

Supplemental Figure 1. Empty nlg targeting to either CD4 or CD8 T cells does not affect T cell cellularity in vivo.

Age and sex matched MRL/*lpr* mice were treated i.v. with either anti-CD4 antibody or anti-CD8 antibody coated-empty-nlg (equivalent to 50 μ g antibody per mouse) for 48 hrs. PBS or empty-nlg were applied to two control groups separately. (a) Flow cytometry analysis of splenic T cells. Data represent the mean \pm SEM (*n* = 4 mice per group). (b) Dot plot graph shows quantitation of the absolute cell numbers of CD4 and CD8 T cells from the spleens of mice subjected to the indicated treatment.



Supplemental Figure 2. Targeted delivery of ATTO590 by antibody-coated nlg.

12 weeks old MRL/*lpr* mice were treated i.v. with either anti-CD4 antibody or anti-CD8 antibody coated-nlg-ATTO590 (a fluorescent dye derived from Rhodamine), and isotype control antibody coated-nlg-ATTO590 was used as control. Mice were euthanized 30 min after nlg administration for analysis, n = 4 mice per group. (a) Confocal microscopic images show the distribution of ATTO590⁺ cells in representative splenic follicles. Original magnification, ×20; Boxed areas in the upper panel are digitally magnified and shown in the bottom panels (IgM : blue, CD3 : green and ATTO590 MFI in different T cell subsets from spleens of mice subjected to the indicated treatment. **P < 0.01, ***P < 0.005 vs. control, a 2-tailed Student's t test.

а



Supplemental Figure 3. Empty nlg targeting to either CD4 or CD8 T cells does not alter production of auto-antibodies in MRL/*lpr* mice. Age matched MRL/*lpr* mice were treated i.v. with either anti-CD4 antibody or anti-CD8 antibody coated empty-nlg (15 μ l nlg-5-Aza/mouse) weekly for 4 weeks starting at 10 weeks of age. Empty-nlg was used in control group. Dot plot graph shows the ELISA analysis of IgG autoantibodies in the serum from mice subjected to the indicated treatment. *n* = 4 mice per group.



b

Supplemental Figure 4. NIg-5-Aza targeting to either CD4 or CD8 T cells dramatically reduces intrarenal CD4, CD8 and DN T cells.

а

MRL/*lpr* mice were treated i.v. with either anti-CD4 antibody coated-nlg-5-Aza (15µl nlg-5-Aza/mouse) or anti-CD8 antibody coated-nlg-5-Aza (15µl nlg-5-Aza/mouse) every ten days for 60 days starting at 12 weeks of age. Free-5-Aza (5µg/mouse) or empty-nlg were applied to two control groups separately. (a) Flow cytometry analysis of intrarenal T cells. Data represent the mean \pm SEM. (b) Dot plot graph shows quantitation of the absolute cell numbers of CD4, CD8 and DN T cells from the kidneys of mice subjected to the indicated treatment. **P* < 0.05, ***P* < 0.01, ****P* < 0.005 vs. control, a 2-tailed Student's *t* test. *n* = 5-6 mice per group for 2 independent experiments.



Supplemental Figure 5. NIg-5-Aza targeting to either CD4 or CD8 T cells dramatically reduces both inflammatory Th1 and Th17 cells in the cervical lymph nodes.

MRL/*lpr* mice were treated i.v. with either anti-CD4 antibody coated-nlg-5-Aza (15 μ l nlg-5-Aza/mouse) or anti-CD8 antibody coated-nlg-5-Aza (15 μ l nlg-5-Aza/mouse) or empty-nlg were applied to two control groups separately. Flow cytometry analysis of IL-17 and IFN γ expression in CD4⁺ T cells of spleens from mice subjected to the indicated treatment (gated in CD3⁺CD4⁺TCR β ⁺CD49b⁻). Data represents the mean \pm SEM (***P* < 0.01 vs. Control, a 2-tailed Student's *t* test. *n* = 5-6 mice per group for 2 independent experiments).



