

Supplementary Figure 1. AEC2s are the Principal Progenitor of AEC2s During Epithelial Regeneration in the LPS Model of Lung Injury. *SftpcCreERT2;mTmG* mice were treated with or without LPS and harvested at Day 27. The percent of proSPC+ cells (AEC2s) that were GFP+, as determined by immunostaining of lung sections, is shown. Mean ± SEM indicated.



Supplementary Figure 2. scRNAseq of Epithelial Cells Isolated from Naïve and LPS-injured mice. A,B) *SftpcCreERT2;mTmG* mice were treated with or without LPS. At Day 7, lungs were digested, stained for EpCAM, CD45, and T1α, and subjected to fluorescence-activated cell sorting (FACS). Tomato+ GFP- CD45- EpCAM+ T1α+ (Non-AEC2 Epithelial) and Tomato- GFP+ (AEC2) cells from naïve mice and Tomato- GFP+ (Injured AEC2-Derived) cells from LPS-treated mice were loaded into a 10X Genomics Chromium Single Cell 3' Solution. Cells and gel beads containing barcoded oligonucleotides and reagents were encapsulated in droplets. Cells were lysed, and mRNA was reverse transcribed to cDNA, which introduced cell barcodes and UMI sequences. cDNA was amplified by PCR and sequenced, and cells were subjected to unsupervised clustering based on their transcriptomes. B) Flow cytometry gating strategy. The experiment with one mouse per treatment group was performed on two separate occasions, for a total of 2 mice/group.



Supplementary Figure 3. scRNAseq of Epithelial Cells Isolated from Naïve and LPS-injured mice. The number of cells sequenced (A) and number of genes (B), transcripts (UMIs) (C,E), and mitochondrial reads (D,E) per cell after quality filtering. n=2 mice/group.



Supplementary Figure 4. Unsupervised Clustering. A) Locations within the tSNE plot of Naïve Non-AEC2 Epithelial, Naïve AEC2, and Injured AEC2-Derived cells from both experiments. B) Expression of GFP (natural log of normalized counts). C) Number and percent of cells in each cluster. D) Heatmap showing the 20 most differentially expressed genes in each cluster compared to all other clusters ranked in order of Bonferroni-corrected p-value. n=2 mice/group.





Supplementary Figure 5. scRNAseq Reveals Naïve Lung Epithelial Cell Types. Gene expression (natural log of normalized counts) of canonical Club Cell (A), Basal Cell (B), Ciliated Cell (C), AEC1 (D), and AEC2 (E) markers. Dark Orange Circles (Cluster 13): Scgb1a1+ Sftpc+ Cells, Light Orange Circles (Cluster 9): Club Cells, Yellow Circles (Cluster 15): Ciliated Cells, Blue Circles (Cluster 7): Basal Cells, Green Circles (Cluster 14): AEC1s, Red Circles (Cluster 5): AEC2s. n=2 mice/group.

All Naive

Foxi1

Ccdc39 1.6 1.2 0.8 0.4 0.0











Supplementary Figure 6. scRNAseq Reveals Rare Contaminating Macrophages, Dendritic Cells, Eosinophils, Endothelial Cells, and Fibroblasts. Gene expression (natural log of normalized counts) of macrophage (A), dendritic cell (B), eosinophil (C), neutrophil (D), lymphocyte (E), endothelial cell (F), and fibroblast (G) markers. Pink Circles (Cluster 11): Hematopoietic Cells, Purple Circles (Cluster 12): Endothelial Cells/Fibroblasts.











Anti-Hamster IgG Anti-Rabbit IgG Anti-Chicken IgG

Supplementary Figure 7. AQP5 is Expressed by AEC2s. Immunostaining of lung sections from naïve *SftpcCreERT2;mTmG* mice for GFP, T1 α , and AQP5. Arrowheads indicate AQP5 staining in AEC2s, particularly at the apical surface, as indicated by breaks in the T1 α (AEC1) staining. Scale bar = 10 µm.



