## SUPPLEMENTARY MATERIALS

## Belimumab promotes negative selection of activated autoreactive B cells in systemic lupus erythematosus patients

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**Supplementary Figure 1: Gating strategy.** (A) Plots show the gating strategy for major B cell populations, including CD27<sup>-</sup>IgD<sup>+</sup> (transitional and naïve), CD27<sup>+</sup>IgD<sup>+</sup> (unswitched memory - non-CSM) CD27<sup>+</sup>IgD<sup>-</sup> (switched memory - CSM) and CD27<sup>-</sup>IgD<sup>-</sup> (double negative - DN) B cells as well as CD20<sup>+</sup>CD38<sup>hi</sup>CD27<sup>hi</sup> (pre-plasmablast and plasmablast) and CD20<sup>+</sup>CD27<sup>+</sup>CD43<sup>+</sup> (B1) B cells. (B) Plots show the gating strategy for major B cell populations, including CD27<sup>-</sup>IgD<sup>+</sup> (transitional and naïve), CD27<sup>+</sup>IgD<sup>+</sup> (unswitched memory) CD27<sup>+</sup>IgD<sup>-</sup> (switched memory) and CD27<sup>-</sup>IgD<sup>-</sup> (double negative - DN) B cells and CD19<sup>+</sup>CD27<sup>hi</sup>CD38<sup>hi</sup> CD38<sup>hi</sup>CD27<sup>hi</sup> (pre-plasmablast and plasmablast - PB). From the mature CD27<sup>-</sup>IgD<sup>+</sup> B cell compartment , cells were further subsetted into transitional T1 (MTG<sup>+</sup>CD38<sup>+++</sup>CD24<sup>+++</sup>), T2 (MTG<sup>+</sup>CD38<sup>++</sup>CD24<sup>++</sup>), T3 (MTG<sup>+</sup>CD38<sup>+</sup>CD24<sup>+</sup>), activated naïve (MTG<sup>+</sup>CD24<sup>-</sup>), naïve (MTG<sup>-</sup>) and MTG<sup>+</sup>CD24<sup>+</sup>CD38<sup>-</sup> B cells.



**Supplementary Figure 2: Comparison of absolute B cell count/ml before and after belimumab treatment.** Connected dot graphs show changes in the number of total B cells (A), and B cell subsets (B-M) at indicated times (in months) after treatment. Each color represents an individual donor.



Supplementary Figure 3: The frequency and number of CD95 and CD21 expressing B cell

**subsets.** A-C. The frequency and number of CD95 and CD21 expressing B cell subsets. Percent (A) of CD95 positive B cells within the major subsets (HD: n=13, LC: n=12, CB: n=14). Frequency (B) and total number (C) of CD21<sup>lo</sup> cells in major subsets of B cells (HD: n=15, LC: n=14, CB: n=12). \*p <0.05; \*\*p < 0.01; \*\*\*p <0.001; \*\*\*p<0.0001; ns: not significant. Comparisons were performed using Mann Whitney analysis (Suppl Fig 3C) and Kruskal-Wallis Test (Suppl Fig 3A and 3B).



**Supplementary Figure 4: VH, DH, JH and VH-JH paired usage.** Bar graphs show frequencies of VH, DH and JH family usage in patients with (n=14) and without (n=10) belimumab treatment (A, C, E) and 5 individual patients before and after 6 months of treatment (B, D, F). Heatmaps (G-K) show the VH-JH paired usage from individual patients before (top) and after (bottom) treatment.



Supplementary Figure 5: Clonotype distribution of different B cell subsets with and without treatment. Clonotype size ranking is shown on the y-axis, and the frequency of sequences for each clone as a percent of total repertoire is shown on the x-axis. Each line represents an individual patient (lupus control (LC) n=10; chronic belimumab (CB) n=14; pre and post belimumab n=5).



Supplementary Figure 6. Clonal representation of PB in patients with and without

**belimumab treatment.** Stacked bar graphs display the size distribution of the most common individual clonotypes constituting 30% of the repertoire (D30) of plasmablasts from (A) lupus controls (n=7) and (B) chronic belimumab-treated patients (n=13). Individual donors are shown at the bottom of each graph and the most frequently represented VH are color coded. Comparisons of individual genes were performed using Mann Whitney analysis.

