

Table S1. The blood cell count in 3-7 years old DMD patients (n=25) before glucocorticoid therapy indicating no severe abnormalities in the level of white blood cells.

<b>Parameter</b>	<b>Average <math>\pm</math> SEM</b>	<b>Norm</b>
<i>White blood cell count (WBC), x 10<sup>3</sup>/<math>\mu</math>l</i>	8.3 $\pm$ 0.3	(4.0 – 11.0)
<i>Neutrophils, x 10<sup>3</sup>/<math>\mu</math>l</i>	3.7 $\pm$ 0.3	(1.9 – 8.0)
<i>Lymphocytes, x 10<sup>3</sup>/<math>\mu</math>l</i>	3.6 $\pm$ 0.2	(0.9 – 5.2)
<i>Monocytes, x 10<sup>3</sup>/<math>\mu</math>l</i>	0.54 $\pm$ 0.03	(0.16 – 1.00)
<i>Eosinophils, x 10<sup>3</sup>/<math>\mu</math>l</i>	0.3 $\pm$ 0.1	(0.0 – 0.8)
<i>Basophils, x 10<sup>3</sup>/<math>\mu</math>l</i>	0.033 $\pm$ 0.005	(0.000 – 0.200)
<i>Neutrophils, % WBC</i>	↓ 44.2 $\pm$ 2.3	(50.0 – 66.0)
<i>Lymphocytes, % WBC</i>	↑ 44.9 $\pm$ 2.1	(20.0 – 40.0)
<i>Monocytes, % WBC</i>	6.6 $\pm$ 0.4	(4.0 – 8.0)
<i>Eosinophils, % WBC</i>	3.7 $\pm$ 0.6	(2.0 – 4.0)
<i>Basophils, % WBC</i>	0.5 $\pm$ 0.1	(0.0 – 1.0)

Table S2. RNA sequencing analysis results showing genes downregulated (green) and upregulated (red) in miR-378<sup>-/-</sup> mice in comparison to the WT mice that are changed at least 1.5 times with  $p_{adj} < 0.1$ . Segregation was done based on the fold change values.

