1 Supplementary Figures





5 Time course of pulse pressure (upper panel) and heart rate (lowe panel) of perfused hearts, 6 from the same mice shown in figure 1, under baseline condition and after addition of Dsg2-LP. 7 VE-Cad-LP served as peptide control. (n=5 mice for Dsg-LP and Pg KO and WT; n=3 mice for 8 VE-Cad-LP). Arrow indicates the time point of addition of Dsg2-LP or VE-Cad-LP. Average of 9 baseline values and 10 minutes of Dsg2-LP were compared. Two-tailed paired Student's t-test 10 with 95 % confidence levels was performed and no significant changes were observed.

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Supplementary figure 2. Dsg2 and Cx43 membrane localization analyzed by STED
 microscopy in WT and Pg KO ventricular cardiac slices.

A, STED images of ICDs of WT and Pg KO mice treated with Dsg2-LP for 1 hour and stained
for Dsg2 (red) and Cx43 (green) (four ICDs per mouse were analyzed for quantification, n=4
mice per condition). Scale bar: 2µm. Colocalization of both proteins was analyzed and
colocalized pixels are visualized in white. B, Quantification of colocalized pixels. **P*<0.05.
Repeated measures two-way ANOVA with Sidak post-hoc test was performed.



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Supplementary figure 3. Dsg2-LP did not alter desmosomal cadherin expression or
 localization in HL-1 cells.

A, Western blot analysis of desmoplakin (Dp), desmoglein 2 (Dsg2), N-cadherin (N-Cad) or
plakoglobin (Pg) after 24 h treatment with Dsg2-LP. □-tubulin (α-tub) was used as a loading
control B, Quantitative analysis of Western blots performed in A (n=4). One-way ANOVA with
Sidak post-hoc test was performed and no significant changes were observed. C,
Representative immunostaining of HL-1 cells stained for N-Cad, Pg and Dp treated with Dsg2LP for 24h (n=3), scale bar: 10µm.

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A, Western blot analysis of Cx43 phosphorylation at serine S368 (p-Cx43) and total Cx43 after
1 h treatment. □-tubulin (α-tub) was used as a loading control. B, C, Quantitative analysis of
Western blots performed in A (n=3 mice per group). Two-way ANOVA with repeated measures
and matching both factors with Sidak post-hoc test was performed for B. Two-tailed unpaired
Student's t-test with 95 % confidence levels was performed for C.



Supplementary Figure 5. MEA of HL-1 cardiomyocyte treated with carbenoxolone or
 Dsg2-IP.

A, Representative MEA heats maps of HL-1 cardiomyocytes treated with the GJ inhibitor
carbenoxolone or the inhibitory peptide Dsg2-IP. Reference electrode is marked by (*). B, C,
Corresponding analysis of conduction velocity and standard deviation of beat-to-beat-intervals
(SDNN) of MEA experiments described in A (n=3). **P*<0.05. One-way ANOVA with Dunnett
post-hoc test.



2 Supplementary Figure 6. AFM force mapping on living cardiomyocytes.

A, Schematic of AFM set-up on living cells. An area spanning the cell-cell boder (marked in
cyan) was selected for force spectroscopy as shown in the overview topography image. Scale
bar overview: 10µm; insert: 1µm. B, Representative immunostaining of cardiomyocytes
isolated form neonatal Dsg2 KO mice and WT littermates stained for Dsg2 (red), N-cadherin
(magenta), F-actin (green) and the nucleus (DAPI, blue) paralleled to AFM experiments shown
in Fig. 5 (n=7 mice per phenotype). Scale bar: 10µm.

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Full Unedited gel for Figure 3B

Full Unedited gel for Figure 41



pCx43 S368



α-tub



Total Cx43

Full Unedited gel for Figure 4J









α-tub

Full Unedited gel for Figure 7A



Full Unedited gel for Supplementary figure 3A

 Dp
 Dsg2
 N-Cad
 Pg
 α-tub

Full Unedited gel for Supplementary figure 4A