Α



**Supplementary Figure 7**. Characteristics of chronic belimumab treated patients at inception. Data from the pre-belimumab screening visit are compared with the study visit (n=15). After >7 years of belimumab treatment patients manifested a significant decrease in SLEDAI score and prednisone dose but changes in anti-dsDNA antibody titers were variable. Comparisons were performed using Wilcoxon Signed Rank test for paired samples. \*p <0.05; \*\*p < 0.01; \*\*\*p <0.001; \*\*\*\*p<0.0001; ns: not significant.

Panel 1	Source	Catalog	Clone
Mitotracker green	Invitrogen	M-7514	
IgD PE	BD	BD 555779	IA6-2
CD3 PE/Texas Red	Beckman Coulter	IM2705U	UCHT1
CD38 PerCP/Cy5.5	Beckman Coulter	A70205	LS198.4.3
CD21 PECy7	BD	BD 561374	B-ly4
CD27 APC	Beckman Coulter	B09983	1A4CD27
CD10 APC/AF700	Beckman Coulter	A86353	ALB1
CD24 APC/AF750	Beckman Coulter	B10738	ALB9
CD19 V450	BD	BD 560354	HIB19
Live/Dead Aqua	Invitrogen	L34957	
Panel 2			
IgM FITC	BD	BD 555782	G20-127
CD38 APC/AF700	Beckman Coulter	B23489	LS198.4.3
CD3 PE/Texas Red	Beckman Coulter	IM 27050	UCHT1
CD95 PerCP/Cy5.5	BD	BD 561655	DX2
IgD PE Cy7	BD	BD 561314	IA6-2
CD27 APC	Beckman Coulter	B09983	1A4CD27
TACI PE	BD	BD 558414	1A1-K21-M22
CD43 APC.AF750	Beckman Coulter	A89307	DFT1
CD20 Pacific Blue	BioLegend	302320	2H7
Live/Dead Aqua	Invitrogen	L34957	
Sorting Panel			
CD19 PE Cy7	BD	341103	SJ25C1
CD27 APC	Beckman Coulter	B09983	1A4CD27
IgD PE	BD	555779	IA6-2
CD38 PerCP/Cy5.5	Beckman Coulter	A70205	LS198.4.3
IgM FITC	BD	555782	G20-127
CD10 APC/AF750	Beckman Coulter	A889310	ALB1
CD43 APC/AF750	Beckman Coulter	A89307	DFT1
CD11b Pacific Blue	BD	558123	ICRF44
CD56 V450	BD	560361	B159
CD3 Pacific Blue	BD	558117	UCHT1

