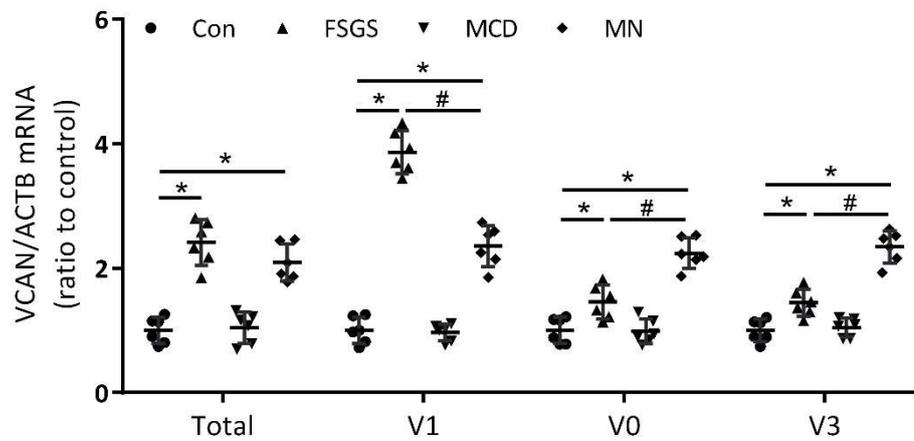


## SUPPLEMENTARY MATERIAL



**Figure S1.** RT-PCR analysis of total versican, versican V1, V0 and V3 levels in the tubulointerstitial tissues of normal controls (Con), focal segmental glomerulosclerosis (FSGS), minimal change disease (MCD) and membranous nephropathy (MN) patients (n=6). For statistical analysis, one-way ANOVA with Tukey's post hoc test was used. \*,  $P < 0.05$  compared with control. #,  $P < 0.05$  compared with FSGS patients.

**Table S1. Clinical and pathological parameters in the discovery and validation cohorts**

Disease type	Discovery cohort		Validation cohort	
	Normal	FSGS	Normal	FSGS
patients	8	8	20	20
Age (years)	35.8±7.5	30.9±10.8	35.3±6.5	33.1±16.0
Gender (% female)	50	50	40	40
Disease course (months)	-	11.0 (1.2-24.3)	-	9.0 (0.9-22.5)
Proteinuria (g/24h)	-	8.2±3.4	-	7.0±4.0
serum albumin (g/L)	45.2±3.3	22.6±2.8	45.8±3.5	25.1±7.6
serum creatinine (mg/dl)	0.68±0.12	1.82±1.74	0.75±0.18	1.64±1.22
focal segmental lesions (%)	-	16.1 (13.5-16.7)	-	22.8 (11.5-32.8)
tubulointerstitial fibrosis	-	1.0 (0.0-2.0)	-	1.0 (0.0-2.0)

**Table S2. Relationship between DEG level and eGFR decline rate in FSGS patients**

DEGs	Correlation coefficient	P value
VCAN	-0.854	0.007
VCAM1	-0.847	0.008
POSTN	-0.844	0.008
ADAMTS9	-0.840	0.009
TGM2	-0.800	0.017
ARG2	-0.791	0.020
ITGAV	-0.790	0.020
CDH6	-0.751	0.032
GBP2	-0.743	0.035
STAT1	-0.743	0.035
FBN1	-0.740	0.036
C3	-0.734	0.038
HAVCR2	-0.730	0.040
ICAM1	-0.721	0.043
UGT1A6	-0.716	0.046
LRRK2	-0.707	0.050
DNPH1	0.710	0.049
RN7SKP137	0.710	0.048
TECRP2	0.712	0.048
SNORD8	0.715	0.046
SNORD9	0.719	0.045
SNORD65	0.720	0.044
HAUS4	0.723	0.043
UBL5P2	0.727	0.041
ACOT4	0.730	0.040
GADD45G	0.731	0.040
RHCG	0.736	0.037
SNORD15A	0.738	0.037
LAMTOR5P1	0.741	0.036
RNA5S9	0.741	0.035
SNORD116-1	0.747	0.033
MISP3	0.747	0.033
EPHX1	0.753	0.031
FXYD2	0.756	0.030
ECHDC3	0.767	0.026
RPL29P4	0.776	0.024
RNY4P7	0.780	0.022
TMEM256	0.784	0.021
HBB	0.794	0.019
RPL10P16	0.795	0.018
PM20D1	0.819	0.013
RNY1P13	0.821	0.013
AHCY	0.874	0.005

