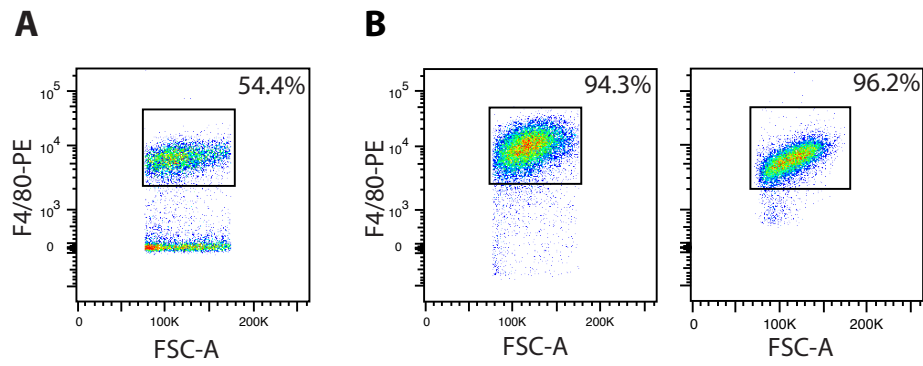


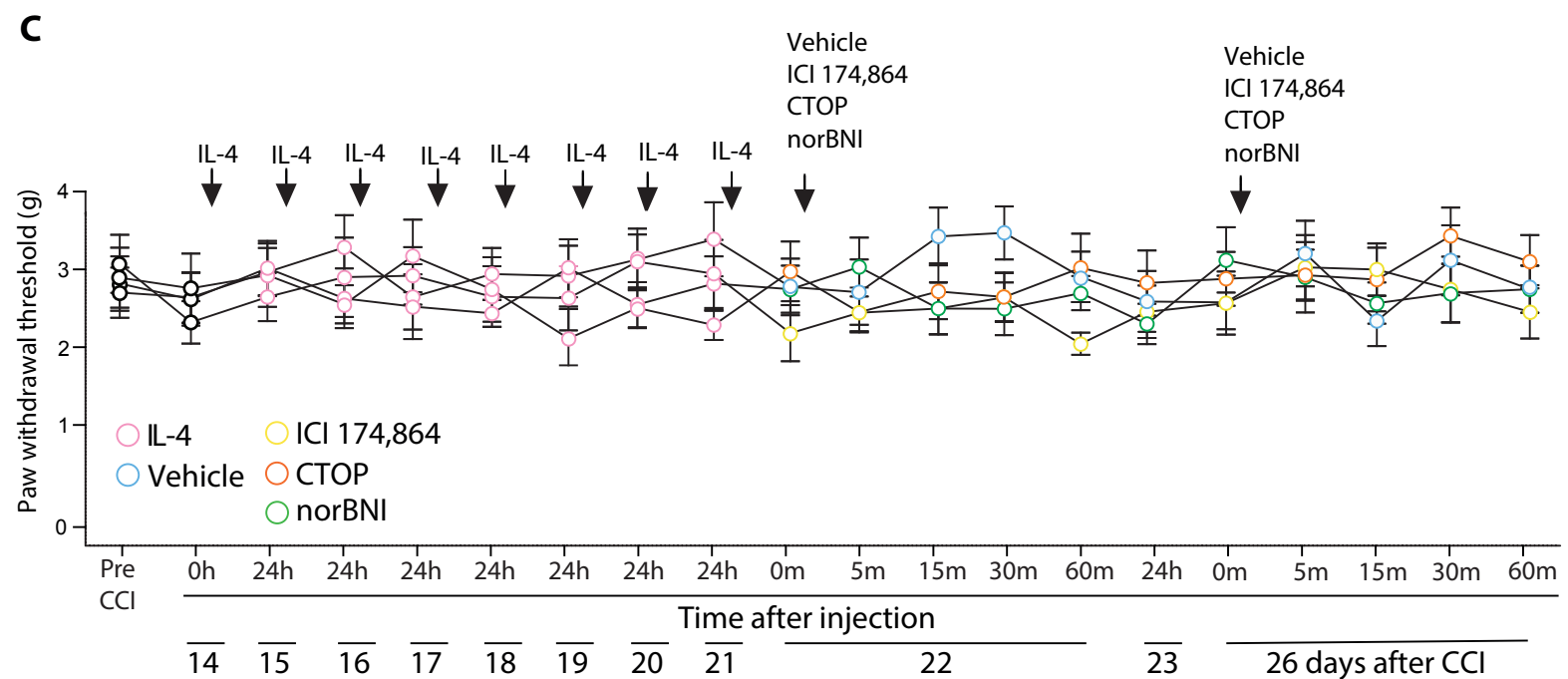
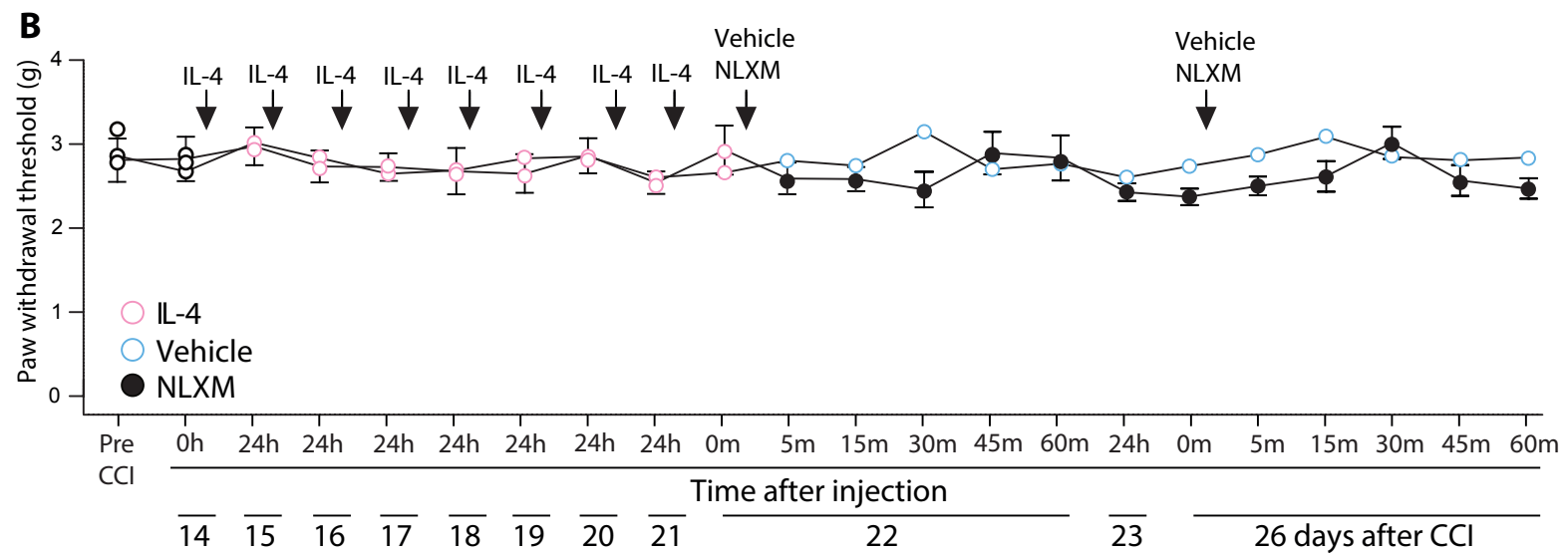
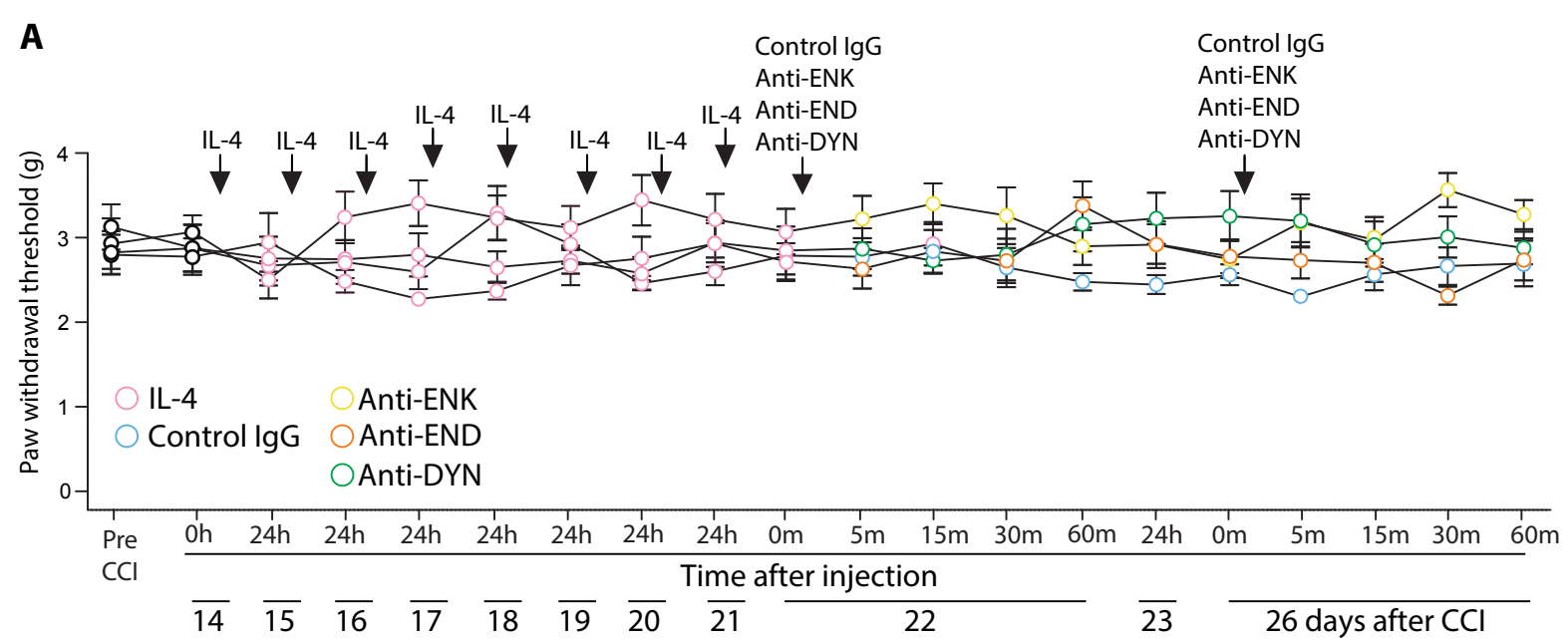
**Supplemental Figure 1. IL-4 application at damaged nerves does not alleviate heat hypersensitivity.** IL-4 (200 ng) was injected daily on days 14–21 after CCI at the CCI site. Heat paw withdrawal latencies were measured before, 5–60 min and 24 h after each injection until day 22, and then on day 23 (48 h after the last injection), and on day 26 after CCI (120 h after the last injection). The latencies were measured in hind paws ipsilateral and contralateral to CCI by the Hargreaves test. Control group was tested accordingly. Arrows indicate injections.  $P > 0.05$  vs. vehicle; 2-way repeated-measures ANOVA. Data are represented as mean  $\pm$  SEM.  $n = 9$  animals per group.