Supplementary Table 1: Antibodies used for flow cytometry and sorting

CD27-	Cells: L	C vs CB								
ID	Cells	Reads	Productive	Mature(<4	Mutated	Mature	Uniq	MutatedUniq	Mutated	Mutated
			Sequences	Mutations)	B	CDR3V	Dlc	CDR3VDJ <sup>D</sup>	Clone 0.85 <sup>D</sup>	D20 <sup>E</sup>
CB1	68241	235401	117902	89770	28132	27988		13167	12007	358
CB2	25340	213666	97555	68321	29234	13896		8782	6836	144
СВЗ	19945	9 212373	149315	135193	14122	67937		12227	12037	993
CB4	25214	117312	53449	45866	7583	11572		4279	4003	101
CB5	72850	212545	142107	128857	13250	50672		10968	10713	671
CB6	38642	228491	130188	118281	11907	32859		8767	8473	361
CB7	52159	1220401	62587	46666	15921	18555		6993	6191	3
CB8	10641	201017	97002	77115	19887	12205		5885	4750	7
	40924	122402	92197	64020	17259	20455		9911	7007	245
CD3	40024	1111002	02107	60100	6760	20400		5442	5206	245
	154420	111993	00070	80460	6919	21000		5001	5300	470
	04420	621080	0/2/0	00400	46644	29010		20102	20054	4/0
0012	84132	031989	342526	295882	40044	80993		32183	29954	0007
CB13	37832	414721	216548	197236	19312	41666		13205	12606	2327
CB14	48213	503570	259373	227004	32369	56579		20698	19302	289
LC1	74922	<b>)</b> 153828	106727	97255	9472	54277		7692	7547	432
LC2	30168	6 468192	314531	266314	48217	156396		37309	34709	1386
LC3	1E+06	201513	90635	81439	9196	53368		7713	7510	547
LC4	63102	B 198173	151369	142799	8570	98195		7970	7902	994
105	25190	373421	232126	203841	28285	91367		23165	22078	1201
106	1E+06	241042	132564	116593	15971	39382		9965	9365	255
107	170921	3 578007	377840	31596/	61885	117536		39431	35702	831
1.02	1502	302044	228547	212554	15002	08751		14078	13840	1081
	62400	007050	147064	100000	10990	20105		14070	10194	1001
1040	45.00	227303	147204	120990	20200	39105		11234	10164	120
CD27-	Cells: P	338344	256174	227263	28911	111551		22090	21101	1055
	Colle	Ponds	Broductivo	Maturo (<4	Mutated	Moturo	Unio	MutatedUpia	Mutated	Mutated
	Cells	Reaus	Sequences	Mutations)	B	CDR3V	DJc	CDR3VDJ <sup>C</sup>	Clone 0.85 <sup>D</sup>	D20 <sup>E</sup>
Pro1	28034	7 415267	304941	273387	31554	155363		28282	27335	2327
Pro2	30767	3 515651	382670	347876	3/79/	82314		32132	31125	3114
Dro2	129170	264022	122070	106009	21906	60231		20260	19009	790
Pro/	74266	314445	182630	121227	61403	34481		20300	18482	243
Pro5	27675	7 265427	244024	215/91	28542	112492		22031	22422	1295
ries	13/0/3	1505427	244024	215401	20343	1113402		23313	22433	1303
Post1	12684	5 282410	177495	151500	25995	57205		17762	16752	615
Post2	80067	490998	305209	274153	31056	63601		21622	20329	678
Post3	60896	306749	128338	83786	44552	17201		14666	11797	257
Post4	14703	314865	180288	89573	90715	13614		15622	8482	132
Post5	17074	5 328001	202810	136701	66109	51599		33592	29139	972
Plasm	ablasts:	LC vs. C	B					7		
			Productive	UniqCDR3	Clone					
ID	Cells	Reads	Sequences	VDJ <sup>C</sup>	0.85 <sup>D</sup>	D20 <sup>E</sup>	D50 <sup>E</sup>			
LC1	1896	24072	1510	161	122	2		11		
LC2	2382	193157	11057	896	561	12		64		
LC3	27311	1010530	58286	7304	5484	82	4	84		
1.04	411	5278	250	1034	Q7	1		3		
105	12122	90575	Z3Z	90	270					
1010	7065	440622	25070	2501	4640	24	1	70		
LC11	1915	687147	36609	2005	522	9		46		
		0.50								
CB1	638	879130	46321	2494	608	11		50		
CB2	1037	834462	45948	2858	738	8		48		
CB3	1957	322060	17137	1384	714	14		70		
CB4	5293	932475	46025	3836	2046	23	1	76		
CB5	9206	746076	38484	4098	2510	6	1	26		
CB6	2244	840141	43912	2558	915	1		10		
CB7	630	490265	26195	1362	309	4	İ	20		
CB8	4856	141356	7584	437	146	3		13		
CB10	1066	796269	43433	2279	609	7		36		
CB10	899	858262	47210	2632	600	5		26		
CB11	914	408268	22001	15/2	605	7		40		
CB13	261	733574	42020	1710	267	<u>'</u>				
CB14	46610	1130275	42020	1/10	6700	10	2	32		
VD14	-+0010	11303/3	1 01038	1 10098	1 0/22	10	, J	V41		

## Supplementary Table 2: Summary of sequences at each stage of analysis per donor

A: Sequences with < 4 nucleotide mutations in the whole VH sequence

B: Sequences with ≥ 4 nucleotide mutations in the whole VH sequence

C: Number of sequences that are 100% identical in VH, DH, JH and CDR3 D: Number of sequences with the same VH, JH CDR3L and  $\geq$  85% identical in the CDR3

