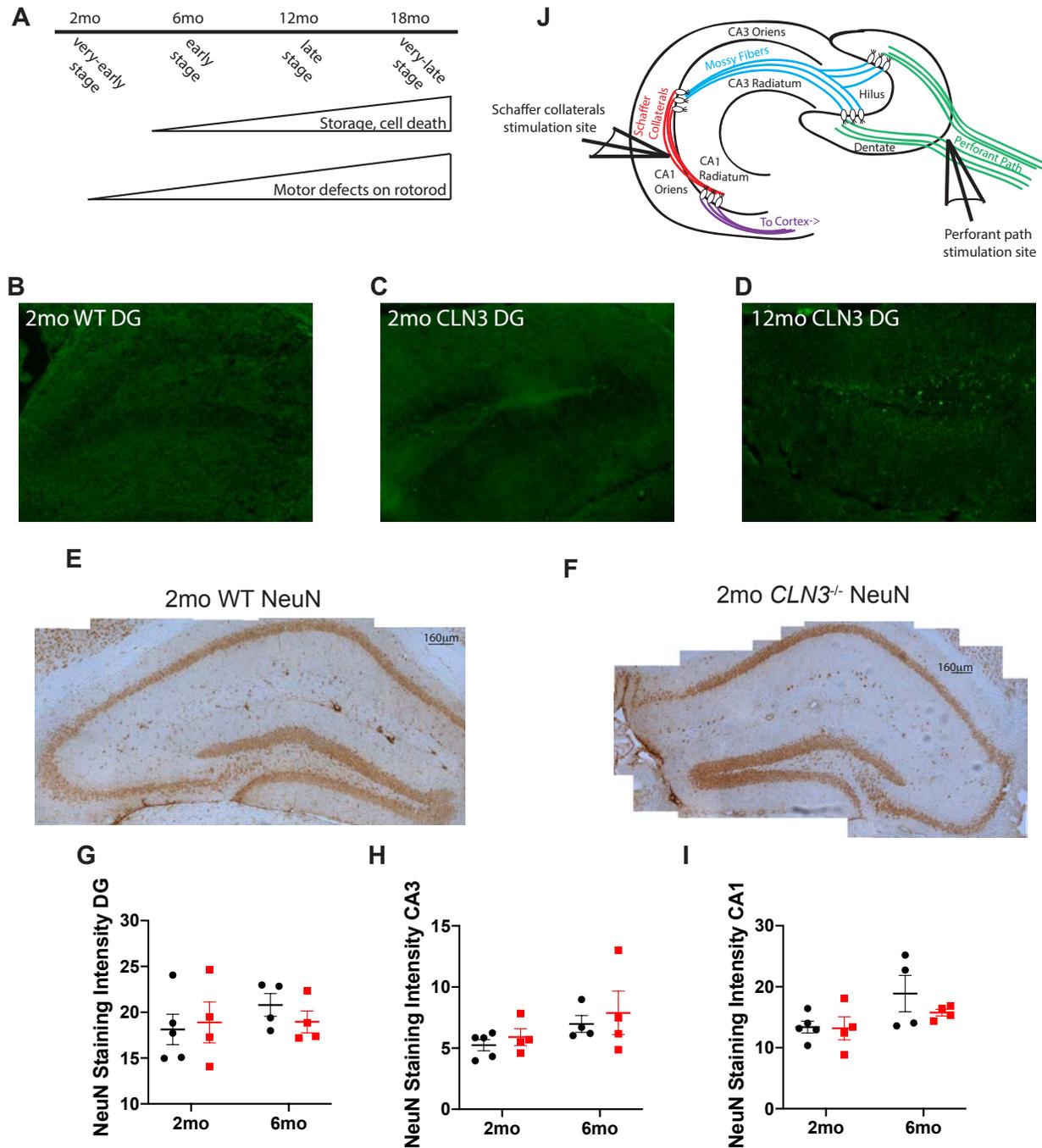
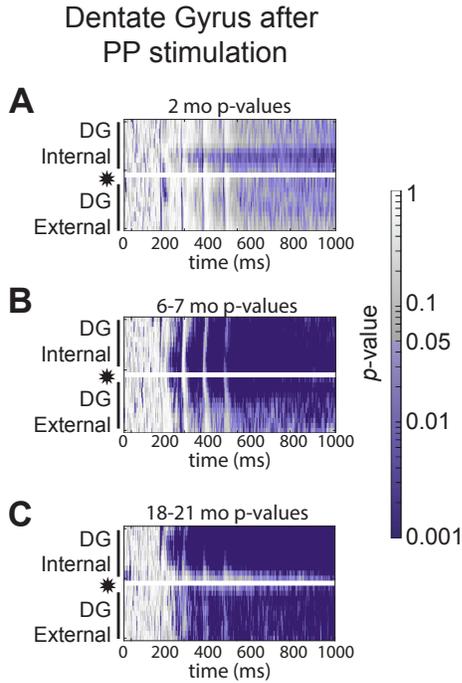


