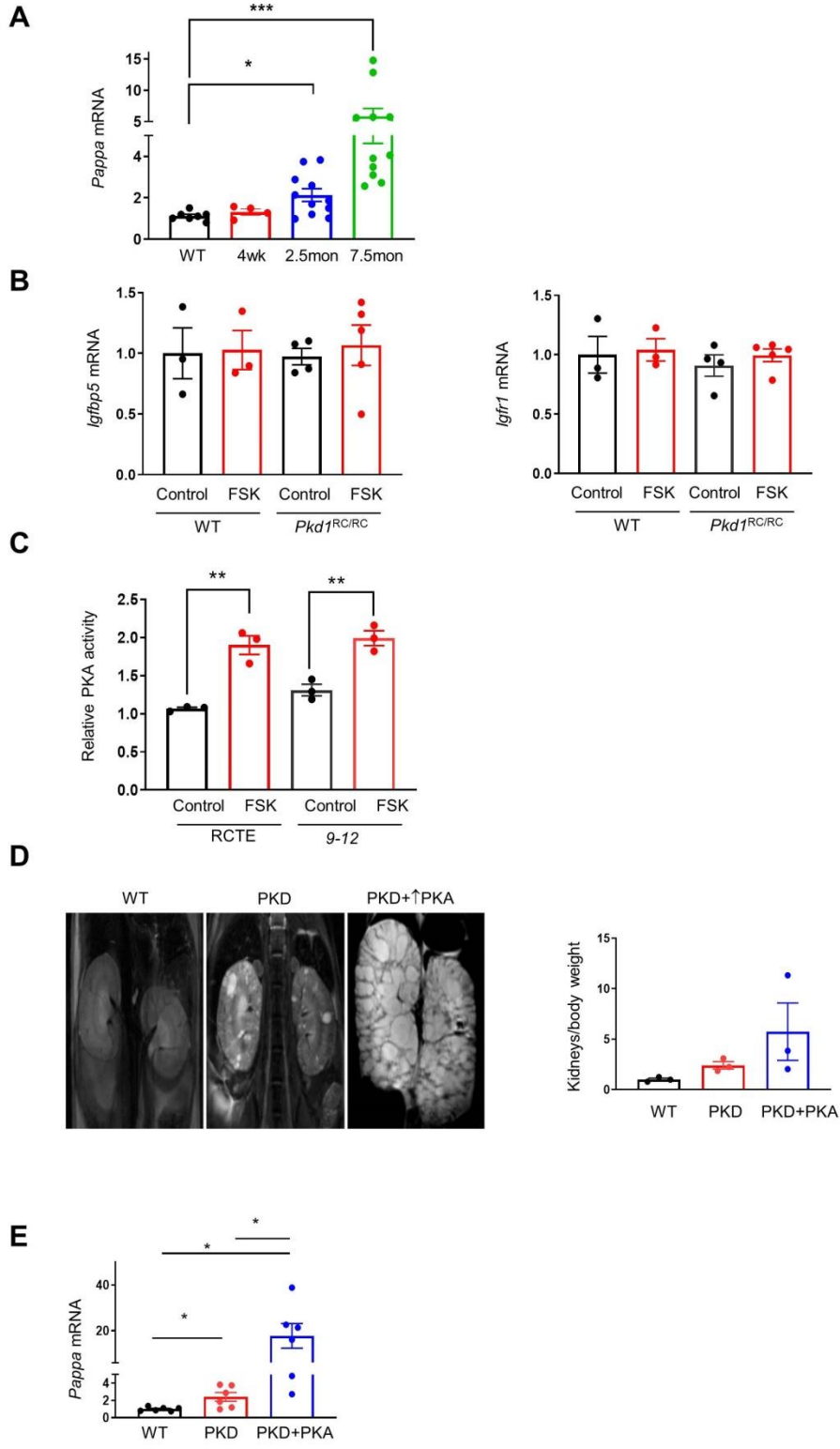
92 figures (A) and (B). (D) Relative mRNA expression of IGF-1 pathway components in

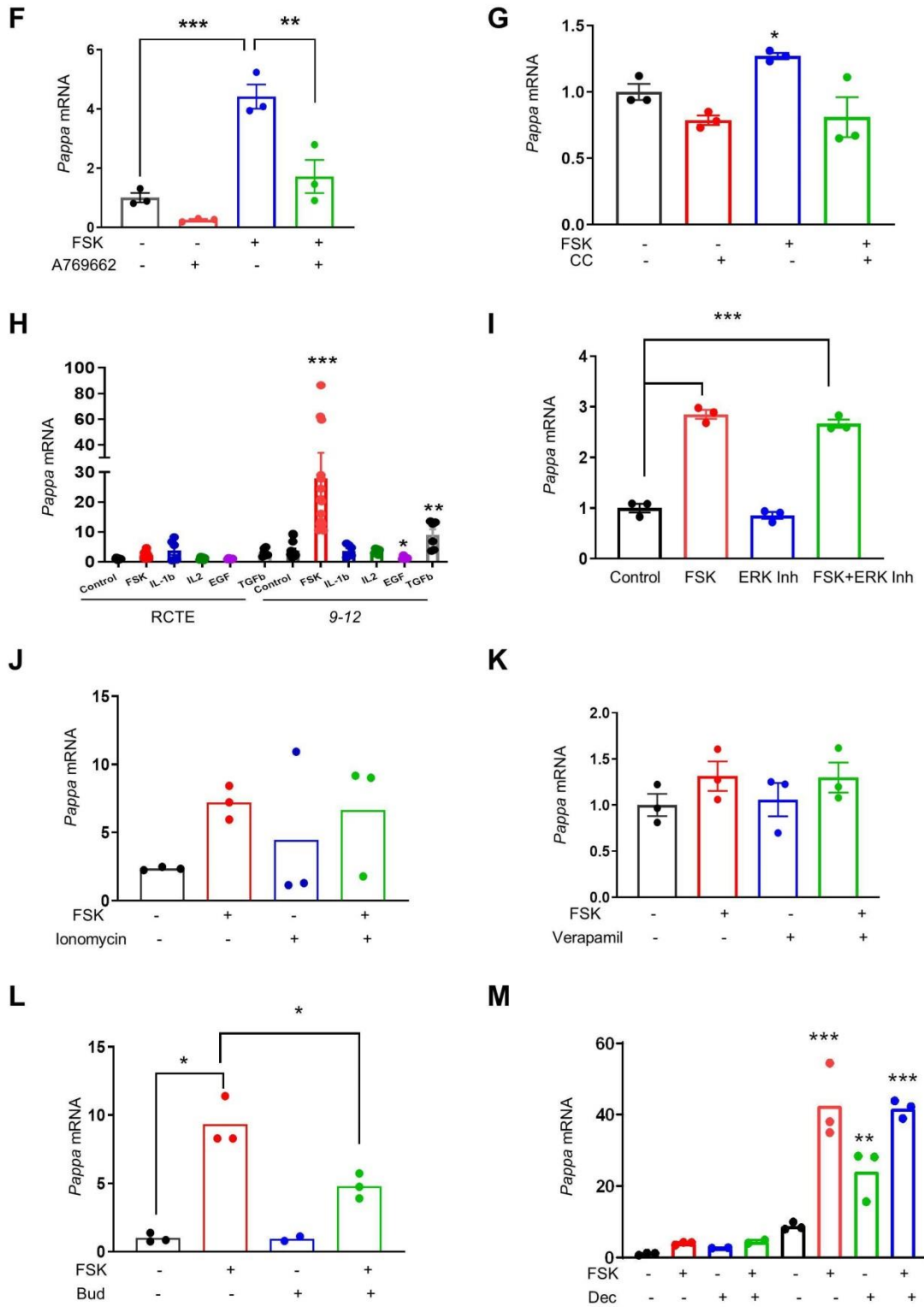
93 kidneys of 7.5 month old WT and *Pkd1*^{RC/RC} mice fed a standard diet ad libitum (AL) or 40%

94 food restriction (FR) for 6 months. PCR data are expressed relative to *Gapdh*, (n=4-8). A *p*

95 value of < 0.05 was considered significant. $*P < 0.05$, $**P < 0.01$, $***P < 0.001$ by two-tailed
