Supplemental Table 1: Median and IQR for cell surface markers in perinatal infections.

	Baseline	**Standard (12h PMA/Ionomycin)**	**Enhanced (18h PHA/PMA/Ionomycin)**
CD69	3.62 (1.03 – 4.45)	96.72 (96.62 – 97.95)	98.42 (98.22 – 98.75)
CD25	13.58 (7.05 – 24.11)	70.41 (55.97 – 81.62)	77.88 (58.57 – 84.3)
HLA-DR	4.55 (2.56 – 7.07)	30.86 (21.76 – 37.27)	35.76 (31.27 – 43.97)
PD-1	10.26 (6.17 – 13.31)	20.95 (16.95 – 25.46)	18.35 (15.05 – 26.96)
TIM-3	7.470 (5.08 – 8.85)	13.17 (10.36 – 13.66)	8.96 (5.95 – 9.47)
TIGIT	20.37 (15.6 – 22.7)	17.77 (12.6 – 19.23)	13.67 (11.1 – 17.93)

Supplemental Table 2: Median and IQR for cell surface markers in adult infections.

	Baseline	Standard (12h PMA/Ionomycin)	Enhanced (18h PHA/PMA/Ionomycin)
CD69	5.88 (1.645 – 23.55)	92.95 (64.6 – 98.29)	94.1 (85.05 – 98.37)
CD25	12.59 (4.26 – 16.9)	66.39 (35.95 – 72.28)	68.24 (42.72 – 76.93)
HLA-DR	10.52 (9.068 – 16.99)	35.87 (21.49 – 60.37)	48.52 (22.01 – 75.39)
PD-1	9.43 (2.47 – 15.1)	15.2 (5.4 – 20.83)	14.55 (5.69 – 20.83)
TIM-3	10.61 (7.88 – 14.06)	12.41 (9.31 – 15.01)	8.56 (7.35 – 9.32)
TIGIT	15.96 (11.67 – 20.41)	11.83 (6.34 – 16.04)	9.805 (6.34 – 12.51)

Supplemental Table 3. Primers and Probes used for viral subtypes.