**Supplementary Figures:**



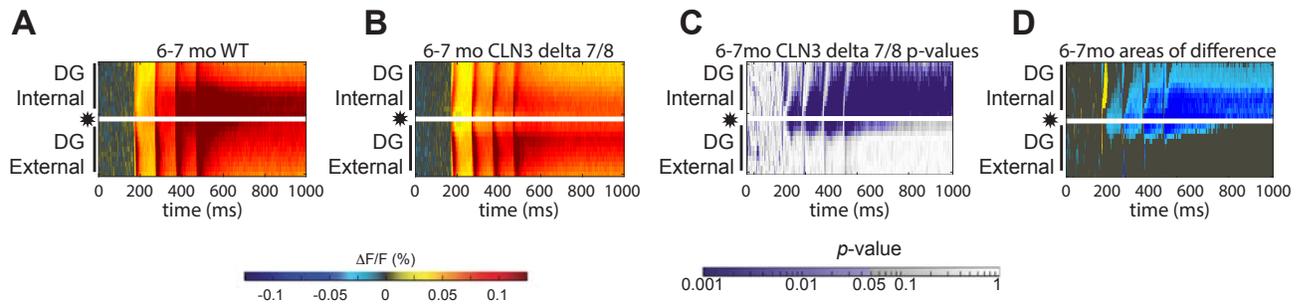
**Supplementary Figure 1: In early-stage disease there is no storage accumulation or neuronal loss. (A)** Timeline of disease progression in the *CLN3<sup>-/-</sup>* mouse. **(B-D)** Representative 10X images of the dentate gyrus (DG) show little autofluorescent storage material in 2-month-old WT or *CLN3* disease mice. However, by 12 months, there is substantial accumulation. **(E-I)** At 2-months of age there is no *CLN3*-disease

associated difference in staining of the neuronal marker NeuN in any of the major hippocampal subfields. **E-F** are representative NeuN images. **G-I** are quantification of staining intensity. N=4-5 animals / condition, mean +/- SEM shown, two-way ANOVA all p-values >0.05. **(J)** Schematic of hippocampal circuit and location of electrode placement for perforant path (as in studies of Fig. 1,2,4,5) and Schaffer collateral (as in studies of Fig. 3) stimulation studies.