Gene symbol	Gene full name	Fold change	P <sub>adj</sub>
<b>Immune system</b>			
<i>Btla</i>	B- and T-lymphocyte attenuator	7.41	0.0078
<i>Iigp1</i>	Interferon-inducible GTPase 1	3.66	0.00020
<i>Ly96</i>	lymphocyte antigen 96	0.22	0.090
<b>Fibrosis and ECM</b>			
<i>Col6a6</i>	collagen type VI alpha 6 chain	2.88	0.0054
<i>Fgf1</i>	fibroblast growth factor 1	0.19	5.09E-15
<b>Metabolism</b>			
<i>Blvrb</i>	biliverdin reductase	2.86	0.028
<i>Rpsud4</i>	RNA pseudouridylation synthase domain containing 4	0.65	0.090
<i>St6galnac4</i>	ST6 N-acetylgalactosaminide alpha-2,6-sialyltransferase 4	0.43	0.062
<i>Gnpdal</i>	glucosamine-6-phosphate deaminase 1	0.25	0.0017
<i>Aldh1b1</i>	aldehyde dehydrogenase 1 family, member B1	0.11	0.098
<i>B3gat1</i>	beta-1,3-glucuronyltransferase 1	0.06	0.083
<b>Genetic Information Processing and Regulation</b>			
<i>Dbp</i>	D site albumin promoter binding protein	3.48	0.012
<i>Egr1</i>	early growth response 1	3.45	0.073
<i>Rpl34</i>	ribosomal protein L34	3.26	0.0069
<i>Klhl38</i>	kelch-like 38	3.20	0.0022
<i>Rplp0</i>	ribosomal protein, large, P0	1.86	0.090
<i>Rnps1</i>	ribonucleic acid binding protein S1	1.81	0.081
<i>Urm1</i>	ubiquitin related modifier 1	0.66	0.069
<i>Noct</i>	nocturnin	0.44	0.091
<i>Snora34</i>	small nucleolar RNA, H/ACA box 34	0.30	0.027
<i>G0s2</i>	G0/G1 switch gene 2	0.30	0.041
<i>Rpl26</i>	ribosomal protein L26	0.08	2.5E-05
<b>Signal transduction</b>			
<i>Map3k8</i>	mitogen-activated protein kinase 8	2.86	0.0039
<i>Pik3ip1</i>	phosphoinositide-3-kinase interacting protein 1	1.97	0.022
<i>Fbxo32</i>	F-box protein 32	1.79	0.028
<b>Muscles</b>			
<i>Tnnt1</i>	troponin T1, skeletal, slow	2.94	0.027
<i>Myh7b</i>	myosin, heavy chain 7B, cardiac muscle, beta	2.83	0.078
<i>Atp2a2</i>	ATPase, Ca <sup>++</sup> transporting, cardiac muscle, slow twitch 2	2.34	0.097
<i>Myod1</i>	myogenic differentiation 1	0.56	0.027
<i>Myhas</i>	myosin heavy chain gene antisense RNA	0.29	0.039
<b>Nervous system</b>			
<i>Ngef</i>	neuronal guanine nucleotide exchange factor	13.70	0.091
<i>Qrfp</i>	pyroglutamylated RFamide peptide	5.18	0.0017
<i>Syt12</i>	synaptotagmin XII	0.35	0.074
<i>Pak3</i>	p21 protein (Cdc42/Rac)-activated kinase 3	0.03	0.052
<b>Heat shock protein</b>			
<i>Hsph1</i>	heat shock 105kDa/110kDa protein 1	0.58	0.027
<i>Hsp90aa1</i>	heat shock protein 90, alpha (cytosolic), class A member 1	0.29	0.042
<i>Hspa1b</i>	heat shock protein 1B	0.21	0.064
<i>Hspa1a</i>	heat shock protein 1A	0.17	0.0048
<b>Other</b>			
<i>Dlk1</i>	delta like non-canonical Notch ligand 1	5.82	0.0046
<i>Adams20</i>	a disintegrin-like and metallopeptidase (reprolysin type) with thrombospondin type 1 motif, 20	3.05	0.0017
<i>Gadd45b</i>	growth arrest and DNA-damage-inducible 45 beta	3.02	0.0017
<i>Nr1d1</i>	nuclear receptor subfamily 1, group D, member 1	2.27	0.00052
<i>Slc47a1</i>	solute carrier family 47, member 1	0.60	0.0093
<i>Mrm1</i>	mitochondrial rRNA methyltransferase 1	0.54	0.097
<i>Rsph1</i>	radial spoke head 1 homolog	0.41	0.014
<i>Abra</i>	actin-binding Rho activating protein	0.36	0.0015
<i>Slc37a1</i>	solute carrier family 37, member 1	0.11	0.0017
<i>Mir133b</i>	microRNA 133b	0.04	0.033

Table S3. RNA sequencing analysis results showing genes downregulated (green) and upregulated (red) in dKO mice in comparison to the mdx mice that are changed at least 1.5 times with  $p_{adj} < 0.1$ . Segregation was done based on the fold change values.