96 Student's t-test (One-tailed for WT vs $Pkd1^{RC/RC}$ AL).

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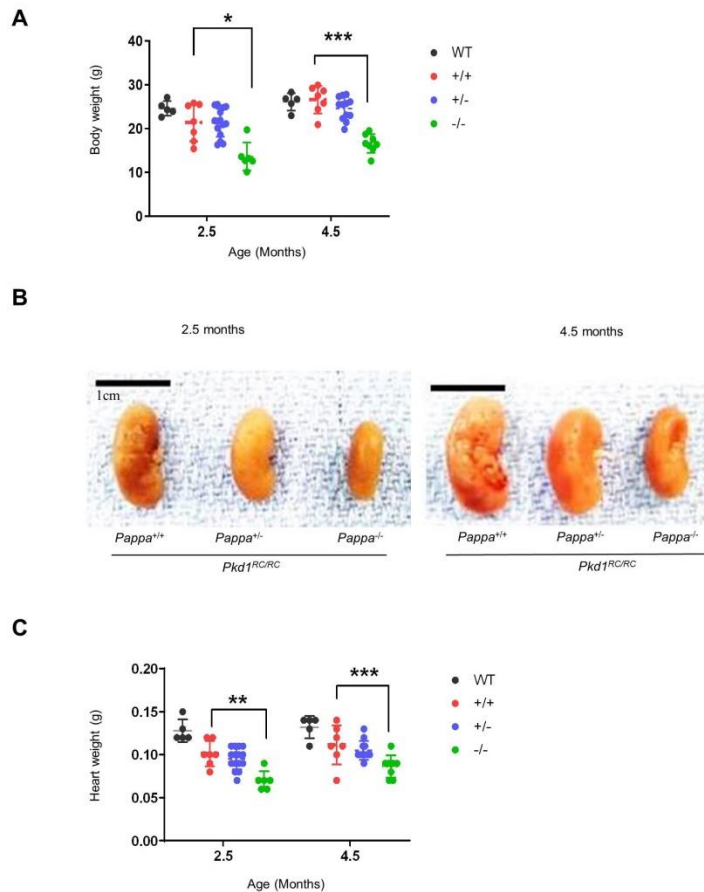
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101 **Fig. S2. cAMP pathway induces PAPP-A expression in ADPKD.** (A) Time course of
 102 *Pappa* expression in *Pkd1^{RC/RC}* mice compared to WT at different time points (n=4-11). (B)
 103 mRNA expression levels of *Igfbp5* and *Igf-1R* in kidney tissues of *Pkd1^{RC/RC}* (n=4-5) and

104 WT (n=3) mice treated with vehicle (5% DMSO) or 5 mg/kg FSK for 24 hours. **(C)** Relative
105 PKA activity in RCTE and 9-12 cells following FSK treatment for 24hrs compared to control.
106 **(D)** Overactivation of PKA exacerbates cystic disease in ADPKD mice. *Pkd1*^{RC/RC} mice were
107 crossed with *Prkar1a*^{fl/fl}; *Pkhd1*-Cre mice to generate ADPKD mice with kidney-specific
108 overactivation of PKA (PKD + ↑PKA). Representative MR Images of WT and PKD kidneys
109 at 3 months old compared to PKD + ↑PKA at 5 weeks old and graph of Kidneys/body
110 weight. **(E)** *Pappa* mRNA expression in kidneys of 3 month old WT and *Pkd1*^{RC/RC} and 5
111 weeks old PKD + ↑PKA mice. **(F)** *Pappa* mRNA expression levels in 9-12 cells treated
112 with/without an AMPK activator, A769662 for 6 hours followed by 10 μM FSK treatment
113 (with/without) for 16 hours. **(G)** mRNA expression levels of *Pappa* in RCTE cells treated
114 with 10 μM FSK in presence or absence of an AMPK inhibitor, compound C for 16 hours.
115 **(H)** mRNA expression levels of *Pappa* in RCTE and 9-12 cells treated with FSK (10 μM),
116 IL1β (5 ng/ml), IL-2 (2 ng/ml), EGF (10 ng/ml) or TGFβ (1 ng/ml) for 16 hours. **(I)** 9-12
117 cells treated with 10 μM FSK in presence or absence of an ERK inhibitor, SCH772984
118 (2μM) for 16 hours. **(J)** 9-12 cells treated with 10 μM FSK in presence or absence of 0.5 μM
119 ionomycin and **(K)** RCTE cells treated with 10 μM FSK in presence or absence of 0.2 μM
120 verapamil for 16 hours. **(L)** 9-12 cells were treated with 10 μM FSK in presence or absence
121 of budesonide (150 μM) or **(M)** RCTE and 9-12 cells treated with 10 μM FSK in presence or
122 absence of decitabine (0.5 μM) for 16 hours. Cells were pre-treated with budesonide or
123 decitabine for 24 hours before FSK treatment. Data are mean ± SEM. A *p* value of < 0.05
124 was considered significant. **P*< 0.05, ***P*< 0.01, ****P*< 0.001 by two-tailed Student's t-test
125 or Mann Whitney test or one way ANOVA and then Tukey's post-hoc test.