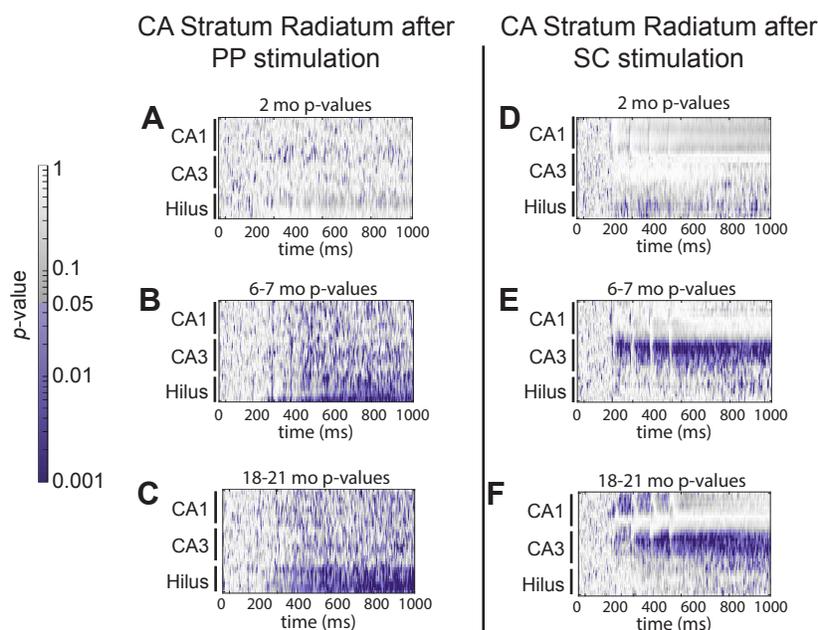
**Supplementary Figure 2: *p*-value maps confirm decreased activation of the dentate gyrus in response to PP stimulation in *CLN3*<sup>-/-</sup> mice.**

Results of permutation sampling statistical analysis (1000 permutations) of data from Figure 1 are shown with *p*-values <0.05 in purple. Dentate gyrus activation in response to perforant path stimulation is statistically different between WT and *CLN3*<sup>-/-</sup> slices at **(A)** 2 months, with more regions of statistical difference in the internal blade as compared to the external blade of the DG, **(B)** 6-7 months, and **(C)** 18-21 months of age. Group sizes (n=slices, N=mice): 2mo WT n=24, N=6; 2mo *CLN3*<sup>-/-</sup> n=26, N=5; 6-7mo WT n=23, N=5; 6-7mo *CLN3*<sup>-/-</sup> n=30, N=8; 18-21mo WT n=25, N=7; 18-21mo *CLN3* n=26, N=7.



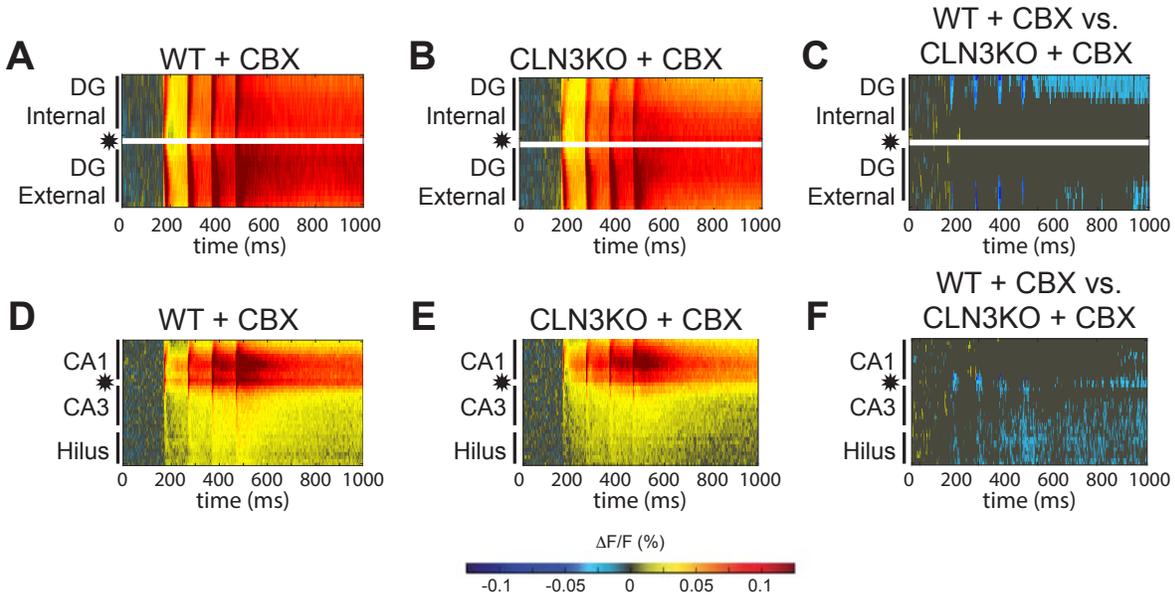
**Supplementary Figure 3: The  $CLN3^{\Delta ex7/8}$  mouse, which models the most common human CLN3 mutation, also demonstrates hypoexcitability of the dentate gyrus early in the disease course.**