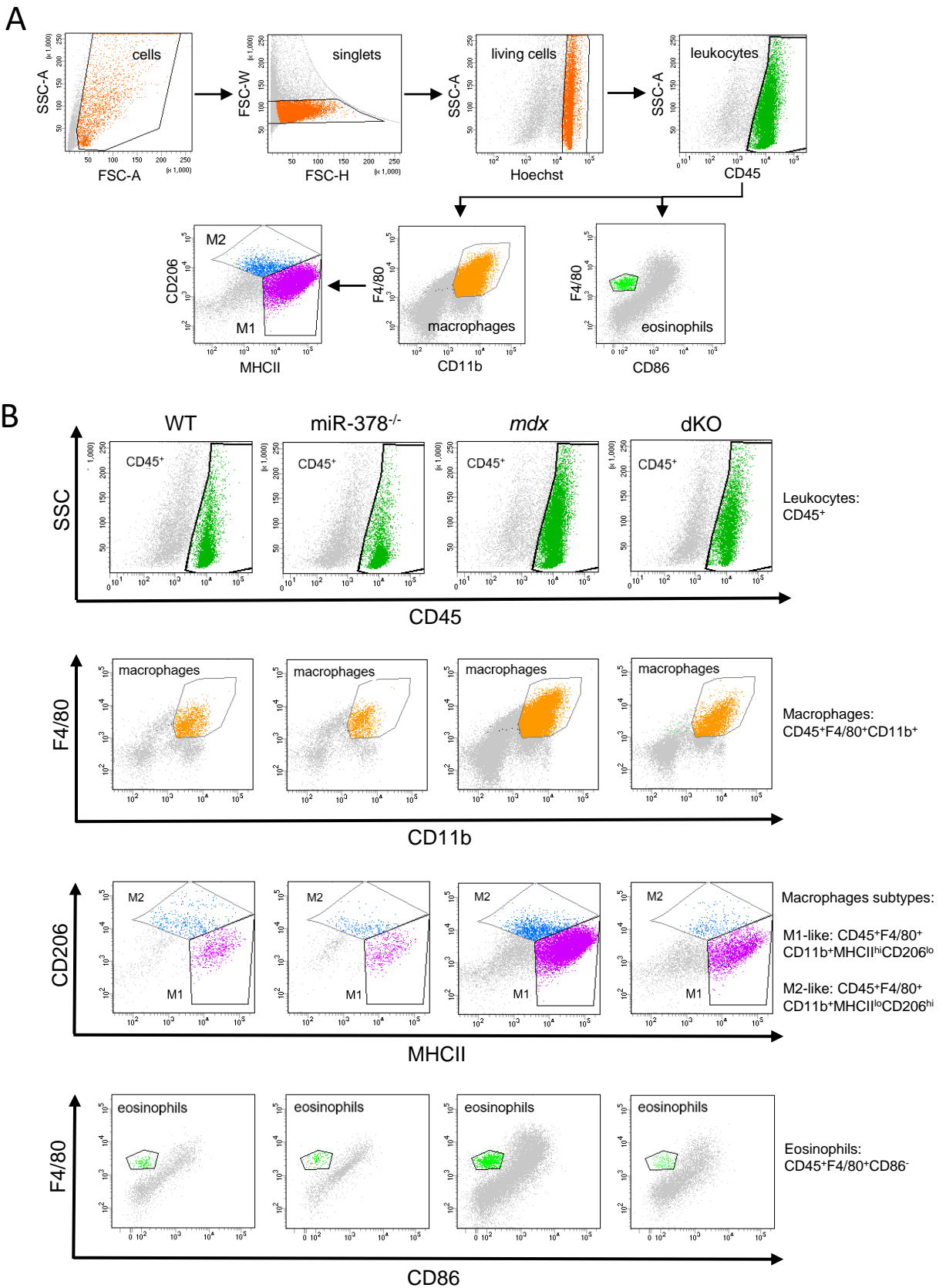
Gene symbol	Gene full name	Fold change	$p_{adj}$
<b>Immune system</b>			
<i>H2-Q10</i>	histocompatibility 2, Q region locus 10	9.93	5.7E-15
<i>Iigp1</i>	Interferon-inducible GTPase 1	4.46	7.8E-07
<i>H2-K1</i>	histocompatibility 2, K region locus 2	0.50	0.058
<i>C1qc</i>	complement component 1, q subcomponent, C chain	0.46	0.069
<i>Mgl2</i>	macrophage galactose N-acetyl-galactosamine specific lectin 2	0.44	1.95E-05
<i>C1qb</i>	complement C1q subcomponent subunit B	0.36	0.0065
<i>H2-D1</i>	histocompatibility 2, D region locus 1	0.14	5.56E-44
<i>H2-T-ps</i>	histocompatibility 2, T region locus, pseudogene	0.09	2.79E-06
<i>H2-Q5</i>	histocompatibility 2, Q region locus 5	0.06	0.0090
<i>H2-Q7</i>	histocompatibility 2, Q region locus 7	0.05	1.36E-10
<i>H2-Ea-ps</i>	histocompatibility 2, class II antigen E alpha, pseudogene	0.01	4.52E-05
<b>Fibrosis and ECM</b>			
<i>Nbl1</i>	neuroblastoma suppressor of tumorigenicity 1	0.57	0.042
<i>Spon2</i>	spondin 2	0.44	0.0052
<i>Fgf1</i>	fibroblast growth factor 1	0.20	3.17E-15
<b>Metabolism</b>			
<i>Eno1b</i>	enolase 1B	48.40	2.79E-06
<i>Tnxa</i>	tenascin XA	14.40	0.018