E: The number of clonotypes in a size-ordered list that span 20% (or 50%) of the sequences

LC: Mutated/Mature			CB: Mutated/Mature				
VH	p-Value	FDR	VH	p-Value	FDR		
IGHV1 18	0.0039	0.0105	IGHV1 46	0.0004	0.0017		
IGHV1 24	0.0039	0.0105	IGHV1_58	0.0052	0.0141		
IGHV1 3	0.0039	0.0105	IGHV2 26	0.0002	0.0016		
IGHV1 46	0.0039	0.0105	IGHV2 5	0.0002	0.0016		
IGHV1_58	0.0020	0.0105	IGHV3_15	0.0006	0.0024		
IGHV1_69	0.0020	0.0105	IGHV3 20	0.0005	0.0021		
IGHV2_26	0.0195	0.0412	IGHV3_21	0.0004	0.0017		
IGHV2_5	0.0020	0.0105	IGHV3_23	0.0001	0.0012		
IGHV2_70	0.0039	0.0105	IGHV3_48	0.0009	0.0028		
IGHV3_15	0.0039	0.0105	IGHV3_49	0.0001	0.0012		
IGHV3_20	0.0039	0.0105	IGHV3_53	0.0004	0.0017		
IGHV3_21	0.0195	0.0412	IGHV3_64	0.0009	0.0028		
IGHV3_23	0.0020	0.0105	IGHV3_69_1	0.0004	0.0017		
IGHV3_30	0.0020	0.0105	IGHV3_74	0.0001	0.0012		
IGHV3_49	0.0020	0.0105	IGHV3_9	0.0002	0.0016		
IGHV3_53	0.0059	0.0144	IGHV4_30_2	0.0085	0.0219		
IGHV3_64	0.0020	0.0105	IGHV4_30_4	0.0052	0.0141		
IGHV3_69_1	0.0039	0.0105	IGHV4_34	0.0001	0.0012		
IGHV3_73	0.0098	0.0230	IGHV4_39	0.0203	0.0478		
IGHV3_74	0.0020	0.0105	IGHV4_4	0.0023	0.0068		
IGHV3_9	0.0039	0.0105	IGHV4_59	0.0009	0.0028		
IGHV4_28	0.0059	0.0144	IGHV5_10_1	0.0156	0.0384		
IGHV4_39	0.0020	0.0105	IGHV5_51	0.0001	0.0012		
IGHV4_4	0.0195	0.0412	IGHV7_4_1	0.0017	0.0053		
IGHV4_59	0.0039	0.0105					
IGHV4-34	0.0488	0.0823					
IGHV5_51	0.0020	0.0105					
[IGHV7_4_1	0.0039	0.0105		_			
CB: PC/Mat	ture <sup>®</sup>		CB: PC/Mu	tated <sup>B</sup>			
VH	p-Value	FDR	IGHV1_3	0.0034	0.0185		
IGHV1_46	0.0017	0.0154	IGHV1_46	0.0002	0.0044		
IGHV1_69	0.0005	0.0099	IGHV1_8	0.0024	0.0185		
IGHV1_8	0.0007	0.0099	IGHV2_26	0.0105	0.0405		
IGHV3_43	0.0005	0.0099	IGHV2_5	0.0134	0.0453		
IGHV3_7	0.0046	0.0313	IGHV3_11	0.0134	0.0453		
IGHV3_72	0.0007	0.0099	IGHV3_23	0.0012	0.0132		
IGHV3_74	0.0024	0.0188	IGHV3_30	0.0081	0.0363		
IGHV4_34	0.0012	0.0132	IGHV3_43	0.0034	0.0185		
			IGHV3_64	0.0046	0.0228		
			IGHV3_7	0.0105	0.0405		
			IGHV3_72	0.0007	0.0099		
			IGHV4_39	0.0034	0.0185		
			IGHV4_4	0.0002	0.0044		
			IGHV4_59	0.0017	0.0154		
			IGHV5_51	0.0002	0.0044		

## Supplementary Table 3: Statistics for Figure 4<sup>A</sup>

A: only those V genes with a statistically significant p value <0.05 and false discovery rate (FDR) <0.05 are shown

B: no significant differences were seen in lupus controls