(A) Stimulation of the perforant path (location indicated by star) reveals robust excitation of the dentate gyrus (DG) in 6-7 month-old wild-type (WT) hippocampal slices, shown as a raster plot of average fluorescence change ( $\Delta F/F_{WT}$ ). (B) By 6-7 months of age, the  $CLN3^{\Delta ex7/8}$  DG is hypoexcitable (as measured by  $\Delta F/F_{CLN3KO}$ ) as compared to WT after PP stimulation. (C) Results of permutation sampling statistical analysis (1000 permutations) comparing WT and  $CLN3^{\Delta ex7/8}$  mice are shown with  $p$ -values < 0.05 in purple. (D) . Regions of the raster plot with  $p > 0.05$  are masked in gray. For regions with  $p < 0.05$ , the difference in fluorescence change ( $\Delta F/F_{WT} - \Delta F/F_{CLN3KO}$ ) is shown. Cooler colors indicate relative hypoexcitability of the  $CLN3^{\Delta ex7/8}$  DG. Group sizes ( $n$ =slices,  $N$ =mice): WT  $n=18$ ,  $N=4$ ;  $CLN3^{-/-}$   $n=24$ ,  $N=5$ . Panel A reproduced from Figure 1D.



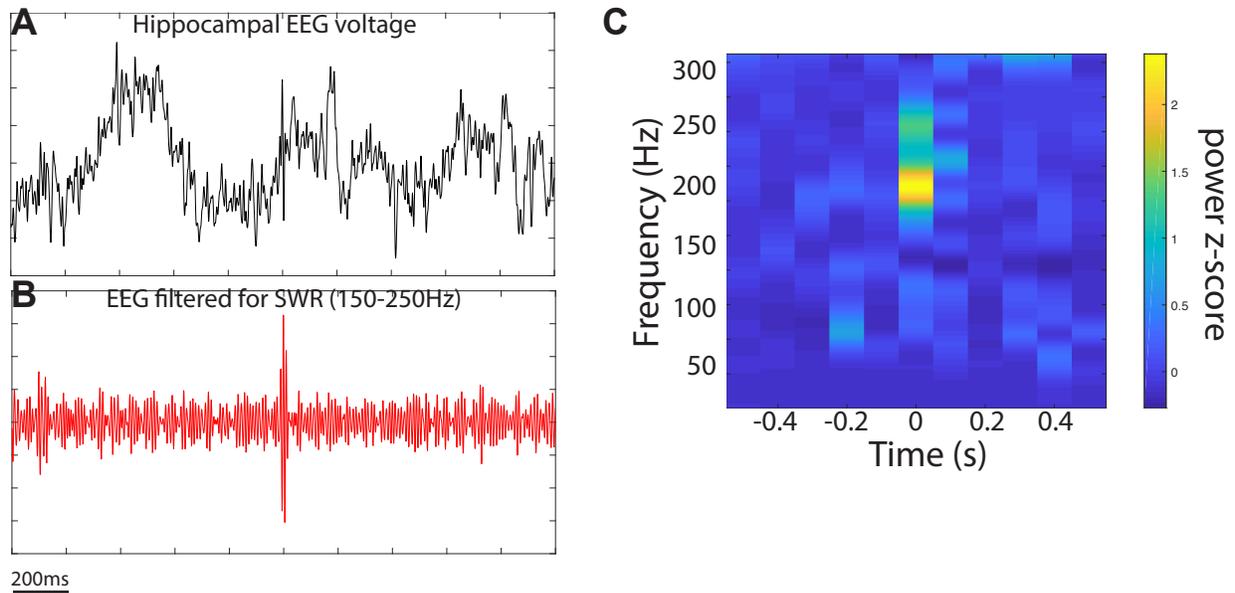
**Supplementary Figure 4:  $p$ -value maps comparing activation of the CA stratum radiatum of WT and  $CLN3^{-/-}$  mice in response to perforant path (PP) and Schaffer collateral (SC) stimulation.**