<i>Aldh1a1</i>	aldehyde dehydrogenase family 1, subfamily A1	1.96	0.098
<i>St8sia5</i>	ST8 alpha-N-acetyl-neuraminide alpha-2,8-sialyltransferase 5	1.60	0.041
<i>Galnt16</i>	polypeptide N-acetylgalactosaminyltransferase 16	0.62	0.030
<i>Cox18</i>	cytochrome c oxidase assembly protein 18	0.47	0.052
<i>Gnpdal</i>	glucosamine-6-phosphate deaminase 1	0.38	0.0092
<i>Hddc3</i>	HD domain containing 3	0.34	0.028
<b>Genetic Information Processing and Regulation</b>			
<i>Nr4a3</i>	nuclear receptor subfamily 4, group A, member 3	5.02	0.074
<i>Igfn1</i>	immunoglobulin-like and fibronectin type III domain containing 1	2.37	0.098
<i>Rps18</i>	ribosomal protein S18	0.31	0.029
<i>Rpl26</i>	ribosomal protein L26	0.08	1.53E-05
<i>Rplp0-ps1</i>	ribosomal protein, large, P0, pseudogene 1	0.02	0.0077
<b>Signal transduction</b>			
<i>Sfrp2</i>	secreted frizzled-related protein 2	0.31	4.52E-05
<b>Other</b>			
<i>Fam20c</i>	family with sequence similarity 20, member C	0.58	0.030
<i>Pex6</i>	peroxisomal biogenesis factor 6	0.57	0.097
<i>Tubal1c</i>	tubulin, alpha 1C	0.39	0.00071
<i>Cdkn2a</i>	cyclin-dependent kinase inhibitor 2A	0.32	0.056