2.0 1.5 1.0 0.5 0.0

Ccne1

1.6 1.2 0.8

0.4 0.0

Cyclin E1

Supplementary Figure 8. scRNAseg Reveals Proliferating, Cell Cycle Arrest, and Transdifferentiating AEC2-Derived Subpopulations. Gene expression levels of A) S phase markers, B) G2 phase markers, C) Mitosis markers, D) G1 arrest markers, E) CDK inhibitors, F) AEC1 markers, and G) AEC2 markers in the Naïve AEC1 and AEC2 and Injured AEC2-Derived cells. Blue Circles (Cluster 10): Proliferating Subpopulation, Green Circles (Cluster 8): Cell Cycle Arrest Subpopulation, Red Circles (Cluster 6): Transdifferentiating Subpopulation. H) *Igfbp2* expression. A-H) Gene expression shown in natural log of normalized counts. I) Enlarged image of mature AEC1 cluster (Cluster 14) with cells derived from the Naive Non-AEC2 Epithelial Cells indicated in blue and cells derived from the Injured AEC2-Derived Cells in orange (left panel) and GFP expression (right panel). J) Number and percent of Injured AEC2-Derived Cells localized to each cluster. K) Percent of Injured AEC2-Derived Cells expressing p15 as determined by costaining with anti-GFP antibody and p15 RNA probe. Mean ± SEM shown. *p<0.05 by Mann-Whitney test. L) Gene Ontology terms for the top 3 biological processes enriched in each cluster, ranked in order of ascending p-value corrected for Elim pruning. n = 4 mice/group.









LPS.







Supplementary Figure 9. scRNAseq Reveals Mildly Perturbed Gene Expression in Other Injured AEC2-Derived Cells. A) Unsupevised clustering at the indicated resolution. B) Canonical correlation analysis of the Other Injured AEC2-Derived and Naive AEC2 populations. Left Panel: tSNE plot showing the overlap of the injured (blue) and naive cells (red). Right Panel: Top genes associated with the first three CCA dimensions, with AEC2 markers highlighted in red. C) Hierarchical clustering of identified subpopulations. D) Heatmap showing the most differentially genes in Naive AEC2 cluster compared to Other Injured AEC2-Derived Cell clusters ranked in order of Bonferroni-corrected p-value. E) Gene Ontology terms for the top 10 biological processes enriched in the Other Injured AEC2-Derived Cells compared to the Naive AEC2s.

В.



C.



Supplementary Figure 10. Expression of TGF β Pathway and EMT Genes. A) Gene expression (natural log of normalized counts) of TGF β signaling molecules. B) Heatmap of TGF β pathway gene expression in the indicated subpopulations. C) Percent of Cell Cycle Arrest subpopulation expressing indicated genes. D) Percent of AEC2-Derived Cells expressing at least 3 RNA molecules of *TGF\beta2* or Integrin β 6 as determined by costaining with anti-GFP antibody and RNA probes. Mean ± SEM shown. *p<0.05 by Mann-Whitney test. E) Immunofluorescence staining for GFP and in situ hybridization for *TGF\beta2*, Integrin β 6, and p15. Arrowheads indicate double positive cells. Scale bar 10 µm. n ≥ 4 mice/group. F) Gene expression (natural log of normalized counts) of epithelial and mesenchymal markers. G) Heatmap of expression of mesenchymal markers.

TGFβ Pathway Genes

В.





D.



Е.





C.





Supplementary Fig. 11. Total and Phosphorylated Smad3 Levels Increase in Cultured AEC2s. Primary rat AEC2s were cultured. Western blotting of cell lysates for pSmad3, Smad3, and actin. Vertical white lines indicate that the lanes were run on the same gel but were noncontiguous.



Supplementary Figure 12. Conditional knockout of TGFβRII in Mouse Tails or Cultured AEC2s. PCR performed on gDNA extracted from tails of *Tgfbr2^{wt/wt}, Tgfbr2^{t/wt}, or Tgfbr2^{t/t}* mice or AEC2s isolated from *Tgfbr2^{t/t}* mice and transduced with AdCre or AdGFP. Expected size of each allele is shown. Representative of n=3.



Supplementary Figure 13. Lineage Relationship Between Subpopulations. A) Pseudotime analysis was performed on scRNAseq data using Monocle2 with various input parameters: top 1000 most differentially expressed genes (1st panel), top 2000 differentially expressed genes (2nd panel), excluding Naive AEC2 populations (3rd, 4th panel), or with downsampling to reduce the number of injured naive AEC2 cells (3rd panel). B) RNA velocity projections overlayed on tSNE (left) and inferred trajectory start points (middle) or endpoints (right) based on Markovchain process. Dark blue indicates the highest density of end or start points. C) PAGA analysis to infer branch points, force directed graph layout (left) or PAGA representation (right). D) Venn diagram showing the overlap of genes highly differentially expressed by each subpopulation. n=2 mice/group.