Results of permutation sampling statistical analysis (1000 permutations) of data from Figure 2 and 3 are shown with  $p$ -values  $< 0.05$  in purple. Hilus, CA3 and CA1 activation in response to perforant path stimulation is minimally different between WT and  $CLN3^{-/-}$  slices at **(A)** 2 months. However, statistically significant regions of hypoexcitability in  $CLN3^{-/-}$  slices is apparent at **(B)** 6-7 months, and **(C)** 18-21 months of age. Group sizes panels A-C (n=slices, N=mice): 2mo WT n=24, N=6; 2mo  $CLN3^{-/-}$  n=26, N=5; 6-7mo WT n=23, N=5; 6-7mo  $CLN3^{-/-}$  n=30, N=8; 18-21mo WT n=25, N=7; 18-21mo  $CLN3^{-/-}$  n=26, N=7. Similar results are seen in response to SC stimulation, with minimal differences at **(D)** 2 months, but clear genotype-associated differences at **(E)** 6-7 months, and **(F)** 18-21 months of age. Group sizes panels D-F (n=slices, N=mice): 2mo WT n=13, N=5; 2mo  $CLN3^{-/-}$  n=11, N=5; 6-7mo WT n=13, N=4; 6-7mo  $CLN3^{-/-}$  n=11, N=3; 18-21mo WT n=13, N=4; 18-21mo  $CLN3^{-/-}$  n=14, N=4.



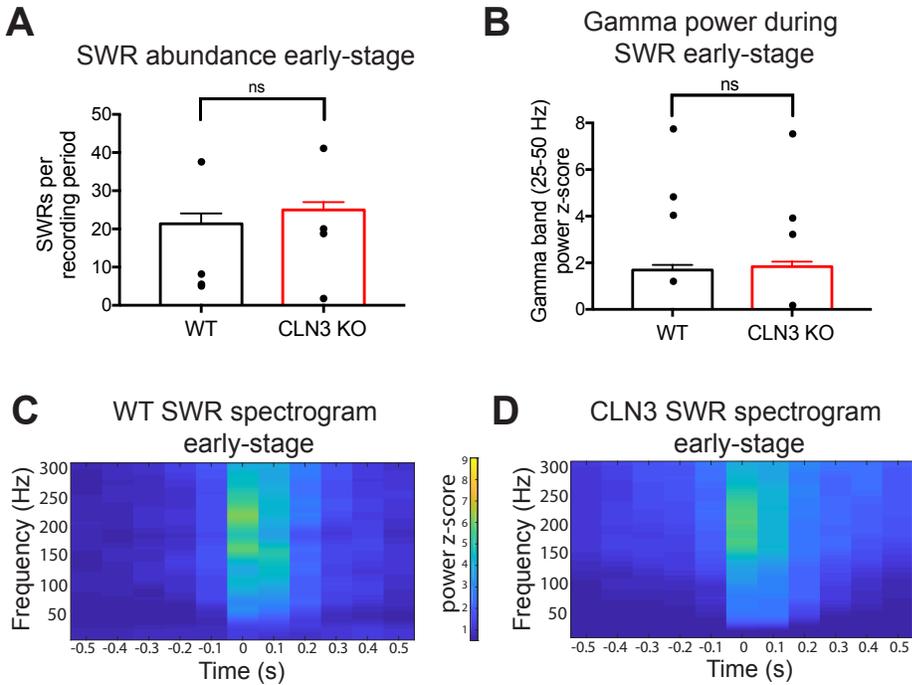
**Supplementary Figure 5: Carbenoxolone (CBX), a drug that reduces storage burden, cannot correct hypoexcitability of the dentate gyrus, and worsens hypoexcitability of the hilus and CA3 in early-stage *CLN3* disease.**