Table S4. The sequences of forward (F) and reverse (R) primers used for the determination of gene expression on mRNA level by qRT-PCR.

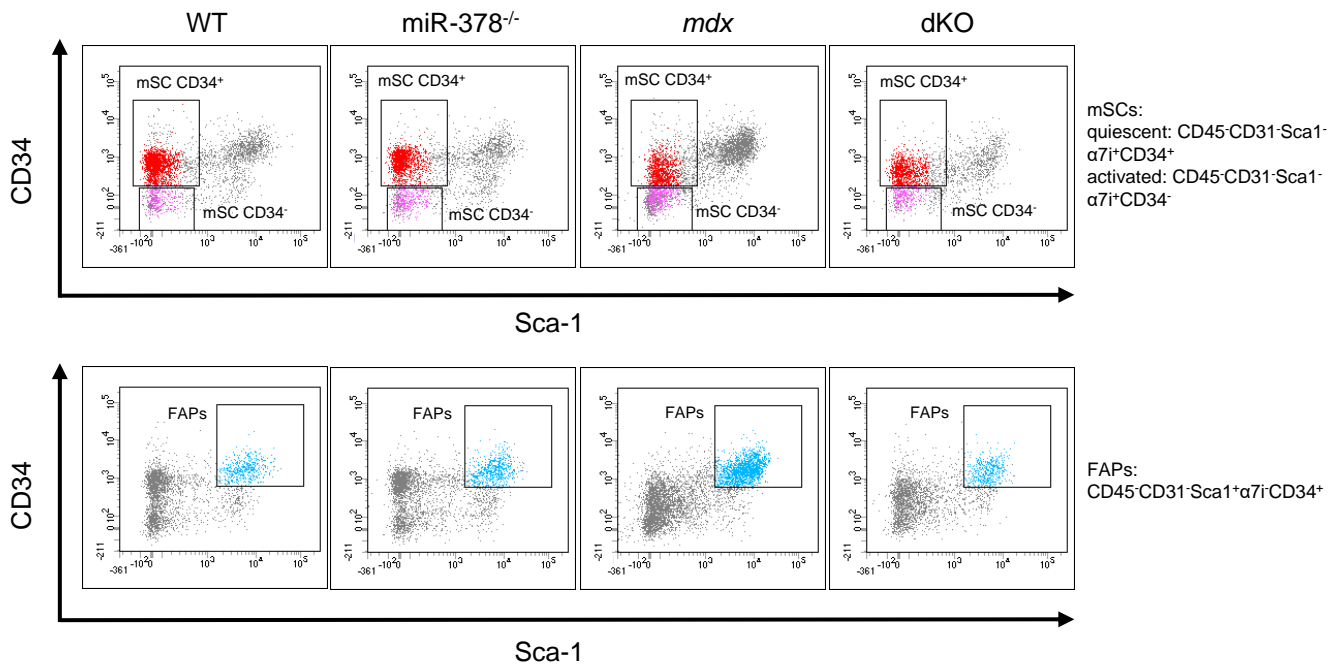
<b>Gene</b>	<b>Sequence 5' → 3'</b>
<b>Ampka1</b>	F: TGTGACAAGCACATTTTCCAA
	R: CCGATCTCTGTGGAGTAGCA
<b>Ampka2</b>	F: ACAGCGCCATGCATATTCCT
	R: TCCGACTGTCTACCAGGTAA
<b>Colla1</b>	F: CGATCCAGTACTCTCCGCTCTTCC
	R: ACTACCGGGCCGATGATGCTAACG
<b>Eef2</b>	F: AGAACATATTATTGCTGGCG
	R: CAACAGGGTCAGATTTCTTG
<b>Essra</b>	F: AGAGCCAGCCAGTCCTGACA
	R: TCACAGGATGCCACACCGTA
<b>Essrg</b>	F: GGAAGAATTCGTCACCCTCA
	R: TTCTGCACAGCTTCCACATC
<b>Fgf1</b>	F: ATGGACACCGAAGGGCTTTT
	R: GAGGCCACAAAACCAGTTCT
<b>Fn1</b>	F: AGCCTGCTCATCAGTTGGGA
	R: GATGGAAACTGGCTTGCTGC
<b>Pecam1</b>	F: CCAAAGCCAGTAGCATCATGGTC
	R: GGATGGTGAAGTTGGCTACAGG
<b>Ppara</b>	F: ACTACGGAGTTCACGCATGTG
	R: TTGTCGTACACCAGCTTCAGC
<b>Ppard</b>	F: GCCCAAGTTCGAGTTTGCTGT
	R: ATTCTAGAGCCCGCAGAATG
<b>Pparg1a</b>	F: TTGACTGGCGTCATTCGGGA
	R: GCAGGCTCATTGTTGTA CTG
<b>Pparg1b</b>	F: CTGACACGCAGGGTGGG
	R: AGGTCAAGCTCTGGCAAGTC