Supplemental Figure 6. Specific targeting of nlg-5-Aza in CD4⁺ T cells promotes expansion of Tregs in cervical lymph nodes.

MRL/*lpr* mice were treated i.v. with either anti-CD4 antibody coated-nlg-5-Aza (15µl nlg-5-Aza/mouse) or anti-CD8 antibody coated-nlg-5-Aza (15µl nlg-5-Aza/mouse) or empty-nlg were applied to two control group separately. (a) Flow cytometry quantization of the percentage of Foxp3⁺CD4 T cells (Thy1.2⁺CD4⁺) in cervical lymph nodes from mice subjected to the indicated treatment. Data represent the mean \pm SEM. (b) Dot plot graph shows quantitation of Foxp3⁺CD4 T cells (Thy1.2⁺CD4⁺) in cervical lymph nodes from mice subjected to the indicated treatment. ****P* < 0.005 vs. control, a 2-tailed Student's *t* test. *n* = 5-6 mice per group for 2 independent experiments.



Empty-nlg 🔷 5-Aza 🔺 anti-CD4-nlg ● anti-CD8-nlg

Supplemental Figure 7. NIg-5-Aza targeting to CD4 T cells but not free 5-Aza dramatically reduces Foxp3-associated DNA methylation in CD4 T cells.

MRL/*lpr* mice were treated i.v. with either anti-CD4 antibody or anti-CD8 antibody coated-nlg-5-Aza (15µl nlg-5-Aza/mouse) for 10 hrs, free-5-Aza (5µg/mouse) or Empty-nlg were applied to two control groups separately. n = 4 mice per group. Dot plot graph shows quantitation of DNA methylation on indicated gene promoters or enhancers in CD4 T cells sorted from mouse spleens 10 hrs after indicated treatment. *P < 0.05, ***P < 0.005 vs. control, a 2-tailed Student's *t* test.



Supplemental Figure 8. 5-Aza or nlg-5-Aza promotes expansion of Tregs under polarization conditions in vitro.

Naive CD4 T-cells from healthy donors were polarized under Treg inducing conditions for 7 days, with Aza (1 μ M), empty nlg or 5-Aza-nlg (equivalent to 1 μ M) added 12 hrs before collection. (a) Flow cytometry analysis shows the induction of Foxp3 in CD4 T cells polarized *in vitro*. Data represent the mean \pm SEM. (b) Dot plot graph shows Mean fluorescence intensity (MFI) of Foxp3 expression in CD4 T cells polarized *in vitro*. *n*= 4 per group. ***P* < 0.01, ****P* < 0.05 vs. control, a 2-tailed Student's *t* test.

а



Supplemental Figure 9. Specific targeting of nlg-5-Aza to CD8⁺ T cells did not induce Foxp3 expression in CD8 T cells.

MRL/*lpr* mice were treated i.v. with either anti-CD4 antibody coated-nlg-5-Aza (15µl nlg-5-Aza/mouse) or anti-CD8 antibody coated-nlg-5-Aza (15µl nlg-5-Aza/mouse) or empty-nlg were applied to two control group separately. Flow cytometry quantization of the percentage of Foxp3⁺ CD8 T cells (Thy1.2⁺CD8⁺) in spleens from mice subjected to the indicated treatment. Data represent the mean \pm SEM (*n* = 5-6 mice per group for 2 independent experiments).



Supplemental Figure 10. Specific targeting of nlg-5-Aza to CD8⁺ T cells significantly reduces DN T cells in cervical lymph nodes.