(A-C) Hippocampal slices from 2-month-old mice treated with 2 weeks of 20mg CBX, a dose that reduces storage burden and normalizes endocytosis in vivo, show residual hypoexcitability of the dentate gyrus after perforant path (PP) stimulation, as measured by fluorescence response in CBX-treated WT vs *CLN3*<sup>-/-</sup> slices. (D-F) The hilus and CA3 regions of slices from CBX-treated *CLN3*<sup>-/-</sup> mice showed worsening hypoexcitability after Schaffer collateral (SC) stimulation. Group sizes (n=slices, N=mice): 2mo PP stimulation WT n=15, N=3; 2mo PP stimulation *CLN3*<sup>-/-</sup> n=25, N=5; 2mo SC stimulation WT n=17, N=4; 2mo SC stimulation *CLN3*<sup>-/-</sup> n=20, N=5. Areas of statistical significance identified using a permutation sampling method with 1000 permutations, for regions of significance with  $p < 0.05$ , the difference in fluorescence change ( $\Delta F/F_{WT} - \Delta F/F_{CLN3KO}$ ) is shown. Panel B reproduced from Figure 5B. Panel E reproduced from Figure 5E.



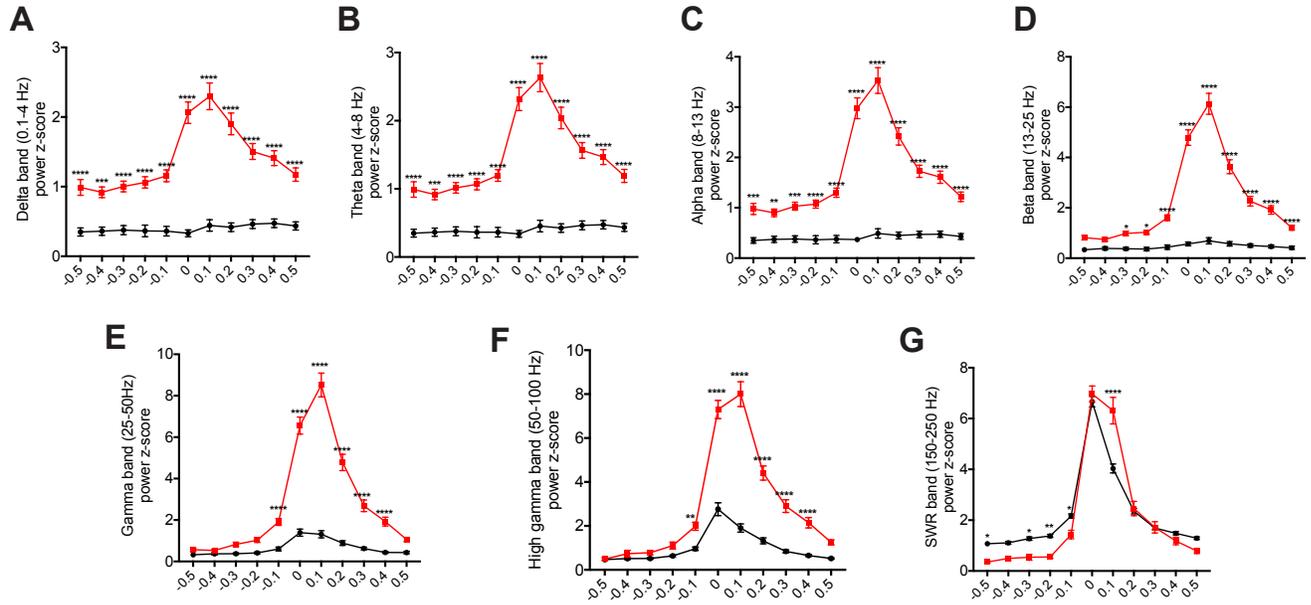
**Supplementary Figure 6: Example sharp wave ripple (SWR) detection from a WT animal.**

(A) Example 2 second raw EEG recording and (B) EEG signal filtered at 150-250Hz to show a SWR. (C) Example spectrogram analysis of the 1 second surrounding a SWR peak recorded from a WT hippocampus.