Table S5. The sequences of primers used for the determination of miRNA level using miRCURY LNA RT-PCR Kit.

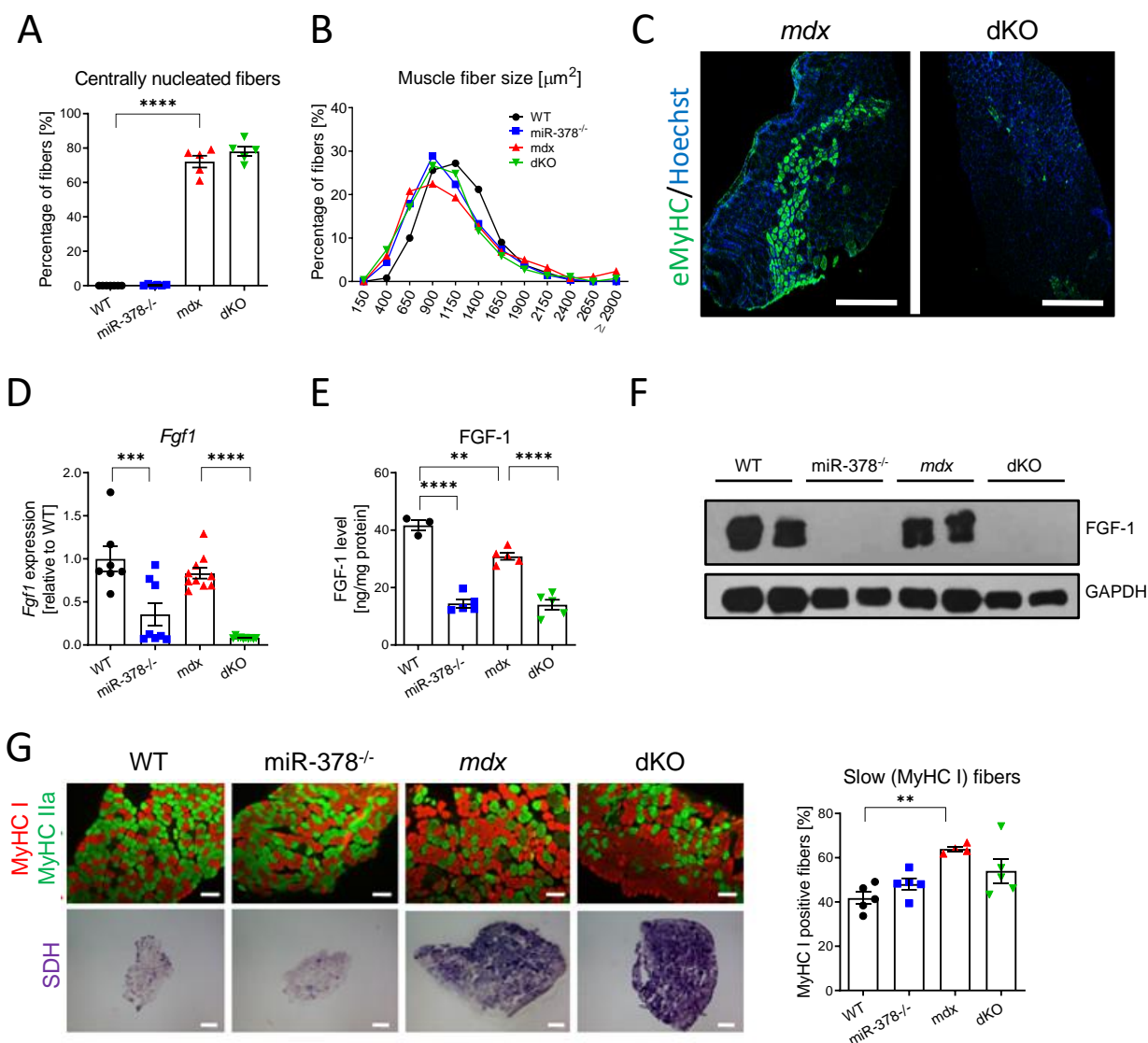
<b>miRNA</b>	<b>Sequence</b>
<i>miR-378-3p</i>	5'-ACUGGACUUGGAGUCAGAAGG-3'
<i>miR-378-5p</i>	5'-CUCCUGACUCCAGGUCCUGUGU-3'
<i>miR-16-3p</i>	5'-CCAGUAUUGACUGUGCUGCUGA-3'
<i>miR-16-5p</i>	5'-UAGCAGCACGUAAAUAUUGGCG-3'
<i>U6 snRNA</i>	5'-CGCAAGGATGACACGCAAATTC-3'



**Supplementary Figure 1. The analysis of immune cells populations within skeletal muscles of hind limbs of WT, *miR-378<sup>-/-</sup>*, *mdx* and *dKO* mice based on flow cytometry analysis. (A) Gating strategy. (B) Examples of two-parameter flow cytometry dot plots for discrimination of leukocytes (CD45<sup>+</sup> cells), macrophages (CD45<sup>+</sup>F4/80<sup>+</sup>CD11b<sup>+</sup> cells), M1-like macrophages (CD45<sup>+</sup>F4/80<sup>+</sup>CD11b<sup>+</sup>MHCII<sup>hi</sup>CD206<sup>lo</sup>), M2-like macrophages (CD45<sup>+</sup>F4/80<sup>+</sup>CD11b<sup>+</sup>MHCII<sup>lo</sup>CD206<sup>hi</sup>) and eosinophils (CD45<sup>+</sup>F4/80<sup>+</sup>CD86<sup>+</sup>).**