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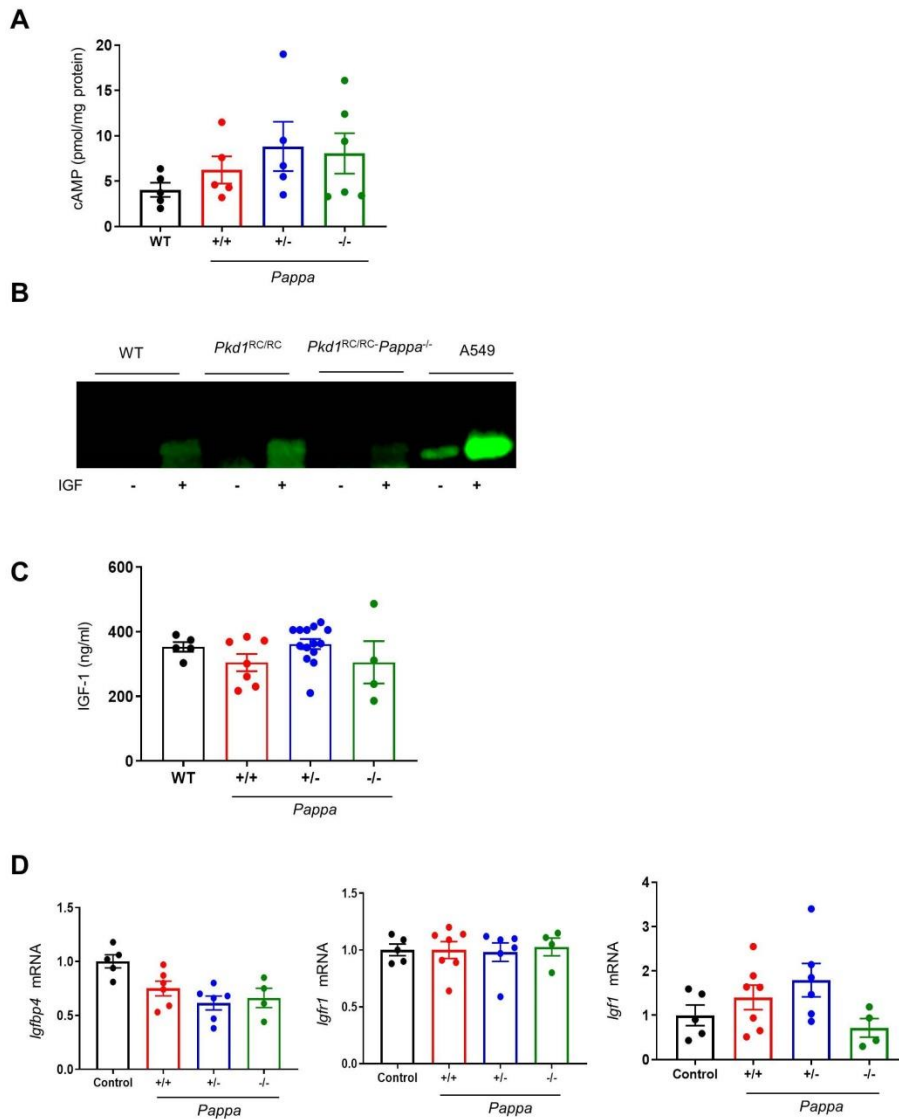
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129 **Fig. S3. PAPP-A deficient mice show reduced body size, heart weight and normal**
 130 **kidneys at different ages. (A)** Body weight of WT and *Pkd1*^{RC/RC}- *Pappa* mutants at
 131 different time points. **(B)** Representative gross kidney images from 2.5 and 4.5 month old
 132 *Pkd1*^{RC/RC} mice that are *Pappa*^{+/+}, *Pappa*^{+/-}, or *Pappa*^{-/-}. **(C)** Heart weight of WT and
 133 *Pkd1*^{RC/RC}- *Pappa* mutants at different ages (n=5-15). Data are mean ± SEM. A *p* value of <
 134 0.05 was considered significant. **P* < 0.05, ***P* < 0.01, ****P* < 0.001 by One-way ANOVA
 135 followed by Tukey's post-hoc test.