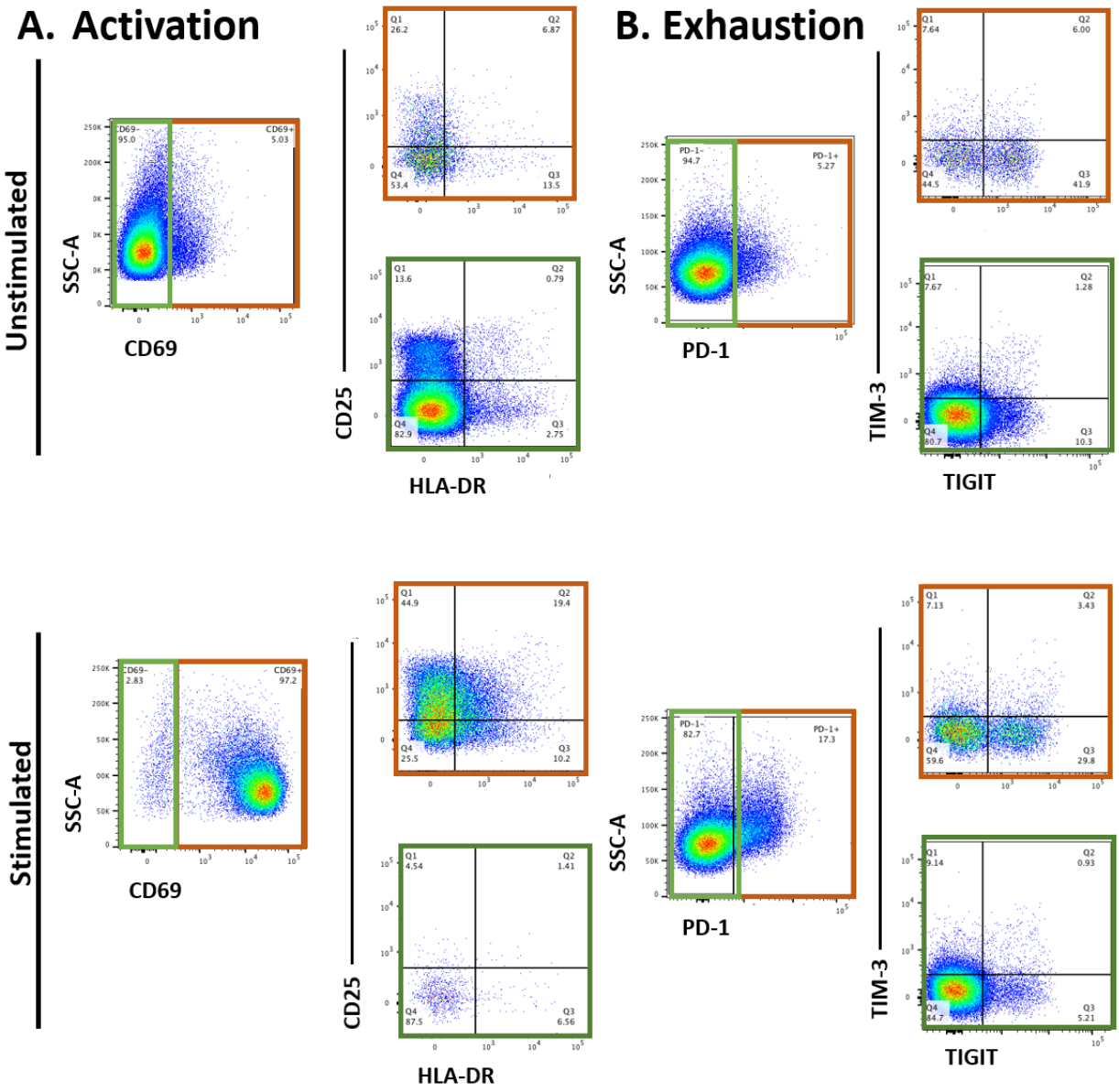
	Tat1.4	**Rev**	**Tat2**	**HIVFamzen**
B[1]	TGG CAG GAA GAA GCG GAG A	GGA TCT GTC TCT GTC TCT CTC TCC ACC	ACA GTC AGA CTC ATC AAG TTT CTC TAT CAA AGC A	/56-FAM/TTC CTT CGG /ZEN/GCC TGT CGG GTC CCGTC CC/3IABkFQ/
A/E[2]	TGG CAG GAA GAA GCG GAA G	TGT CTC TGY CTT GCT CKC CAC C	GCA GTA AGG ATC ATC AAA ATC CTA TAC CAG AGC A	/56-FAM/TTC YTT CGG/ZEN/GCC TGT CGG GTT CC/3IABkFQ
A/D	TGG CAG GAA GAA GCG GAG A	TGG TTC TGY CTT GCT CTC CAC C	GCA GTC AGG ATC ATC AAA ATC CTA TAC CAA AGC A	/56-FAM/TTCYTCCGG/ZEN/GCCTGTC GAGATCC/3IABkFQ/
A/G	TGG CAG GAA GAA GCG GAG A	TGT CTC TGY CTT GCT CKC CAC C	GCC GTC AGG ATC ATC AAA ATC CTG TAC CAA AGC A	/56-FAM/TTC YTT CGG/ZEN/GCC TGT CGG GTT CC/3IABkFQ
A	TGG CAG GAA GAA GCG GAR R	GAT CTG YCT CTG YCT TGC TCT CCA CC	GCA GTA AGG ATC ATC AAA ATC CTR TAC CAA AGC A	/56-FAM/TTCTTCCGG/ZEN/GCCTGTC GGGWYCC/3IABkFQ/
C	TGG CAG GAA GAA GCG GAG A	GAT CTG YCT CTG YCT TGC TCT CCA CC	GCA GTG AGG ATC ATC AAA ATC YTR TAT CAA AGC A	/56-FAM/TTCYTTCGG/ZEN/GCCTGTC GGGTYCC/3IABkFQ/

[1]Procopio FA, Fromentin R, Kulpa DA, et al. A Novel Assay to Measure the Magnitude of the Inducible Viral Reservoir in HIV-infected Individuals. EBioMedicine 2015;2:874-83.

[2]Colby DJ, Trautmann L, Pinyakorn S, et al. Rapid HIV RNA rebound after antiretroviral treatment interruption in persons durably suppressed in Fiebig I acute HIV infection. Nat Med 2018;24:923-6.

Supplemental Table 4. Antibodies used for flow cytometry.

Antibodies	**Clone**	**Manufacturer**
Live/dead FVS780	N/A	BD Biosciences, San Jose, CA
CD8 BV510	SK1	BD Biosciences, San Jose, CA
CD14 BV510	MϕP9	BD Biosciences, San Jose, CA
CD19 BV510	SJ25C1	BD Biosciences, San Jose, CA
CD3 APC R700	UCHT1	BD Biosciences, San Jose, CA
CD4 BV421	RPA-T4	BD Biosciences, San Jose, CA
HLA-DR BB515	G46-6	BD Biosciences, San Jose, CA
CD25 PE	MA251	BD Biosciences, San Jose, CA
CD69 FITC	FN50	BD Biosciences, San Jose, CA
TIM-3 PE	7D3	BD Biosciences, San Jose, CA
PD-1 BV786	EH12.1	BD Biosciences, San Jose, CA
TIGIT PerCP eF710	MBSA43	eBioscience, San Diego, CA



Supplemental Figure 1. Gating strategy for flow cytometry experiments. Cells were gated on lymphocyte, singlet, live CD3+CD8-/CD14-/CD19- and then gated for activation and exhaustion markers as above.