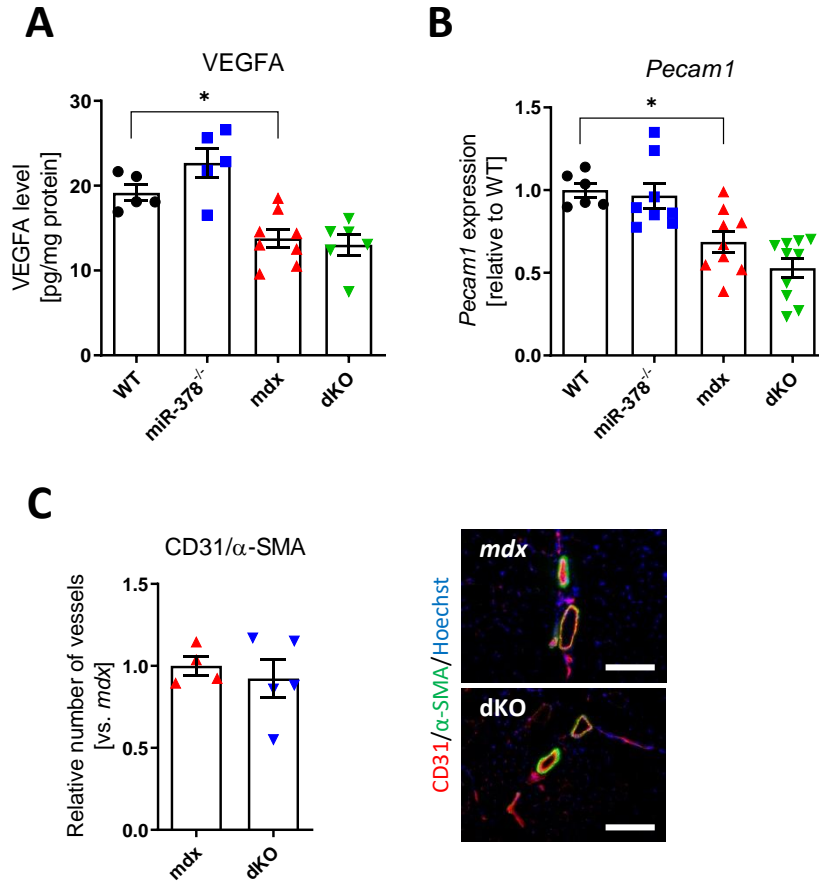


**Supplementary Figure 2.** The analysis of mSCs and FAPs populations within skeletal muscles of hind limbs of WT, miR-378<sup>-/-</sup>, mdx and dKO mice. Examples of two-parameter flow cytometry dot plots for discrimination of quiescent (CD45<sup>-</sup>CD31<sup>-</sup>Sca1<sup>-</sup>α7i<sup>+</sup>CD34<sup>+</sup>) mSCs, activated (CD45<sup>+</sup>CD31<sup>+</sup>Sca1<sup>-</sup>α7i<sup>+</sup>CD34<sup>-</sup>) mSCs and FAPs (CD45<sup>+</sup>CD31<sup>+</sup>Sca1<sup>+</sup>α7i<sup>-</sup>CD34<sup>+</sup>).