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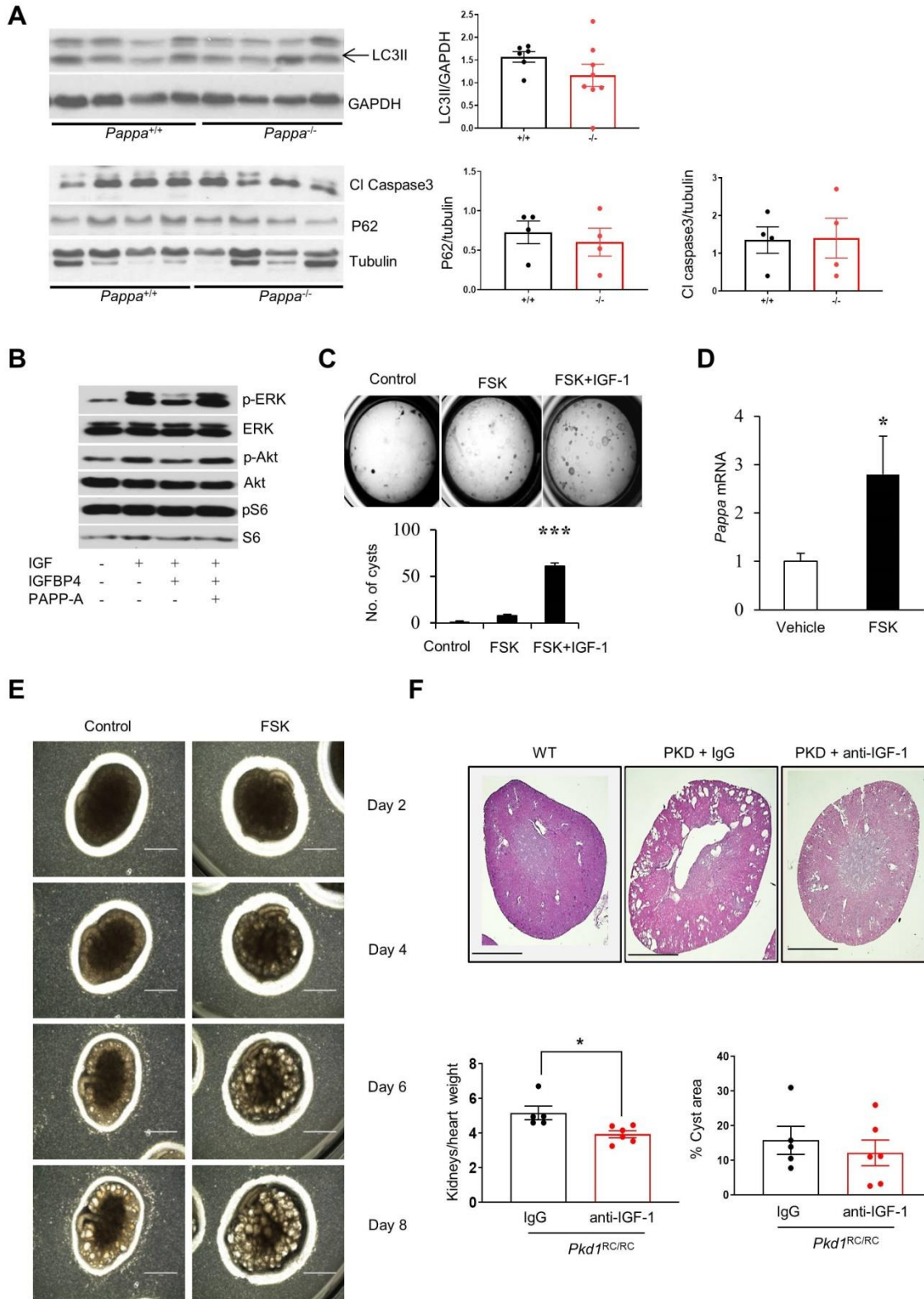
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139 **Fig. S4. cAMP and IGF pathway components in $Pkd1^{RC/RC}$ -*Pappa* mutant mice. (A)**
 140 cAMP levels in kidney tissues of WT and $Pkd1^{RC/RC}$ -*Pappa* mutants (n=5-6). **(B)** Proteolytic
 141 assay of PAPPAs mediated IGFBP4 using kidney membrane fractions from WT, $Pkd1^{RC/RC}$
 142 and $Pkd1^{RC/RC}$ -*Pappa*^{-/-} mice at 6 months age. Membrane fractions were incubated for 72 h at
 143 37 °C with IGFBP-4 without (-) or with (+) pre-complexing to IGF. N-terminal cleaved band
 144 is shown strongly in $Pkd1^{RC/RC}$ compared to WT mice when IGF is pre-incubated but nearly
 145 absent in $Pkd1^{RC/RC}$ -*Pappa*^{-/-} mice. A549 cell line is used as positive control. **(C)** Plasma

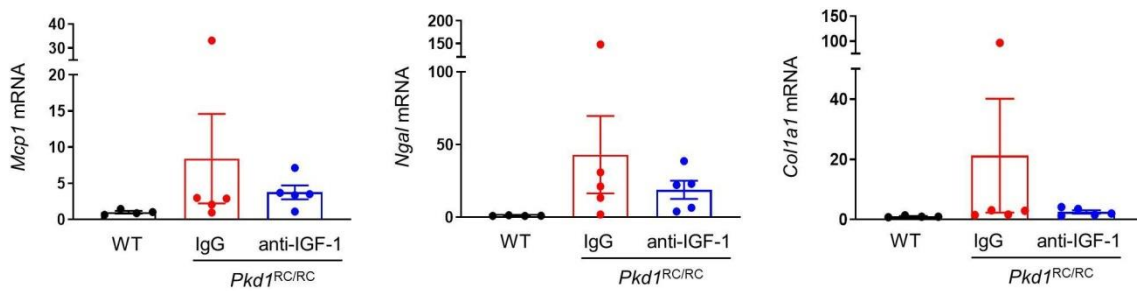
146 IGF-1 levels and **(D)** renal mRNA expression levels of *Igfbp4*, *Igfr1* and *Igf1* in WT and
147 *Pkd1*^{RC/RC}-*Pappa* mutants mice. Data are mean \pm SEM.

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 152 **Fig. S5. Role of PAPP-A-IGF-1 pathway in pathogenesis of ADPKD in experimental**
 153 **models. (A)** Western blot analysis of LC3, cleaved caspase 3 (Cl Caspase3) and p62 in
 154 kidneys of 2.5 month old *Pkd1^{RC/RC}*;*-Pappa^{+/+}* and *Pkd1^{RC/RC}*-*Pappa^{-/-}* mice. Graphs show