MRL/*lpr* mice were treated i.v. with anti-CD8 antibody coated-nlg-5-Aza (15µl nlg-5-Aza/mouse) every ten days for total 60 days total starting at 12 weeks of age. Free-5-Aza (5µg/mouse) or empty-nlg were applied to two control group separately. (a) Flow cytometry quantization of the percentage of Thy1.2+TCR β +TCR β -CD49b-CD4-CD8-T cells in cervical lymph nodes from mice subjected to the indicated treatment. Data represent the mean ± SEM. (b) Dot plots show quantitation of the absolute cell numbers of Thy1.2+TCR β +TCR γ δ -CD49b-CD4-CD8-T cells in cervical lymph nodes from mice subjected to the indicated treatment. ***P* < 0.01, ****P* < 0.005 vs. control, a 2-tailed Student's *t* test. *n* = 5-6 mice per group for 2 independent experiments.

а



Empty-nlg 🔷 5-Aza 🔺 anti-CD4-nlg 🔵 anti-CD8-nlg

Supplemental Figure 11. NIg-5-Aza targeting to CD8 T cells but not free 5-Aza dramatically reduces cytolytic activity associated DNA methylation in CD8 T cells.

MRL/*lpr* mice were treated i.v. with either anti-CD4 antibody or anti-CD8 antibody coated-nlg-5-Aza (15µl nlg-5-Aza/mouse) for 10 hrs, free-5-Aza (5µg/mouse) or Empty-nlg were applied to two control groups separately. n = 4 mice per group. Dot plots show quantitation of DNA methylation on indicated gene promoters or enhancers in CD8 T cells sorted from mouse spleens 10 hrs after indicated treatment. *P < 0.05, **P < 0.01, ***P < 0.005 vs. control, a 2-tailed Student's *t* test.



Empty-nlg 🔷 5-Aza 🔺 anti-CD4-nlg 🔵 anti-CD8-nlg

Supplemental Figure 12. Free-5-Aza but not NIg-5-Aza targeting to either CD4 or CD8 T cells dramatically reduces Inflammation-associated DNA methylation in macrophages and B cells.

Age matched MRL/*lpr* mice were treated i.v. with either anti-CD4 antibody or anti-CD8 antibody coated-nlg-5-Aza (15µl nlg-5-Aza/mouse) for 10 hrs, free-5-Aza (5µg/mouse) or Empty-nlg were applied to two control groups separately. n = 4 mice per group. Dot plot graph shows show quantitation of *DNA methylation* on indicated gene promoters or enhancers in macrophages (M ϕ ,CD11b⁺) or B cells (CD19⁺) sorted from MRL/*lpr* mouse spleens 10 hrs after indicated treatment. **P* < 0.05, ***P* < 0.01, ****P* < 0.005 vs. control, a 2-tailed Student's *t* test.



Supplemental Figure 13. 5-Aza did not induce perforin expression in naïve CD8 T cells but could enhance perforin expression in activated CD8 T cells .

Flow cytometry analysis of intracellular perforin expression in CD45.1⁺OT-I TCR Tg T cells from CD45.1 OT-I TCR Tg Rag1^{-/-} B6 mice (CD45.1⁺TCR β^+ gated) co-cultured with or without OVA₂₅₇₋₂₆₄ loaded antigen-presenting cells (*APC*) for 12 hrs in the absence or presence of 5-Aza (1 μ M). Data represent the mean \pm SEM (*n* = 3-4 per group. **P* < 0.05, ****P* < 0.005 vs. control, a 2-tailed Student's *t* test).



Supplemental Figure 14. 5-Aza or nlg-5-Aza reduced activation induced CD8 downregulation in vitro.

MACS enriched peripheral CD8 T cells from healthy donors were cultured with anti-CD3/anti-CD28 stimulus for 12 hrs with 5-Aza (1 μ M), emptynlg or nlg-5-Azd (equivalent to 1 μ M). *n* = 4 per group. (a) Flow cytometry analysis shows the activation induced CD8 downregulation on cell surface. Data represent the mean \pm SEM. (b) Dot plots graph shows MFI of CD8 expression after stimulation. **P* < 0.05, ***P* < 0.01 vs. indicated control, a 2-tailed Student's *t* test.