**Supplemental Figure 2. No changes in mechanical sensitivity in paws contralateral to CCI following IL-4 and anti-IL-4R $\alpha$  applied at damaged nerves.** This Figure refers to Figure 1. **(A)** Time-course following IL-4 injection. IL-4 (200 ng) was injected daily on days 14–21 after CCI at the CCI site. Mechanical von Frey thresholds were measured before, 5–60 min and 24 h after each injection until day 22, and then on day 23 (48 h after the last injection), and on day 26 after CCI (120 h after the last injection). **(B)** Time-course following IL-4 and anti-IL-4R $\alpha$  injection. Anti-IL-4R $\alpha$  (6  $\mu$ g) was injected with IL-4 (200 ng) on day 21 after CCI (when IL-4 was applied last time), and again alone (without IL-4) on days 22 and 26 after CCI. Von Frey thresholds were measured before and 24 h after each IL-4 injection (on days 14–21), 5 min after IL-4 and anti-IL-4R $\alpha$  co-injection (on day 21), before and 5–60 min (on day 22) and 24 h (on day 23) after the second anti-IL-4R $\alpha$  injection, and before and 5–30 min after the third anti-IL-4R $\alpha$  injection (on day 26). Control groups were tested accordingly. Arrows indicate injections.  $P > 0.05$  vs. control (vehicle or control IgG); 2-way repeated-measures ANOVA. Data are represented as mean  $\pm$  SEM.  $n = 9$  animals per group.



**Supplemental Figure 3. Flow cytometric verification of purity of macrophages isolated by F4/80+ IMS from damaged nerves.** (A and B) Representative dot blot plots showing that F4/80+ macrophages constituted 54% of all cells isolated from damaged nerves (on day 22 after CCI) before IMS (A), but they represented 94% (left blot) to 96% (right blot) of all cells after IMS (B). F4/80+ macrophages are shown inside the rectangular gates. The data were analyzed using FlowJo software.



**Supplemental Figure 4. No changes in mechanical sensitivity in paws contralateral to CCI following application of IL-4 with opioid peptide antibodies or opioid receptor antagonists at damaged nerves.** This Figure refers to Figure 8. (A) Time-course following application of IL-4 (200 ng) with antibodies to opioid peptides, anti-ENK (2  $\mu$ g), anti-END (2  $\mu$ g), or anti-DYN (4  $\mu$ g). (B) Time-course following application of IL-4 (200 ng) with peripherally restricted opioid receptor antagonist NLXM (10  $\mu$ g). (C) Time-course following application of IL-4 (200 ng) with antagonists selective at  $\delta$ - (ICI 174,864; 8  $\mu$ g),  $\mu$ - (CTOP; 2  $\mu$ g), and  $\kappa$ -receptors (norBNI; 20  $\mu$ g). Antibodies and antagonists were injected 24 h after the last IL-4 injection (on day 22 after CCI) and again 5 days after IL-4 injection (on day 26 after CCI). Von Frey thresholds were determined before and 24 h after each IL-4 injection (on days 14–21), before and 5–60 min after the first injection of antibodies and antagonists (on day 22), once on day 23 (24 h after the first injection of antibodies and antagonists), and on day 26, before and 5–60 min after the second injection of antibodies and antagonists. Control groups were tested accordingly. Arrows indicate injections.  $P > 0.05$  vs. control (control IgG or vehicle); 2-way repeated-measures ANOVA. Data are represented as mean  $\pm$  SEM.  $n = 9$  animals per group.