 155 quantitative analysis of the specific bands by densitometry. **(B)** Western blot analysis of IGF-
 156 1 and MAP kinase pathways in 9-12, human PKD cells in presence of IGF-1, IGFBP4 or
 157 IGFBP+PAPP-A. **(C)** three dimensional MDCK cystogenic assay in the presence of FSK
 158 alone or FSK + IGF1 (top) and quantification of MDCK cysts (bottom) IGF-1. **(D)** *Pappa*
 159 mRNA expression in metanephros treated with FSK or vehicle. **(E)** Photomicrographs
 160 showing the cystic growth in a metanephric model of cystogenesis in *Pkd1^{RC/RC}* mice at Day
 161 13.5 embryonic kidneys that were stimulated with FSK (10 μ M) or control in absence of any
 162 growth hormone. Scale bar, 1 mm. **(F)** *Pkd1^{RC/RC}* mice were treated with IGF-1 neutralizing
 163 antibody (0.2 mg/kg, n=6) or control IgG (n=5) weekly by I.P. injection for 6 weeks starting
 164 at 4 months old age. Representative H&E sections of kidneys and graphs of kidneys/body
 165 weight and cystic area in IgG (n=5) and anti-IGF-1 (n=6) treated *Pkd1^{RC/RC}* mice. **(G)** mRNA
 166 levels of *Mcp1*, *Ng2* and *Col1a1* in IgG and anti-IGF1 (n=5 for each group) treated
 167 *Pkd1^{RC/RC}* mice compared to WT mice (n=4). Data are mean \pm SEM. A *p* value of < 0.05 was
 168 considered significant. **P* < 0.05, ****P* < 0.001 compared to control by two-tailed Student's
 169 t-test.

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