**Table S3. Primers**

Primer list	
<b>Hs-VCAN-total forward primer</b>	TTCAACCTTAATAGTAACCCATGC
<b>Hs-VCAN-total reverse primer</b>	AAGGTAGGCTGACTTTTCCAGAG
<b>Hs-VCAN-V1 forward primer</b>	TCGTTTTGAGAACCAGACAGG
<b>Hs-VCAN-V1 reverse primer</b>	CTCAAATCACTCATTTCGACGTT
<b>Hs-VCAN-V0 forward primer</b>	CAGCAAGCACAAAATTTACC
<b>Hs-VCAN-V0 reverse primer</b>	CTCAAATCACTCATTTCGACCTG
<b>Hs-VCAN-V3 forward primer</b>	TTGAGAACCAGACAGGCT
<b>Hs-VCAN-V3 reverse primer</b>	CAGCGATCAGGTCGTTTA
<b>Hs-ACTB forward primer</b>	CTTGACAAAACCTAACTTGCG
<b>Hs-ACTB reverse primer</b>	TGCTGTCACCTTCACCGTTC
<b>Hs-COL1A1 forward primer</b>	GAGGGCCAAGACGAAGACATC
<b>Hs-COL1A1 reverse primer</b>	CAGATCACGTCATCGCACAAAC
<b>Hs-VCAN-V0/V1 forward primer</b>	AGGTGGTCTACTTGGGGTGA
<b>Hs-VCAN -V0/V1 reverse primer</b>	TGGGGTGGTTGTCACATCAG
<b>Mm-VCAN-total forward primer</b>	TGGCCAGAACGGAAATATCA
<b>Mm-VCAN-total reverse primer</b>	ACTAGCCCGGAGTTTGACCAT
<b>Mm-VCAN-V1 forward primer</b>	TCGTTTTGAGAACCAGACATG
<b>Mm-VCAN-V1 reverse primer</b>	ATCATATCACTCAATCGACGTT
<b>Mm-VCAN-V0 forward primer</b>	GGTCAGAGAAAACAAGACAGGTCCG
<b>Mm-VCAN-V0 reverse primer</b>	TGTTTCTTCACTGCAAGGTTCTCT
<b>Mm-VCAN-V3 forward primer</b>	TGCTTCCCTCTCCCTGATAG
<b>Mm-VCAN-V3 reverse primer</b>	TGCAGAGATCAGGTCGTTTA
<b>Mm-ACTB forward primer</b>	GTGACGTTGACATCCGTAAAGA
<b>Mm-ACTB reverse primer</b>	GCCGGACTCATCGTACTCC
<b>Hs-VCAN-Chip-forward</b>	CTTCCCTCTAGGTCCCCGA
<b>Hs-VCAN-Chip-reverse</b>	ACTTCTTCTCGGGAGCACAC
<b>Hs-VCAN-RIP-forward</b>	GCACTTAACAAACTGGGC
<b>Hs-VCAN-RIP-reverse</b>	TGATGCAGTTTCTGCGAGGAT
<b>Hs-VCAN-RAP-probe</b>	GAATACACCATACTTTCAAGAATCTTAAGTCCTGAA
<b>Hs-VCAN-RAP-forward</b>	AATACTCCCTACAAAATACTCAGGA
<b>Hs-VCAN-RAP-reverse</b>	AAACTTGTATGCCAGTGTTTGT
<b>Hs-LNA antisense site 1</b>	ATTTCAAAGAGTTGATTAGA
<b>Hs-LNA antisense site 2</b>	TCTAATTCTTATGAAACAT
<b>Mm-VCAN-Chip-forward</b>	AGGAAAGACGCGGATTTGTG
<b>Mm-VCAN-Chip-reverse</b>	AGTCCTTAGGAGCGGACGG
<b>Mm-VCAN-RIP-forward</b>	CTGGTCGTGCAACCCCTTA
<b>Mm-VCAN-RIP-reverse</b>	CTGGGTGATGAAGTTTCTGCG
<b>Mm-VCAN-RAP-probe</b>	TATATGTATACAACACTGCAGTTTCAAAAATCTTACATTA
<b>Mm-VCAN-RAP-forward</b>	GTCCAATTAATAACAGAGCCAC
<b>Mm-VCAN-RAP-reverse</b>	AAGGGGTTGCACGACCA

**Table S4. Antibodies used in this study**

Antibody	Catalog no.	Company	Reactivity	Application (dilution)
<b>Versican</b>	ab19345	Abcam	Human, Mouse	IHC (1:150), western (1 µg/ml)
<b>Collagen I</b>	14695	Proteintech	Human, Mouse	Western (1:2000)
<b>CD44</b>	217594	Merck Millipore	Human, Mouse	Western (1:1000)
<b>p-smad3</b>	9520	Cell Signaling Technology	Human, Mouse	Western (1:1000)
<b>Smad3</b>	9523	Cell Signaling Technology	Human, Mouse	Western (1:1000)
<b>C3a</b>	ab11873	Abcam	Human	IF (1:50)
<b>C3a</b>	558250	BD Biosciences	Mouse	IF (1:50)
<b>uPAR</b>	MAB807	R&D Systems	Human	Blocking study (10µg/ml)
<b>uPAR</b>	sc-10815	Santa Cruz	Human, Mouse	IHC (1:150), IF (1:75), IP (5µg/mg protein)
<b>ITGB6</b>	ab240302	Abcam	Human, Mouse	IF (1:75), western (1:2000)
<b>β-catenin</b>	66379	Proteintech	Human, Mouse	western (1:2000), ChIP (2µg per 15µg chromatin)
<b>SRp40</b>	06-1365	Merk Millipore	Human, Mouse	IF (1:75), western (1:500), RIP (5µg per immunoprecipitation)
<b>Rac1</b>	ab33186	Abcam	Human, Mouse	IF (1:50), western (1 µg/mL), IP (5µg/mg protein), RIP (5µg per immunoprecipitation)
<b>U2AF1</b>	ab86305	Abcam	Human	RIP (5µg per immunoprecipitation)
<b>Akt</b>	9272	Cell Signaling Technology	Human, Mouse	western (1:1000)
<b>p-AKT</b>	9271	Cell Signaling Technology	Human, Mouse	western (1:1000)
<b>Histone H3</b>	05-928	Merck Millipore	Human, Mouse	western (1:2000)