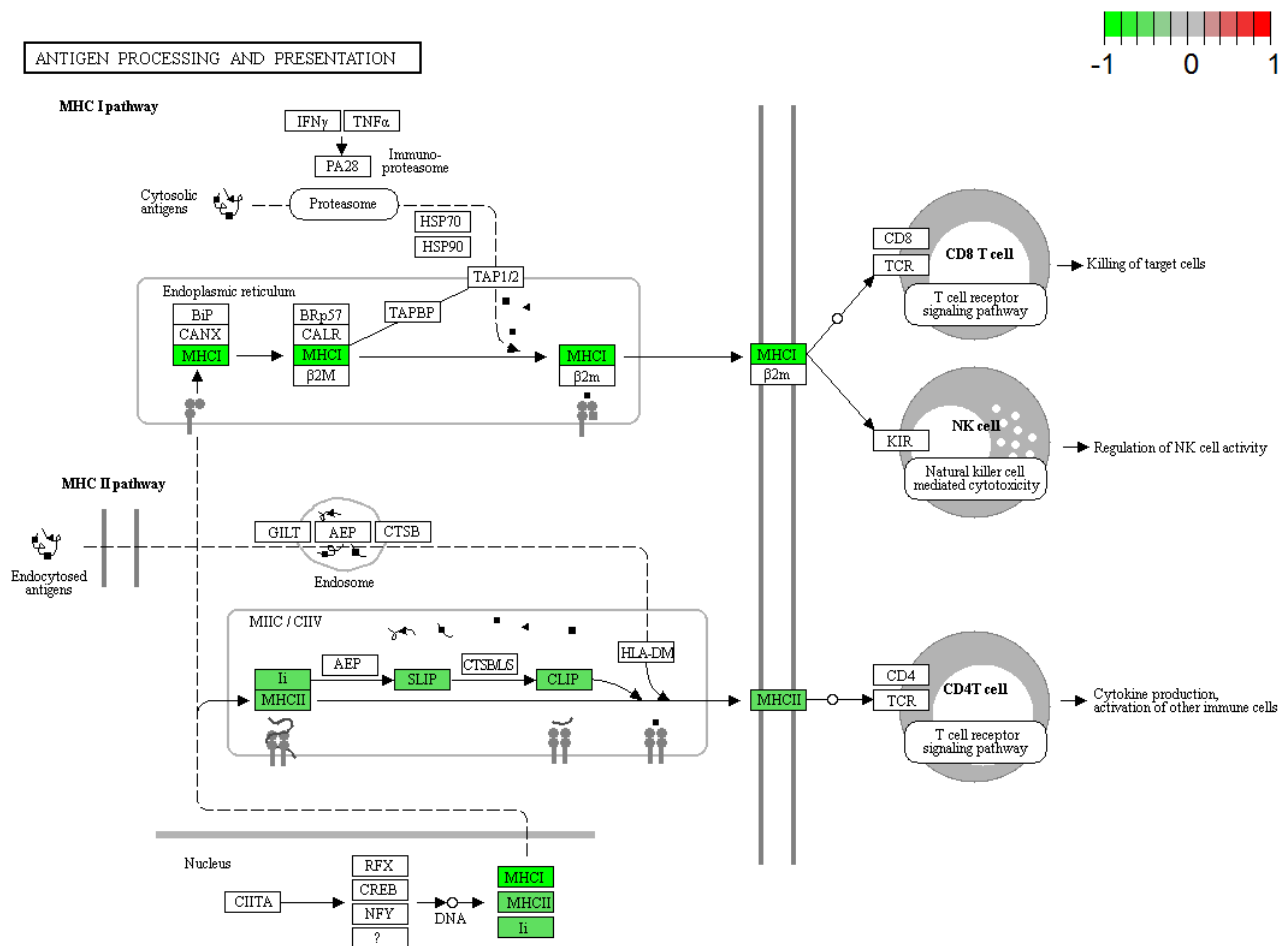


**Supplementary Figure 3. The effect of miR-378 deficiency in the soleus muscle of 3-month-old mice.** (A) The analysis of centrally nucleated fibers performed based on H&E staining; microscopic assessment using Nikon Eclipse microscope; n=5-7/group. (B) The quantification of muscle fibers size based on laminin staining indicating no effect of miR-378 deficiency; n=4-5/group. (C) Immunofluorescent staining of eMyHC expression (green) showing its decreased level in dKO mice; Hoechst (blue) was used to visualize nuclei; representative pictures; confocal microscope (LSM-510; Carl Zeiss); scale bar: 100  $\mu\text{m}$ ; n=4-5/group. The expression of (D) *Fgf1* mRNA and FGF-1 protein level analyzed by (E) ELISA and (F) Western blot indicating a potent downregulatory effect of miR-378 deficiency; qRT-PCR results calculated as the relative expression level compared to WT animals; n=7-10/group, ELISA test calculated as ng of FGF-1 per mg protein; n=3-5/group; representative Western Blot result. (G) Representative pictures of immunofluorescent staining of slow (MyHC I, red), fast (MyHC IIa, green) MyHC isoforms (upper panel) with a histochemical assessment of SDH activity (bottom panel) indicating the increased abundance of oxidative fibers in *mdx* mice; n=4-5/group, microscopic assessment using Nikon Eclipse microscope, scale bar: 100  $\mu\text{m}$ . Data are presented as mean  $\pm$  SEM. \*p<0.05; \*\*p<0.01; \*\*\*p<0.001; \*\*\*\*p<0.0001; one-way ANOVA with Tukey's post-hoc test.





**Supplementary Figure 4. miR-378 loss does not affect angiogenesis markers in the muscles of 3-month-old animals.** A decreased (A) VEGFA protein level in gastrocnemius and (B) expression of *Pecam1* in diaphragm muscles of *mdx* animals without any apparent changes driven by the lack of miR-378; ELISA (n=5-8/group) and qRT-PCR (n=6-10/group) analyses, respectively. (C) No alterations in the blood vessels abundance in gastrocnemius muscle of dKO vs. *mdx* mice; microscopic assessment using Nikon Eclipse microscope, scale bar: 100  $\mu$ m. Data are presented as mean  $\pm$  SEM. \*p<0.05; (A-B) one-way ANOVA with Tukey's post-hoc test; (C) unpaired 2-tailed Student's t-test.



Data on KEGG graph  
Rendered by Pathview

**Supplementary Figure 5.** KEGG pathway analysis based on RNA-sequencing data. Antigen processing and presentation pathway is diminished in dKO vs. *mdx* mice.