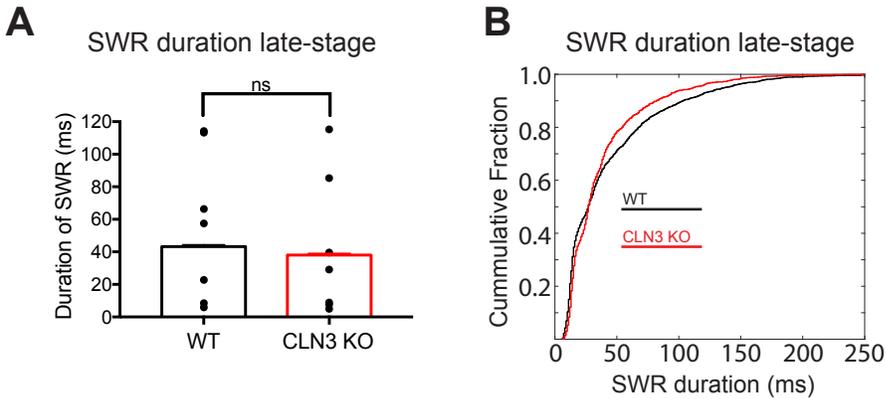
**Supplementary Figure 7: In early stage disease there are no differences between WT and *CLN3*<sup>-/-</sup> hippocampal sharp wave ripple (SWR) abundance or gamma power modulation during SWRs.**

**(A)** Quantification of SWRs per 30 min recording period from hippocampal EEG recordings of 6-month-old WT (black) and *CLN3*<sup>-/-</sup> (red) mice reveals no difference in SWR abundance (Mann-Whitney test  $p=0.51$ ). **(B)** There was also difference in peak gamma power during an SWR (two-way ANOVA to evaluate animal and genotype variation,  $p=0.02$  for genotype effect). **(C-D)** Average spectrograms from SWRs are similar between 6-month-old WT and *CLN3* mice. Group sizes WT: 37 thirty-minute recording periods, 1334 SWRs,  $N=4$  mice; *CLN3*<sup>-/-</sup>  $n=84$  thirty-minute recording periods, 2040 SWRs,  $N=4$  mice. Bar graphs show mean  $\pm$  SEM SWR number from all recording periods, black points show mean SWR abundance for each mouse.



**Supplementary Figure 8: Unlike in control mice, hippocampal sharp wave ripples (SWR) in *CLN3*<sup>-/-</sup> mice trigger increased power in all frequency bands that lags behind and persists longer than the peak power in the SWR band.**

In the major EEG frequency bands including (A) delta (0.1-4 Hz), (B) theta (4-8 Hz), (C) alpha (8-13 Hz), (D) beta (13-25 Hz), (E) gamma (25-50 Hz) and (F) high gamma (50-100 Hz), there is increased power after a SWR in *CLN3*<sup>-/-</sup> (red) hippocampus as compared to WT (black). (G) While, in the SWR frequency range (150-250 Hz) there is no difference in power at the SWR peak. Group sizes for panels: WT n=2059 SWRs, N=7 mice; *CLN3*<sup>-/-</sup> n=1363 SWRs, N=7 mice. Graphs show mean +/- SEM. Groups compared using repeated measures two-way ANOVA followed Sidak's multiple comparison test. \*p<0.05, \*\*p<0.01, \*\*\*p<0.001, \*\*\*\*p<0.0001.



**Supplementary Figure 9: In late-stage disease, there is no difference between WT and *CLN3*<sup>-/-</sup> hippocampal sharp wave ripple (SWR) duration.**

The duration of SWRs was calculated for all hippocampal ripples detected in WT (black) and *CLN3*<sup>-/-</sup> (red) 11-month-old mice. There was no significant difference in the **(A)** mean ( $p=0.43$ , Mann-Whitney test) or **(B)** cumulative fraction duration between groups. Bar graphs show mean  $\pm$  SEM duration of all SWRs, black points show mean SWR duration for each mouse.